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(54) Title: MAIZE CELLULOSE SYNTHASES AND USES THEREOF

(57) Abstract

The invention provides isolated cellulose synthase nucleic acids and their encoded proteins. The present invention provides methods and compositions relating to altering cellulose synthase concentration and/or composition of plants. The invention further provides recombinant expression cassettes, host cells, and transgenic plants.

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Maize Cellulose Synthases and Uses Thereof

TECHNICAL FIELD

The present invention relates generally to plant molecular biology. More specifically, it relates to nucleic acids and methods for modulating their expression in plants.

BACKGROUND OF THE INVENTION

Polysaccharides constitute the bulk of the plant cell walls and have been
traditionally classified into three categories: cellulose, hemicellulose, and pectin. Fry, S.
C. (1988), The growing plant cell wall: Chemical and metabolic analysis. New York:
Longman Scientific & Technical. Whereas cellulose is made at the plasma membrane and directly laid down into the cell wall, hemicellulosic and pectic polymers are first made in the Golgi apparatus and then exported to the cell wall by exocytosis. Ray, P.
M., et al., (1976), Ber. Deutsch. Bot. Ges. Bd. 89, 121-146. The variety of chemical linkages in the pectic and hemicellulosic polysaccharides indicates that there must be tens of polysaccharide synthases in the Golgi apparatus. Darvill et al., (1980). The primary cell walls of flowering plants. In The Plant Cell (N. E. Tolbert, ed.), Vol. 1 in Series:
The biochemistry of plants: A comprehensive treatise, eds. P.K. Stumpf and E.E. Conn
(New York: Academic Press), pp. 91-162.

Cellulose, by virtue of its ability to form semicrystalline microfibrils, has a very high tensile strength which approaches that of some metals. Niklas, K. J. (1992). Plant Biomechanics: An engineering approach to plant form and function, The University of Chicago Press, pp. 607. Bending strength of the culm of normal and brittle-culm mutants of barley has been found to be directly correlated with the concentration of cellulose in the cell wall. Kokubo, et al., (1989), Plant Physiology 91, 876-882; Kokubo, et al., (1991) Plant Physiology 97, 509-514.

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Even though sugar and polysaccharide compositions of the plant cell walls have been well characterized, very limited progress has been made toward identification of the enzymes involved in polysaccharides formation, the reason being their labile nature and recalcitrance to solubilization by available detergents. Sporadic claims for the identification of cellulose synthase from plant sources have been made over the years. Callaghan, T., and Benziman, M. (1984), Nature 311, 165-167; Okuda, et al., (1993),

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Plant Physiol. 101, 1131-1142. However, these claims have been met with skepticism. Callaghan, T., and Benziman, M. (1985), *Nature* 314, 383-384; Delmer, *et al.*, (1993), Plant Physiol. 103, 307-308. It was only recently that a putative gene for plant cellulose synthase (CelA) was cloned from the developing cotton fibers based on homology to the bacterial gene. Pear, *et al.*, *Proc. Natl. Acad. Sci.* (USA) 93, 12637-12642; Saxena, *et al.*, (1990), *Plant Molecular Biology* 15, 673-684; see also, WO 9818949.

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As brittle snap is a major problem in corn breeding, what is needed in the art are compositions and methods for manipulating cellulose concentration in the cell wall and thereby altering plant stalk quality for improved standability or silage. The present invention provides these and other advantages.

SUMMARY OF THE INVENTION

Generally, it is the object of the present invention to provide nucleic acids and proteins relating to cellulose synthases. It is an object of the present invention to provide: 1) nucleic acids and proteins relating to maize cellulose synthases; 2) transgenic plants comprising the nucleic acids of the present invention; 3) methods for modulating, in a transgenic plant, the expression of the nucleic acids of the present invention.

Therefore, in one aspect, the present invention relates to an isolated nucleic acid comprising a member selected from the group consisting of (a) a polynucleotide having a specified sequence identity to a polynucleotide encoding a polypeptide of the present invention;; (b) a polynucleotide which is complementary to the polynucleotide of (a); and (c) a polynucleotide comprising a specified number of contiguous nucleotides from a polynucleotide of (a) or (b). The isolated nucleic acid can be DNA or RNA.

In another aspect, the present invention relates to recombinant expression cassettes, comprising a nucleic acid of the present invention operably linked to a promoter. In some embodiments, the nucleic acid is operably linked in antisense orientation to the promoter.

In another aspect, the present invention is directed to a host cell transfected with the recombinant expression cassette.

In a further aspect, the present invention relates to an isolated protein comprising a polypeptide having a specified number of contiguous amino acids encoded by an isolated nucleic acid of the present invention.

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In another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide of specified length which selectively hybridizes under stringent conditions to a polynucleotide of the present invention, or a complement thereof. In some embodiments, the isolated nucleic acid is operably linked to a promoter.

In yet another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide, the polynucleotide having a specified sequence identity to an identical length of a nucleic acid of the present invention or a complement thereof.

In another aspect, the present invention relates to an isolated nucleic acid comprising a polynucleotide having a sequence of a nucleic acid amplified from a Zea mays nucleic acid library using at least two primers or their complements, one of which selectively hyridizes under stringent conditions to a locus of the nucleic acid comprising the 5' terminal coding region and the other primer selectively hybridizing, under stringent conditions, to a locus of the nucleic acid comprising the 3' terminal coding region, and wherein both primers selectively hybridize within the coding region. In some embodiments, the nucleic acid library is a cDNA library.

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In another aspect, the present invention relates to a recombinant expression cassette comprising a nucleic acid, wherein the nucleic acid is operably linked to a promoter. In some embodiments, the present invention relates to a host cell transfected with this recombinant expression cassette. In some embodiments, the present invention relates to a protein of the present invention which is produced from this host cell.

In a further aspect, the present invention relates to a heterologous promoter operably linked to a non-isolated polynucleotide of the present invention, wherein the polypeptide is encoded by a nucleic acid amplified from a nucleic acid library.

In yet another aspect, the present invention relates to a transgenic plant comprising a recombinant expression cassette comprising a plant promoter operably linked to any of the isolated nucleic acids of the present invention. In some embodiments, the transgenic plant is *Zea mays*. The present invention also provides transgenic seed from the transgenic plant.

In a further aspect, the present invention relates to a method of modulating expression of the genes encoding the proteins of the present invention in a plant cell capable of plant regeneration, comprising the steps of (a) transforming a plant cell with a recombinant expression cassette comprising a polynucleotide of the present invention

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operably linked to a promoter; (b) growing the plant cell under plant growing conditions; and (c) inducing expression of the polynucleotide for a time sufficient to modulate expression of the genes in the plant. In some embodiments, the plant is maize. Expression of the genes encoding the proteins of the present invention can be increased or decreased relative to a non-transformed control plant.

Definitions

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Units, prefixes, and symbols may be denoted in their SI accepted form. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. Numeric ranges are inclusive of the numbers defining the range and include each integer within the defined range. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes. Unless otherwise provided for, software, electrical, and electronics terms as used herein are as defined in The New IEEE Standard Dictinary of Electrical and electronics Terms (5th edition, 1993). The terms defined below are more fully defined by reference to the specification as a whole.

By "amplified" is meant the construction of multiple copies of a nucleic acid sequence or multiple copies complementary to the nucleic acid sequence using at least one of the nucleic acid sequences as a template. Amplification systems include the polymerase chain reaction (PCR) system, ligase chain reaction (LCR) system, nucleic acid sequence based amplification (NASBA, Cangene, Mississauga, Ontario), *Q-Beta* Replicase systems, transcription-based amplification system (TAS), and strand displacement amplification (SDA). See, e.g., *Diagnostic Molecular Microbiology: Principles and Applications*, D. H. Persing *et al.*, Ed., American Society for Microbiology, Washington, D.C. (1993). The product of amplification is termed an amplicon.

The term "antibody" includes reference to antigen binding forms of antibodies (e.g., Fab, F(ab)₂). The term "antibody" frequently refers to a polypeptide substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments thereof which specifically bind and recognize an analyte (antigen). However, while various antibody fragments can be defined in terms of the digestion of an intact antibody, one of

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skill will appreciate that such fragments may be synthesized *de novo* either chemically or by utilizing recombinant DNA methodology. Thus, the term antibody, as used herein, also includes antibody fragments such as single chain Fv, chimeric antibodies (i.e., comprising constant and variable regions from different species), humanized antibodies

(i.e., comprising a complementarity determining region (CDR) from a non-human source) and heteroconjugate antibodies (e.g., bispecific antibodies).

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(1996).

The term "antigen" includes reference to a substance to which an antibody can be generated and/or to which the antibody is specifically immunoreactive. The specific immunoreactive sites within the antigen are known as epitopes or antigenic determinants. These epitopes can be a linear array of monomers in a polymeric composition - such as amino acids in a protein - or consist of or comprise a more complex secondary or tertiary structure. Those of skill will recognize that all immunogens (i.e., substances capable of eliciting an immune response) are antigens; however some antigens, such as haptens, are not immunogens but may be made immunogenic by coupling to a carrier molecule. An antibody immunologically reactive with a particular antigen can be generated *in vivo* or by recombinant methods such as selection of libraries of recombinant antibodies in phage or similar vectors. See, e.g., Huse et al., Science 246: 1275-1281 (1989); and Ward, et al., Nature 341: 544-546 (1989); and Vaughan et al., Nature Biotech. 14: 309-314

As used herein, "antisense orientation" includes reference to a duplex polynucleotide sequence which is operably linked to a promoter in an orientation where the antisense strand is transcribed. The antisense strand is sufficiently complementary to an endogenous transcription product such that translation of the endogenous transcription product is often inhibited.

As used herein, "chromosomal region" includes reference to a length of a chromosome which may be measured by reference to the linear segment of DNA which it comprises. The chromosomal region can be defined by reference to two unique DNA sequences, i.e., markers.

The term "conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or conservatively modified variants of the amino acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein.

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For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations" and represent one species of conservatively modified variation. Every nucleic acid sequence herein which encodes a polypeptide also describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine; and UGG, which is ordinarily the only codon for for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid which encodes a polypeptide of the present invention is implicit in each described polypeptide sequence and incorporated herein by reference.

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As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" where the alteration results in the substitution of an amino acid with a chemically similar amino acid. Thus, any number of amino acid residues selected from the group of integers consisting of from 1 to 15 can be so altered. Thus, for example, 1, 2, 3, 4, 5, 7, or 10 alterations can be made. Conservatively modified variants typically provide similar biological activity as the unmodified polypeptide sequence from which they are derived. For example, substrate specificity, enzyme activity, or ligand/receptor binding is generally at least 30%, 40%, 50%, 60%, 70%, 80%, or 90% of the native protein for it's native substrate. Conservative substitution tables providing functionally similar amino acids are well known in the art.

The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- 2) Aspartic acid (D), Glutamic acid (E);
- 30 3) Asparagine (N), Glutamine (Q);
 - 4) Arginine (R), Lysine (K);
 - 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
 - 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

See also, Creighton (1984) Proteins W.H. Freeman and Company.

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By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as are present in some plant, animal, and fungal mitochondria, the bacterium Mycoplasma capricolum (Proc. Natl. Acad. Sci. (USA), 82: 2306-2309 (1985)), or the ciliate Macronucleus, may be used when the nucleic acid is expressed using these organisms.

When the nucleic acid is prepared or altered synthetically, advantage can be taken of known codon preferences of the intended host where the nucleic acid is to be expressed. For example, although nucleic acid sequences of the present invention may be expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledons or dicotyledons as these preferences have been shown to differ (Murray et al. Nucl. Acids Res. 17: 477-498 (1989)). Thus, the maize preferred codon for a particular amino acid may be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants are listed in Table 4 of Murray et al., above.

As used herein "full-length sequence" in reference to a specified polynucleotide or its encoded protein means having the entire amino acid sequence of, a native (non-synthetic), endogenous, catalytically active form of the specified protein. Methods to determine whether a sequence is full-length are well known in the art including such exemplary techniques as northern or western blots, primer extension, S1 protection, and ribonuclease protection. See, e.g., *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997). Comparison to known full-length homologous (orthologous and/or paralogous) sequences can also be used to identify full-length sequences of the present invention. Additionally, consensus sequences typically present at the 5' and 3' untranslated regions of mRNA aid in the identification of a polynucleotide as full-length. For example, the consensus sequence ANNNNAUGG,

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where the underlined codon represents the N-terminal methionine, aids in determining whether the polynucleotide has a complete 5' end. Consensus sequences at the 3' end, such as polyadenylation sequences, aid in determining whether the polynucleotide has a complete 3' end.

The term "gene activity" refers to one or more steps involved in gene expression, including transcription, translation, and the functioning of the protein encoded by the gene.

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As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species or, if from the same species, is substantially modified from its original form by deliberate human intervention.

By "host cell" is meant a cell which contains a vector and supports the replication and/or expression of the expression vector. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. Preferably, host cells are monocotyledonous or dicotyledonous plant cells. A particularly preferred monocotyledonous host cell is a maize host cell.

The term "hybridization complex" includes reference to a duplex nucleic acid structure formed by two single-stranded nucleic acid sequences selectively hybridized with each other.

By "immunologically reactive conditions" or "immunoreactive conditions" is meant conditions which allow an antibody, generated to a particular epitope, to bind to that epitope to a detectably greater degree (e.g., at least 2-fold over background) than the antibody binds to substantially all other epitopes in a reaction mixture comprising the particular epitope. Immunologically reactive conditions are dependent upon the format of the antibody binding reaction and typically are those utilized in immunoassay protocols. See Harlow and Lane, *Antibodies, A Laboratory Manual*, Cold Spring Harbor Publications, New York (1988), for a description of immunoassay formats and conditions.

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The term "introduced" in the context of inserting a nucleic acid into a cell, means "transfection" or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA).

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The terms "isolated" refers to material, such as a nucleic acid or a protein, which is: (1) substantially or essentially free from components which normally accompany or interact with it as found in its naturally occurring environment. The isolated material optionally comprises material not found with the material in its natural environment; or (2) if the material is in its natural environment, the material has been synthetically (nonnaturally) altered by deliberate human intervention to a composition and/or placed at a locus in the cell (e.g., genome or subcellular organelle) not native to a material found in that environment. The alteration to yield the synthetic material can be performed on the material within or removed from its natural state. For example, a naturally occurring nucleic acid becomes an isolated nucleic acid if it is altered, or if it is transcribed from DNA which has been altered, by non-natural, synthetic (i.e., "man-made") methods performed within the cell from which it originates. See, e.g., Compounds and Methods for Site Directed Mutagenesis in Eukaryotic Cells, Kmiec, U.S. Patent No. 5,565,350; In Vivo Homologous Sequence Targeting in Eukaryotic Cells; Zarling et al., PCT/US93/03868. Likewise, a naturally occurring nucleic acid (e.g., a promoter) becomes isolated if it is introduced by non-naturally occurring means to a locus of the genome not native to that nucleic acid. Nucleic acids which are "isolated" as defined herein, are also referred to as "heterologous" nucleic acids.

Unless otherwise stated, the term "cellulose synthase nucleic acid" is a nucleic acid of the present invention and means a nucleic acid comprising a polynucleotide of the present invention (a "cellulose synthase polynucleotide") encoding a cellulose synthase polypeptide. A "cellulose synthase gene" is a gene of the present invention and refers to a non-heterologous genomic form of a full-length cellulose synthase polynucleotide.

As used herein, "localized within the chromosomal region defined by and including" with respect to particular markers includes reference to a contiguous length of a chromosome delimited by and including the stated markers.

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As used herein, "marker" includes reference to a locus on a chromosome that serves to identify a unique position on the chromosome. A "polymorphic marker" includes reference to a marker which appears in multiple forms (alleles) such that different forms of the marker, when they are present in a homologous pair, allow transmission of each of the chromosomes in that pair to be followed. A genotype may be defined by use of one or a plurality of markers.

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As used herein, "nucleic acid" includes reference to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues having the essential nature of natural nucleotides in that they hybridize to single-stranded nucleic acids in a manner similar to naturally occurring nucleotides (e.g., peptide nucleic acids).

By "nucleic acid library" is meant a collection of isolated DNA or RNA molecules which comprise and substantially represent the entire transcribed fraction of a genome of a specified organism. Construction of exemplary nucleic acid libraries, such as genomic and cDNA libraries, is taught in standard molecular biology references such as Berger and Kimmel, Guide to Molecular Cloning Techniques, Methods in Enzymology, Vol. 152, Academic Press, Inc., San Diego, CA (Berger); Sambrook et al., Molecular Cloning - A Laboratory Manual, 2nd ed., Vol. 1-3 (1989); and Current Protocols in Molecular Biology, F.M. Ausubel et al., Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1994 Supplement).

As used herein "operably linked" includes reference to a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

As used herein, the term "plant" includes reference to whole plants, plant parts or organs (e.g., leaves, stems, roots, etc.), plant cells, seeds and progeny of same. Plant cell, as used herein includes, without limitation, cells obtained from or found in: seeds, suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. Plant cells can also be understood to include modified cells, such as protoplasts, obtained from the aforementioned tissues.

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The class of plants which can be used in the methods of the invention is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants. Particularly preferred plants include maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley and millet.

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As used herein, "polynucleotide" includes reference to a deoxyribopolynucleotide, ribopolynucleotide, or analogs thereof that have the essential nature of a natural ribonucleotide in that they hybridize, under stringent hybridization conditions, to substantially the same nucleotide sequence as naturally occurring nucleotides and/or allow translation into the same amino acid(s) as the naturally occurring nucleotide(s). A polynucleotide can be full-length or a subsequence of a native or heterologous structural or regulatory gene. Unless otherwise indicated, the term includes reference to the specified sequence as well as the complementary sequence thereof. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein. It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well as the chemical forms of DNA and RNA characteristic of viruses and cells, including among other things, simple and complex cells.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers. The essential nature of such analogues of naturally occurring amino acids is that, when incorporated into a protein, that protein is specifically reactive to antibodies elicited to the same protein but consisting entirely of naturally occurring amino acids. The terms "polypeptide", "peptide" and "protein" are also inclusive of modifications including, but not limited to, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation. Exemplary modifications are described in most basic texts, such as, *Proteins - Structure*

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and Molecular Properties, 2nd ed., T. E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as, for example, those provided by Wold, F., Post-translational Protein Modifications: Perspectives and Prospects, pp. 1-12 in Posttranslational Covalent Modification of Proteins, B. C. Johnson, Ed., Academic Press, New York (1983); Seifter et al., Meth. Enzymol. 182: 626-646 (1990) and Rattan et al., Protein Synthesis: Posttranslational Modifications and Aging, Ann. N.Y. Acad. Sci. 663: 48-62 (1992). It will be appreciated, as is well known and as noted above, that polypeptides are not always entirely linear. For instance, polypeptides may be branched as a result of ubiquitination, and they may be circular, with or without branching, generally as a result of posttranslation events, including natural processing event and events brought about by human manipulation which do not occur naturally. Circular, branched and branched circular polypeptides may be synthesized by nontranslation natural process and by entirely synthetic methods, as well. Modifications can occur anywhere in a polypeptide, including the peptide backbone, the amino acid sidechains and the amino or carboxyl termini. In fact, blockage of the amino or carboxyl group in a polypeptide, or both, by a covalent modification, is common in naturally occurring and synthetic polypeptides and such modifications may be present in polypeptides of the present invention, as well. For instance, the amino terminal residue of polypeptides made in E. coli or other cells, prior to proteolytic processing, almost invariably will be N-formylmethionine. During post-translational modification of the peptide, a methionine residue at the NH₂-terminus may be deleted. Accordingly, this invention contemplates the use of both the methionine-containing and the methionine-less amino terminal variants of the protein of the invention. In general, as used herein, the term polypeptide encompasses all such modifications, particularly those that are present in polypeptides synthesized by expressing a polynucleotide in a host cell.

As used herein "promoter" includes reference to a region of DNA upstream from the start of transcription and involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating transcription in plant cells. Exemplary plant promoters include, but are not limited to, those that are obtained from plants, plant viruses, and bacteria which comprise genes expressed in plant cells such Agrobacterium or Rhizobium. Examples of promoters under developmental control include promoters that preferentially initiate transcription in certain tissues, such as leaves, roots, or seeds. Such promoters are

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referred to as "tissue preferred". Promoters which initiate transcription only in certain tissue are referred to as "tissue specific". A "cell type" specific promoter primarily drives expression in certain cell types in one or more organs, for example, vascular cells in roots or leaves. An "inducible" promoter is a promoter which is under environmental control. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions or the presence of light. Tissue specific, tissue preferred, cell type specific, and inducible promoters constitute the class of "non-constitutive" promoters. A "constitutive" promoter is a promoter which is active under most environmental conditions.

The term "cellulose synthase polypeptide" is a polypeptide of the present invention and refers to one or more amino acid sequences, in glycosylated or non-glycosylated form. The term is also inclusive of fragments, variants, homologs, alleles or precursors (e.g., preproproteins or proproteins) thereof. A "cellulose synthase protein" is a protein of the present invention and comprises a cellulose synthase polypeptide.

As used herein "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found in identical form within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under-expressed or not expressed at all as a result of deliberate human intervention. The term "recombinant" as used herein does not encompass the alteration of the cell or vector by naturally occurring events (e.g., spontaneous mutation, natural transformation/transduction/transposition) such as those occurring without deliberate human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements which permit transcription of a particular nucleic acid in a host cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid to be transcribed, and a promoter.

The term "residue" or "amino acid residue" or "amino acid" are used interchangeably herein to refer to an amino acid that is incorporated into a protein,

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polypeptide, or peptide (collectively "protein"). The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass known analogs of natural amino acids that can function in a similar manner as naturally occurring amino acids.

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The term "selectively hybridizes" includes reference to hybridization, under stringent hybridization conditions, of a nucleic acid sequence to a specified nucleic acid target sequence to a detectably greater degree (e.g., at least 2-fold over background) than its hybridization to non-target nucleic acid sequences and to the substantial exclusion of non-target nucleic acids. Selectively hybridizing sequences typically have about at least 80% sequence identity, preferably 90% sequence identity, and most preferably 100% sequence identity (i.e., complementary) with each other.

The term "specifically reactive", includes reference to a binding reaction between an antibody and a protein having an epitope recognized by the antigen binding site of the antibody. This binding reaction is determinative of the presence of a protein having the recognized epitope amongst the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind to an analyte having the recognized epitope to a substantially greater degree (e.g., at least 2-fold over background) than to substantially all other analytes lacking the epitope which are present in the sample.

The terms "stringent conditions" or "stringent hybridization conditions" includes reference to conditions under which a probe will hybridize to its target sequence, to a detectably greater degree than other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which are 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50

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nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C.

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Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl, Anal. Biochem., 138:267-284 (1984): $T_m = 81.5 \, ^{\circ}\text{C} + 16.6 \, (\log M) + 0.41 \, (\% GC) - 10.41 \, (\% GC) + 10.41 \, (\% GC)$ 0.61 (% form) - 500/L; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1 °C for each 1% of mismatching; thus, T_m, hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10 °C. Generally, stringent conditions are selected to be about 5 °C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4 °C lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10 °C lower than the thermal melting point (T_m) ; low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20 °C lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45 °C (aqueous solution) or 32 °C (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive

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guide to the hybridization of nucleic acids is found in Tijssen, Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes, Part I, Chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays", Elsevier, New York (1993); and Current Protocols in Molecular Biology, Chapter 2, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

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As used herein, "transgenic plant" includes reference to a plant which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic. The term "transgenic" as used herein does not encompass the alteration of the genome (chromosomal or extra-chromosomal) by conventional plant breeding methods or by naturally occurring events such as random cross-fertilization, non-recombinant viral infection, non-recombinant bacterial transformation, non-recombinant transposition, or spontaneous mutation.

As used herein, "vector" includes reference to a nucleic acid used in transfection of a host cell and into which can be inserted a polynucleotide. Vectors are often replicons. Expression vectors permit transcription of a nucleic acid inserted therein.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

- (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.
- (b) As used herein, "comparison window" means includes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence may be compared to a reference sequence and wherein the

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portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

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Methods of alignment of sequences for comparison are well-known in the art. 10 Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman, Adv. Appl. Math. 2: 482 (1981); by the homology alignment algorithm of Needleman and Wunsch, J. Mol. Biol. 48: 443 (1970); by the search for similarity method of Pearson and Lipman, Proc. Natl. Acad. Sci. 85: 2444 (1988); by computerized implementations of these algorithms, including, but not limited 15 to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California, GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the CLUSTAL program is well described by Higgins and Sharp, Gene 73: 237-244 (1988); Higgins and Sharp, CABIOS 5: 151-153 (1989); Corpet, et al., Nucleic Acids Research 16: 10881-90 (1988); Huang, et al., Computer Applications in the 20 Biosciences 8: 155-65 (1992), and Pearson, et al., Methods in Molecular Biology 24: 307-331 (1994). The BLAST family of programs which can be used for database similarity searches includes: BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX for nucleotide query sequences against protein database sequences; BLASTP for protein query sequences against protein database sequences; 25 TBLASTN for protein query sequences against nucleotide database sequences; and TBLASTX for nucleotide query sequences against nucleotide database sequences. See, Current Protocols in Molecular Biology, Chapter 19, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using the BLAST 2.0 suite of programs using default parameters. Altschul *et al.*, *Nucleic Acids Res.* 25:3389-3402 (1997). Software for performing BLAST analyses is publicly available, e.g., through the National Center for Biotechnology

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Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., supra). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff (1989) Proc. Natl. Acad. Sci. USA 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul, Proc. Nat'l. Acad. Sci. USA 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance.

BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, *Comput. Chem.*, 17:149-163 (1993)) and

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XNU (Claverie and States, *Comput. Chem.*, 17:191-201 (1993)) low-complexity filters can be employed alone or in combination.

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- (c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences includes reference to the residues in the two sequences which are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and therefore do not change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences which differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well-known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., according to the algorithm of Meyers and Miller, Computer Applic. Biol. Sci., 4: 11-17 (1988) e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).
- (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

(e) (i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity, preferably at least 80%, more preferably at least 90% and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

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Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. However, nucleic acids which do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides which they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is that the polypeptide which the first nucleic acid encodes is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e) (ii) The terms "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70% sequence identity to a reference sequence, preferably 80%, more preferably 85%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970). An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. Peptides which are "substantially similar" share sequences as noted above except that residue positions which are not identical may differ by conservative amino acid changes.

Overview

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The present invention provides, among other things, compositions and methods for modulating (i.e., increasing or decreasing) the level of polypeptides of the present invention in plants. In particular, the polypeptides of the present invention can be expressed at developmental stages, in tissues, and/or in quantities which are uncharacteristic of non-recombinantly engineered plants. Thus, the present invention provides utility in such exemplary applications as improvement of stalk quality for improved stand or silage. Further, the present invention provides for an increased concentration of cellulose in the pericarp; hardening the kernel and thus improving its handling ability.

The present invention also provides isolated nucleic acid comprising polynucleotides of sufficient length and complementarity to a gene of the present invention to use as probes or amplification primers in the detection, quantitation, or isolation of gene transcripts. For example, isolated nucleic acids of the present invention can be used as probes in detecting deficiencies in the level of mRNA in screenings for desired transgenic plants, for detecting mutations in the gene (e.g., substitutions, deletions, or additions), for monitoring upregulation of expression or changes in enzyme activity in screening assays of compounds, for detection of any number of allelic variants (polymorphisms) of the gene, or for use as molecular markers in plant breeding programs. The isolated nucleic acids of the present invention can also be used for recombinant expression of their encoded polypeptides, or for use as immunogens in the preparation and/or screening of antibodies. The isolated nucleic acids of the present invention can also be employed for use in sense or antisense suppression of one or more genes of the present invention in a host cell, tissue, or plant. Attachment of chemical agents which bind, intercalate, cleave and/or crosslink to the isolated nucleic acids of the present invention can also be used to modulate transcription or translation.

The present invention also provides isolated proteins comprising a polypeptide of the present invention (e.g., preproenzyme, proenzyme, or enzymes). The present invention also provides proteins comprising at least one epitope from a polypeptide of the present invention. The proteins of the present invention can be employed in assays for enzyme agonists or antagonists of enzyme function, or for use as immunogens or antigens to obtain antibodies specifically immunoreactive with a protein of the present invention. Such antibodies can be used in assays for expression levels, for identifying

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and/or isolating nucleic acids of the present invention from expression libraries, or for purification of polypeptides of the present invention.

The isolated nucleic acids and proteins of the present invention can be used over a broad range of plant types, particularly monocots such as the species of the Family Graminiae including Sorghum bicolor and Zea mays. The isolated nucleic acid and proteins of the present invention can also be used in species from the genera: Cucurbita, Rosa, Vitis, Juglans, Fragaria, Lotus, Medicago, Onobrychis, Trifolium, Trigonella, Vigna, Citrus, Linum, Geranium, Manihot, Daucus, Arabidopsis, Brassica, Raphanus, Sinapis, Atropa, Capsicum, Datura, Hyoscyamus, Lycopersicon, Nicotiana, Solanum, Petunia, Digitalis, Majorana, Ciahorium, Helianthus, Lactuca, Bromus, Asparagus, Antirrhinum, Heterocallis, Nemesis, Pelargonium, Panieum, Pennisetum, Ranunculus, Senecio, Salpiglossis, Cucumis, Browaalia, Glycine, Pisum, Phaseolus, Lolium, Oryza, Avena, Hordeum, Secale, Triticum, Bambusa, Dendrocalamus, and Melocanna.

15 Nucleic Acids

The present invention provides, *among other things*, isolated nucleic acids of RNA, DNA, and analogs and/or chimeras thereof, comprising a polynucleotide of the present invention.

A polynucleotide of the present invention is inclusive of:

- 20 (a) a polynucleotide encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58, and conservatively modified and polymorphic variants thereof, including exemplary polynucleotides of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57;
- (b) a polynucleotide which is the product of amplification from a Zea mays
 nucleic acid library using primer pairs which selectively hybridize under stringent conditions to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57, wherein the polynucleotide has substantial sequence identity to a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57;
 - (c) a polynucleotide which selectively hybridizes to a polynucleotide of (a) or (b);
 - (d) a polynucleotide having a specified sequence identity with polynucleotides of (a), (b), or (c);

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(e) a polynucleotide encoding a protein having a specified number of contiguous amino acids from a prototype polypeptide, wherein the protein is specifically recognized by antisera elicited by presentation of the protein and wherein the protein does not detectably immunoreact to antisera which has been fully immunosorbed with the protein;

(f) complementary sequences of polynucleotides of (a), (b), (c), (d), or (e); and

(g) a polynucleotide comprising at least a specific number of contiguous nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f).

A. Polynucleotides Encoding A Polypeptide of the Present Invention or Conservatively Modified or Polymorphic Variants Thereof

As indicated in (a), above, the present invention provides isolated nucleic acids comprising a polynucleotide of the present invention, wherein the polynucleotide encodes a polypeptide of the present invention, or conservatively modified or polymorphic variants thereof. Those of skill in the art will recognize that the degeneracy of the genetic code allows for a plurality of polynucleotides to encode for the identical amino acid sequence. Such "silent variations" can be used, for example, to selectively hybridize and detect allelic variants of polynucleotides of the present invention. Accordingly, the present invention includes polynucleotides of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57, and silent variations of polynucleotides encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58. The present invention further provides isolated nucleic acids comprising polynucleotides encoding conservatively modified variants of a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58. Additionally, the present invention further provides isolated nucleic acids comprising polynucleotides encoding one or more polymorphic (allelic) variants of polypeptides/polynucleotides. Polymorphic variants are frequently used to follow segregation of chromosomal regions in, for example, marker assisted selection methods for crop improvement.

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As indicated in (b), above, the present invention provides an isolated nucleic acid comprising a polynucleotide of the present invention, wherein the polynucleotides are amplified from a Zea mays nucleic acid library. Zea mays lines B73, PHRE1, A632, BMS-P2#10, W23, and Mo17 are known and publicly available. Other publicly known and available maize lines can be obtained from the Maize Genetics Cooperation (Urbana, IL). The nucleic acid library may be a cDNA library, a genomic library, or a library generally constructed from nuclear transcripts at any stage of intron processing. cDNA libraries can be normalized to increase the representation of relatively rare cDNAs. In optional embodiments, the cDNA library is constructed using a full-length cDNA synthesis method. Examples of such methods include Oligo-Capping (Maruyama, K. and Sugano, S. Gene 138: 171-174, 1994), Biotinylated CAP Trapper (Carninci, P., Kyan, C., et al. Genomics 37: 327-336, 1996), and CAP Retention Procedure (Edery, E., Chu, L.L., et al. Molecular and Cellular Biology 15: 3363-3371, 1995). cDNA synthesis is often catalyzed at 50-55°C to prevent formation of RNA secondary structure. Examples of reverse transcriptases that are relatively stable at these temperatures are SuperScript II Reverse Transcriptase (Life Technologies, Inc.), AMV Reverse Transcriptase (Boehringer Mannheim) and RetroAmp Reverse Transcriptase (Epicentre). Rapidly growing tissues, or rapidly dividing cells are preferably used as mRNA sources such as from the elongating internode of corn plants.

The polynucleotides of the present invention include those amplified using the following primer pairs:

SEQ ID NOS: 3 and 4 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 1;

SEQ ID NOS: 7 and 8 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 5; and

SEQ ID NOS: 11 and 12 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 9.

SEQ ID NOS: 15 and 16 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 13.

SEQ ID NOS: 19 and 20 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 17;

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SEQ ID NOS: 23 and 24 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 21; and

SEQ ID NOS: 27 and 28 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 25.

SEQ ID NOS: 31 and 32 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 29.

SEQ ID NOS: 35 and 36 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 33;

SEQ ID NOS: 39 and 40 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 37; and

SEQ ID NOS: 43 and 44 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 41.

SEQ ID NOS: 47 and 48 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 45.

SEQ ID NOS: 51 and 52 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 49;

SEQ ID NOS: 55 and 56 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 53; and

SEQ ID NOS: 59 and 60 which yield an amplicon comprising a sequence having substantial identity to SEQ ID NO: 57.

The present invention also provides subsequences of the polynucleotides of the present invention. A variety of subsequences can be obtained using primers which selectively hybridize under stringent conditions to at least two sites within a polynucleotide of the present invention, or to two sites within the nucleic acid which flank and comprise a polynucleotide of the present invention, or to a site within a polynucleotide of the present invention and a site within the nucleic acid which comprises it. Primers are chosen to selectively hybridize, under stringent hybridization conditions, to a polynucleotide of the present invention. Generally, the primers are complementary to a subsequence of the target nucleic acid which they amplify. As those skilled in the art will appreciate, the sites to which the primer pairs will selectively hybridize are chosen such that a single contiguous nucleic acid can be formed under the desired amplification conditions.

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In optional embodiments, the primers will be constructed so that they selectively hybridize under stringent conditions to a sequence (or its complement) within the target nucleic acid which comprises the codon encoding the carboxy or amino terminal amino acid residue (i.e., the 3' terminal coding region and 5' terminal coding region, respectively) of the polynucleotides of the present invention. Optionally within these embodiments, the primers will be constructed to selectively hybridize entirely within the coding region of the target polynucleotide of the present invention such that the product of amplification of a cDNA target will consist of the coding region of that cDNA. The primer length in nucleotides is selected from the group of integers consisting of from at least 15 to 50. Thus, the primers can be at least 15, 18, 20, 25, 30, 40, or 50 nucleotides in length. Those of skill will recognize that a lengthened primer sequence can be employed to increase specificity of binding (i.e., annealing) to a target sequence. A non-annealing sequence at the 5'end of a primer (a "tail") can be added, for example, to introduce a cloning site at the terminal ends of the amplicon.

The amplification products can be translated using expression systems well known to those of skill in the art and as discussed, *infra*. The resulting translation products can be confirmed as polypeptides of the present invention by, for example, assaying for the appropriate catalytic activity (e.g., specific activity and/or substrate specificity), or verifying the presence of one or more linear epitopes which are specific to a polypeptide of the present invention. Methods for protein synthesis from PCR derived templates are known in the art and available commercially. See, e.g., Amersham Life Sciences, Inc, Catalog '97, p.354.

Methods for obtaining 5' and/or 3' ends of a vector insert are well known in the art. See, e.g., RACE (Rapid Amplification of Complementary Ends) as described in Frohman, M. A., in PCR Protocols: A Guide to Methods and Applications, M. A. Innis, D. H. Gelfand, J. J. Sninsky, T. J. White, Eds. (Academic Press, Inc., San Diego, 1990), pp. 28-38.); see also, U.S. Pat. No. 5,470,722, and Current Protocols in Molecular Biology, Unit 15.6, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995); Frohman and Martin, Techniques 1:165 (1989).

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C. Polynucleotides Which Selectively Hybridize to a Polynucleotide of (A) or (B)

As indicated in (c), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides

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selectively hybridize, under selective hybridization conditions, to a polynucleotide of paragraphs (A) or (B) as discussed, above. Thus, the polynucleotides of this embodiment can be used for isolating, detecting, and/or quantifying nucleic acids comprising the polynucleotides of (A) or (B). For example, polynucleotides of the present invention can be used to identify, isolate, or amplify partial or full-length clones in a deposited library. In some embodiments, the polynucleotides are genomic or cDNA sequences isolated or otherwise complementary to a cDNA from a dicot or monocot nucleic acid library. Exemplary species of monocots and dicots include, but are not limited to: corn, canola, soybean, cotton, wheat, sorghum, sunflower, oats, sugar cane, millet, barley, and rice. Optionally, the cDNA library comprises at least 80% full-length sequences, preferably at least 85% or 90% full-length sequences, and more preferably at least 95% full-length sequences. The cDNA libraries can be normalized to increase the representation of rare sequences. Low stringency hybridization conditions are typically, but not exclusively, employed with sequences having a reduced sequence identity relative to complementary sequences. Moderate and high stringency conditions can optionally be employed for sequences of greater identity. Low stringency conditions allow selective hybridization of sequences having about 70% sequence identity and can be employed to identify orthologous or paralogous sequences.

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D. Polynucleotides Having a Specific Sequence Identity with the Polynucleotides of (A), (B) or (C)

As indicated in (d), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides have a specified identity at the nucleotide level to a polynucleotide as disclosed above in paragraphs (A), (B), or (C). The percentage of identity to a reference sequence is at least 60% and, rounded upwards to the nearest integer, can be expressed as an integer selected from the group of integers consisting of from 60 to 99. Thus, for example, the percentage of identity to a reference sequence can be at least 70%, 75%, 80%, 85%, 90%, or 95%.

Optionally, the polynucleotides of this embodiment will share an epitope with a polypeptide encoded by the polynucleotides of (A), (B), or (C). Thus, these polynucleotides encode a first polypeptide which elicits production of antisera comprising antibodies which are specifically reactive to a second polypeptide encoded by

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a polynucleotide of (A), (B), or (C). However, the first polypeptide does not bind to antisera raised against itself when the antisera has been fully immunosorbed with the first polypeptide. Hence, the polynucleotides of this embodiment can be used to generate antibodies for use in, for example, the screening of expression libraries for nucleic acids comprising polynucleotides of (A), (B), or (C), or for purification of, or in immunoassays for, polypeptides encoded by the polynucleotides of (A), (B), or (C). The polynucleotides of this embodiment embrace nucleic acid sequences which can be employed for selective hybridization to a polynucleotide encoding a polypeptide of the present invention.

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Screening polypeptides for specific binding to antisera can be conveniently achieved using peptide display libraries. This method involves the screening of large collections of peptides for individual members having the desired function or structure. Antibody screening of peptide display libraries is well known in the art. The displayed peptide sequences can be from 3 to 5000 or more amino acids in length, frequently from 5-100 amino acids long, and often from about 8 to 15 amino acids long. In addition to direct chemical synthetic methods for generating peptide libraries, several recombinant DNA methods have been described. One type involves the display of a peptide sequence on the surface of a bacteriophage or cell. Each bacteriophage or cell contains the nucleotide sequence encoding the particular displayed peptide sequence. Such methods are described in PCT patent publication Nos. 91/17271, 91/18980, 91/19818, and 93/08278. Other systems for generating libraries of peptides have aspects of both in vitro chemical synthesis and recombinant methods. See, PCT Patent publication Nos. 92/05258, 92/14843, and 96/19256. See also, U.S. Patent Nos. 5,658,754; and 5,643,768. Peptide display libraries, vectors, and screening kits are commercially available from such suppliers as Invitrogen (Carlsbad, CA).

E. Polynucleotides Encoding a Protein Having a Subsequence from a Prototype Polypeptide and is Cross-Reactive to the Prototype Polypeptide

As indicated in (e), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides encode a protein having a subsequence of contiguous amino acids from a prototype polypeptide of the present invention such as are provided in (a), above. The length of contiguous amino acids from the prototype polypeptide is selected from the group of integers

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consisting of from at least 10 to the number of amino acids within the prototype sequence. Thus, for example, the polynucleotide can encode a polypeptide having a subsequence having at least 10, 15, 20, 25, 30, 35, 40, 45, or 50, contiguous amino acids from the prototype polypeptide. Further, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

The proteins encoded by polynucleotides of this embodiment, when presented as an immunogen, elicit the production of polyclonal antibodies which specifically bind to a prototype polypeptide such as but not limited to, a polypeptide encoded by the polynucleotide of (a) or (b), above. Generally, however, a protein encoded by a polynucleotide of this embodiment does not bind to antisera raised against the prototype polypeptide when the antisera has been fully immunosorbed with the prototype polypeptide. Methods of making and assaying for antibody binding specificity/affinity are well known in the art. Exemplary immunoassay formats include ELISA, competitive immunoassays, radioimmunoassays, Western blots, indirect immunofluorescent assays and the like.

In a preferred assay method, fully immunosorbed and pooled antisera which is elicited to the prototype polypeptide can be used in a competitive binding assay to test the protein. The concentration of the prototype polypeptide required to inhibit 50% of the binding of the antisera to the prototype polypeptide is determined. If the amount of the protein required to inhibit binding is less than twice the amount of the prototype protein, then the protein is said to specifically bind to the antisera elicited to the immunogen. Accordingly, the proteins of the present invention embrace allelic variants, conservatively modified variants, and minor recombinant modifications to a prototype polypeptide.

A polynucleotide of the present invention optionally encodes a protein having a molecular weight as the non-glycosylated protein within 20% of the molecular weight of the full-length non-glycosylated polypeptides of the present invention. Molecular weight can be readily determined by SDS-PAGE under reducing conditions. Preferably, the molecular weight is within 15% of a full length polypeptide of the present invention, more preferably within 10% or 5%, and most preferably within 3%, 2%, or 1% of a full

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length polypeptide of the present invention. Molecular weight determination of a protein can be conveniently performed by SDS-PAGE under denaturing conditions.

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Optionally, the polynucleotides of this embodiment will encode a protein having a specific activity at least 50%, 60%, 80%, or 90% of the native, endogenous (i.e., nonisolated), full-length polypeptide of the present invention. Further, the proteins encoded by polynucleotides of this embodiment will optionally have a substantially similar affinity constant (K_m) and/or catalytic activity (i.e., the microscopic rate constant, k_{cat}) as the native endogenous, full-length protein. Those of skill in the art will recognize that k_{cat}/K_m value determines the specificity for competing substrates and is often referred to as the specificity constant. Proteins of this embodiment can have a k_{car}/K_m value at least 10% of a non-isolated full-length polypeptide of the present invention as determined using the endogenous substrate of that polypeptide. Optionally, the k_{car}/K_m value will be at least 20%, 30%, 40%, 50%, and most preferably at least 60%, 70%, 80%, 90%, or 95% the k_{cat}/K_m value of the non-isolated, full-length polypeptide of the present invention. Determination of k_{cat} , K_{m} , and $k_{\text{cat}}/K_{\text{m}}$ can be determined by any number of means well known to those of skill in the art. For example, the initial rates (i.e., the first 5% or less of the reaction) can be determined using rapid mixing and sampling techniques (e.g., continuous-flow, stopped-flow, or rapid quenching techniques), flash photolysis, or relaxation methods (e.g., temperature jumps) in conjunction with such exemplary methods of measuring as spectrophotometry, spectrofluorimetry, nuclear magnetic resonance, or radioactive procedures. Kinetic values are conveniently obtained using a Lineweaver-Burk or Eadie-Hofstee plot.

F. Polynucleotides Complementary to the Polynucleotides of (A)-(E)

As indicated in (f), above, the present invention provides isolated nucleic acids comprising polynucleotides complementary to the polynucleotides of paragraphs A-E, above. As those of skill in the art will recognize, complementary sequences base-pair throughout the entirety of their length with the polynucleotides of (A)-(E) (i.e., have 100% sequence identity over their entire length). Complementary bases associate through hydrogen bonding in double stranded nucleic acids. For example, the following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

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G. Polynucleotides Which are Subsequences of the Polynucleotides of (A)-(F)

As indicated in (g), above, the present invention provides isolated nucleic acids comprising polynucleotides which comprise at least 15 contiguous bases from the polynucleotides of (A) through (F) as discussed above. The length of the polynucleotide is given as an integer selected from the group consisting of from at least 15 to the length of the nucleic acid sequence from which the polynucleotide is a subsequence of. Thus, for example, polynucleotides of the present invention are inclusive of polynucleotides comprising at least 15, 20, 25, 30, 40, 50, 60, 75, or 100 contiguous nucleotides in length from the polynucleotides of (A)-(F). Optionally, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

The subsequences of the present invention can comprise structural characteristics of the sequence from which it is derived. Alternatively, the subsequences can lack certain structural characteristics of the larger sequence from which it is derived. For example, a subsequence from a polynucleotide encoding a polypeptide having at least one linear epitope in common with a prototype polypeptide sequence as provided in (a), above, may encode an epitope in common with the prototype sequence. Alternatively, the subsequence may not encode an epitope in common with the prototype sequence but can be used to isolate the larger sequence by, for example, nucleic acid hybridization with the sequence from which it's derived. Subsequences can be used to modulate or detect gene expression by introducing into the subsequences compounds which bind, intercalate, cleave and/or crosslink to nucleic acids. Exemplary compounds include acridine, psoralen, phenanthroline, naphthoquinone, daunomycin or chloroethylaminoaryl conjugates.

Construction of Nucleic Acids

The isolated nucleic acids of the present invention can be made using (a) standard recombinant methods, (b) synthetic techniques, or combinations thereof. In some embodiments, the polynucleotides of the present invention will be cloned, amplified, or otherwise constructed from a monocot. In preferred embodiments the monocot is Zea mays.

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The nucleic acids may conveniently comprise sequences in addition to a polynucleotide of the present invention. For example, a multi-cloning site comprising one or more endonuclease restriction sites may be inserted into the nucleic acid to aid in isolation of the polynucleotide. Also, translatable sequences may be inserted to aid in the isolation of the translated polynucleotide of the present invention. For example, a hexa-histidine marker sequence provides a convenient means to purify the proteins of the present invention. A polynucleotide of the present invention can be attached to a vector, adapter, or linker for cloning and/or expression of a polynucleotide of the present invention. Additional sequences may be added to such cloning and/or expression sequences to optimize their function in cloning and/or expression, to aid in isolation of the polynucleotide, or to improve the introduction of the polynucleotide into a cell. Typically, the length of a nucleic acid of the present invention less the length of its polynucleotide of the present invention is less than 20 kilobase pairs, often less than 15 kb, and frequently less than 10 kb. Use of cloning vectors, expression vectors, adapters, and linkers is well known and extensivley described in the art. For a description of various nucleic acids see, for example, Stratagene Cloning Systems, Catalogs 1995, 1996, 1997 (La Jolla, CA); and, Amersham Life Sciences, Inc, Catalog '97 (Arlington Heights, IL).

20 A. Recombinant Methods for Constructing Nucleic Acids

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The isolated nucleic acid compositions of this invention, such as RNA, cDNA, genomic DNA, or a hybrid thereof, can be obtained from plant biological sources using any number of cloning methodologies known to those of skill in the art. In some embodiments, oligonucleotide probes which selectively hybridize, under stringent conditions, to the polynucleotides of the present invention are used to identify the desired sequence in a cDNA or genomic DNA library. While isolation of RNA, and construction of cDNA and genomic libraries is well known to those of ordinary skill in the art, the following highlights some of the methods employed.

A1. mRNA Isolation and Purification

Total RNA from plant cells comprises such nucleic acids as mitochondrial RNA, chloroplastic RNA, rRNA, tRNA, hnRNA and mRNA. Total RNA preparation typically involves lysis of cells and removal of proteins, followed by precipitation of nucleic

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acids. Extraction of total RNA from plant cells can be accomplished by a variety of means. Frequently, extraction buffers include a strong detergent such as SDS and an organic denaturant such as guanidinium isothiocyanate, guanidine hydrochloride or phenol. Following total RNA isolation, poly(A)⁺ mRNA is typically purified from the remainder RNA using oligo(dT) cellulose. Exemplary total RNA and mRNA isolation protocols are described in *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); and, *Current Protocols in Molecular Biology*, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995). Total RNA and mRNA isolation kits are commercially available from vendors such as Stratagene (La Jolla, CA), Clonetech (Palo Alto, CA), Pharmacia (Piscataway, NJ), and 5'-3' (Paoli, PA). See also, U.S. Patent Nos. 5,614,391; and, 5,459,253. The mRNA can be fractionated into populations with size ranges of about 0.5, 1.0, 1.5, 2.0, 2.5 or 3.0 kb. The cDNA synthesized for each of these fractions can be size selected to the same size range as its mRNA prior to vector insertion. This method helps eliminate truncated cDNA formed by incompletely reverse transcribed mRNA.

A2. Construction of a cDNA Library

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Construction of a cDNA library generally entails five steps. First, first strand cDNA synthesis is initiated from a poly(A)⁺ mRNA template using a poly(dT) primer or random hexanucleotides. Second, the resultant RNA-DNA hybrid is converted into double stranded cDNA, typically by a combination of RNAse H and DNA polymerase I (or Klenow fragment). Third, the termini of the double stranded cDNA are ligated to adaptors. Ligation of the adaptors will produce cohesive ends for cloning. Fourth, size selection of the double stranded cDNA eliminates excess adaptors and primer fragments, and eliminates partial cDNA molecules due to degradation of mRNAs or the failure of reverse transcriptase to synthesize complete first strands. Fifth, the cDNAs are ligated into cloning vectors and packaged. cDNA synthesis protocols are well known to the skilled artisan and are described in such standard references as: *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); and, *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995). cDNA synthesis kits are available from a variety of commercial vendors such as Stratagene or Pharmacia.

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A number of cDNA synthesis protocols have been described which provide substantially pure full-length cDNA libraries. Substantially pure full-length cDNA libraries are constructed to comprise at least 90%, and more preferably at least 93% or 95% full-length inserts amongst clones containing inserts. The length of insert in such libraries can be from 0 to 8, 9, 10, 11, 12, 13, or more kilobase pairs. Vectors to accommodate inserts of these sizes are known in the art and available commercially. See, e.g., Stratagene's lambda ZAP Express (cDNA cloning vector with 0 to 12 kb cloning capacity).

An exemplary method of constructing a greater than 95% pure full-length cDNA library is described by Carninci et al., Genomics, 37:327-336 (1996). In that protocol, the cap-structure of eukaryotic mRNA is chemically labeled with biotin. By using streptavidin-coated magnetic beads, only the full-length first-strand cDNA/mRNA hybrids are selectively recovered after RNase I treatment. The method provides a high yield library with an unbiased representation of the starting mRNA population. Other methods for producing full-length libraries are known in the art. See, e.g., Edery et al., Mol. Cell Biol., 15(6):3363-3371 (1995); and, PCT Application WO 96/34981.

A3. Normalized or Subtracted cDNA Libraries

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A non-normalized cDNA library represents the mRNA population of the tissue it was made from. Since unique clones are out-numbered by clones derived from highly expressed genes their isolation can be laborious. Normalization of a cDNA library is the process of creating a library in which each clone is more equally represented.

A number of approaches to normalize cDNA libraries are known in the art. One approach is based on hybridization to genomic DNA. The frequency of each hybridized cDNA in the resulting normalized library would be proportional to that of each corresponding gene in the genomic DNA. Another approach is based on kinetics. If cDNA reannealing follows second-order kinetics, rarer species anneal less rapidly and the remaining single-stranded fraction of cDNA becomes progressively more normalized during the course of the hybridization. Specific loss of any species of cDNA, regardless of its abundance, does not occur at any Cot value. Construction of normalized libraries is described in Ko, *Nucl. Acids. Res.*, 18(19):5705-5711 (1990); Patanjali *et al.*, *Proc. Natl. Acad. U.S.A.*, 88:1943-1947 (1991); U.S. Patents 5,482,685, and 5,637,685. In an exemplary method described by Soares *et al.*, normalization resulted in reduction of

the abundance of clones from a range of four orders of magnitude to a narrow range of only 1 order of magnitude. *Proc. Natl. Acad. Sci. USA*, 91:9228-9232 (1994).

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Subtracted cDNA libraries are another means to increase the proportion of less abundant cDNA species. In this procedure, cDNA prepared from one pool of mRNA is depleted of sequences present in a second pool of mRNA by hybridization. The cDNA:mRNA hybrids are removed and the remaining un-hybridized cDNA pool is enriched for sequences unique to that pool. See, Foote et al. in, Plant Molecular Biology: A Laboratory Manual, Clark, Ed., Springer-Verlag, Berlin (1997); Kho and Zarbl, Technique, 3(2):58-63 (1991); Sive and St. John, Nucl. Acids Res., 16(22):10937 (1988); Current Protocols in Molecular Biology, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995); and, Swaroop et al., Nucl. Acids Res., 19)8):1954 (1991). cDNA subtraction kits are commercially available. See, e.g., PCR-Select (Clontech).

15 A4. Construction of a Genomic Library

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To construct genomic libraries, large segments of genomic DNA are generated by random fragmentation, e.g. using restriction endonucleases, and are ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. Methodologies to accomplish these ends, and sequencing methods to verify the sequence of nucleic acids are well known in the art. Examples of appropriate molecular biological 20 techniques and instructions sufficient to direct persons of skill through many construction, cloning, and screening methodologies are found in Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor Laboratory Vols. 1-3 (1989), Methods in Enzymology, Vol. 152: Guide to Molecular Cloning Techniques, Berger and Kimmel, Eds., San Diego: Academic Press, Inc. (1987), Current 25 Protocols in Molecular Biology, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995); Plant Molecular Biology: A Laboratory Manual, Clark, Ed., Springer-Verlag, Berlin (1997). Kits for construction of genomic libraries are also commercially available.

A5. Nucleic Acid Screening and Isolation Methods

The cDNA or genomic library can be screened using a probe based upon the sequence of a polynucleotide of the present invention such as those disclosed herein.

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Probes may be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes in the same or different plant species. Those of skill in the art will appreciate that various degrees of stringency of hybridization can be employed in the assay; and either the hybridization or the wash medium can be stringent. As the conditions for hybridization become more stringent, there must be a greater degree of complementarity between the probe and the target for duplex formation to occur. The degree of stringency can be controlled by temperature, ionic strength, pH and the presence of a partially denaturing solvent such as formamide. For example, the stringency of hybridization is conveniently varied by changing the polarity of the reactant solution through manipulation of the concentration of formamide within the range of 0% to 50%. The degree of complementarity (sequence identity) required for detectable binding will vary in accordance with the stringency of the hybridization medium and/or wash medium. The degree of complementarity will optimally be 100 percent; however, it should be understood that minor sequence variations in the probes and primers may be compensated for by reducing the stringency of the hybridization and/or wash medium.

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The nucleic acids of interest can also be amplified from nucleic acid samples using amplification techniques. For instance, polymerase chain reaction (PCR) technology can be used to amplify the sequences of polynucleotides of the present invention and related genes directly from genomic DNA or cDNA libraries. PCR and other *in vitro* amplification methods may also be useful, for example, to clone nucleic acid sequences that code for proteins to be expressed, to make nucleic acids to use as probes for detecting the presence of the desired mRNA in samples, for nucleic acid sequencing, or for other purposes. Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods are found in Berger, Sambrook, and Ausubel, as well as Mullis *et al.*, U.S. Patent No. 4,683,202 (1987); and, *PCR Protocols A Guide to Methods and Applications*, Innis *et al.*, Eds., Academic Press Inc., San Diego, CA (1990). Commercially available kits for genomic PCR amplification are known in the art. See, e.g., Advantage-GC Genomic PCR Kit (Clontech). The T4 gene 32 protein (Boehringer Mannheim) can be used to improve yield of long PCR products.

PCR-based screening methods have also been described. Wilfinger *et al*. describe a PCR-based method in which the longest cDNA is identified in the first step so that incomplete clones can be eliminated from study. *BioTechniques*, 22(3): 481-486 (1997). In that method, a primer pair is synthesized with one primer annealing to the 5'

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end of the sense strand of the desired cDNA and the other primer to the vector. Clones are pooled to allow large-scale screening. By this procedure, the longest possible clone is identified amongst candidate clones. Further, the PCR product is used solely as a diagnostic for the presence of the desired cDNA and does not utilize the PCR product itself. Such methods are particularly effective in combination with a full-length cDNA construction methodology, above.

B. Synthetic Methods for Constructing Nucleic Acids

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The isolated nucleic acids of the present invention can also be prepared by direct 10 chemical synthesis by methods such as the phosphotriester method of Narang et al., Meth. Enzymol. 68: 90-99 (1979); the phosphodiester method of Brown et al., Meth. Enzymol. 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage et al., Tetra. Lett. 22: 1859-1862 (1981); the solid phase phosphoramidite triester method described by Beaucage and Caruthers, Tetra. Letts. 22(20): 1859-1862 (1981), e.g., 15 using an automated synthesizer, e.g., as described in Needham-VanDevanter et al., Nucleic Acids Res., 12: 6159-6168 (1984); and, the solid support method of U.S. Patent No. 4,458,066. Chemical synthesis generally produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the 20 single strand as a template. One of skill will recognize that while chemical synthesis of DNA is limited to sequences of about 100 bases, longer sequences may be obtained by the ligation of shorter sequences.

Recombinant Expression Cassettes

The present invention further provides recombinant expression cassettes comprising a nucleic acid of the present invention. A nucleic acid sequence coding for the desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a full length polypeptide of the present invention, can be used to construct a recombinant expression cassette which can be introduced into the desired host cell. A recombinant expression cassette will typically comprise a polynucleotide of the present invention operably linked to transcriptional initiation regulatory sequences which will direct the transcription of the polynucleotide in the intended host cell, such as tissues of a transformed plant.

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For example, plant expression vectors may include (1) a cloned plant gene under the transcriptional control of 5' and 3' regulatory sequences and (2) a dominant selectable marker. Such plant expression vectors may also contain, if desired, a promoter regulatory region (e.g., one conferring inducible or constitutive, environmentally- or developmentally-regulated, or cell- or tissue-specific/selective expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site, and/or a polyadenylation signal.

A plant promoter fragment can be employed which will direct expression of a polynucleotide of the present invention in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the 1'- or 2'- promoter derived from T-DNA of Agrobacterium tumefaciens, the ubiquitin 1 promoter, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (U.S. Patent No. 5,683,439), the Nos promoter, the pEmu promoter, the rubisco promoter, the GRP1-8 promoter, the actin promoter, the F3.7 promoter, and other transcription initiation regions from various plant genes known to those of skill.

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Alternatively, the plant promoter can direct expression of a polynucleotide of the present invention in a specific tissue or may be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Environmental conditions that may effect transcription by inducible promoters include pathogen attack, anaerobic conditions, or the presence of light. Examples of inducible promoters are the Adh1 promoter which is inducible by hypoxia or cold stress, the Hsp70 promoter which is inducible by heat stress, and the PPDK promoter which is inducible by light.

Examples of promoters under developmental control include promoters that initiate transcription only, or preferentially, in certain tissues, such as leaves, roots, fruit, seeds, or flowers. The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

Both heterologous and non-heterologous (i.e., endogenous) promoters can be employed to direct expression of the nucleic acids of the present invention. These

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promoters can also be used, for example, in recombinant expression cassettes to drive expression of antisense nucleic acids to reduce, increase, or alter concentration and/or composition of the proteins of the present invention in a desired tissue. Thus, in some embodiments, the nucleic acid construct will comprise a promoter functional in a plant cell, such as in Zea mays, operably linked to a polynucleotide of the present invention. Promoters useful in these embodiments include the endogenous promoters driving expression of a polypeptide of the present invention.

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In some embodiments, isolated nucleic acids which serve as promoter or enhancer elements can be introduced in the appropriate position (generally upstream) of a non-heterologous form of a polynucleotide of the present invention so as to up or down regulate expression of a polynucleotide of the present invention. For example, endogenous promoters can be altered *in vivo* by mutation, deletion, and/or substitution (see, Kmiec, U.S. Patent 5,565,350; Zarling *et al.*, PCT/US93/03868), or isolated promoters can be introduced into a plant cell in the proper orientation and distance from a gene of the present invention so as to control the expression of the gene. Gene expression can be modulated under conditions suitable for plant growth so as to alter the total concentration and/or alter the composition of the polypeptides of the present invention in plant cell. Thus, the present invention provides compositions, and methods for making, heterologous promoters and/or enhancers operably linked to a native, endogenous (i.e., non-heterologous) form of a polynucleotide of the present invention.

Methods for identifying promoters with a particular expression pattern, in terms of, e.g., tissue type, cell type, stage of development, and/or environmental conditions, are well known in the art. See, e.g., *The Maize Handbook*, Chapters 114-115, Freeling and Walbot, Eds., Springer, New York (1994); *Corn and Corn Improvement*, 3rd edition, Chapter 6, Sprague and Dudley, Eds., American Society of Agronomy, Madison, Wisconsin (1988). A typical step in promoter isolation methods is identification of gene products that are expressed with some degree of specificity in the target tissue. Amongst the range of methodologies are: differential hybridization to cDNA libraries; subtractive hybridization; differential display; differential 2-D protein gel electrophoresis; DNA probe arrays; and isolation of proteins known to be expressed with some specificity in the target tissue. Such methods are well known to those of skill in the art. Commercially available products for identifying promoters are known in the art such as Clontech's (Palo Alto, CA) Universal GenomeWalker Kit.

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For the protein-based methods, it is helpful to obtain the amino acid sequence for at least a portion of the identified protein, and then to use the protein sequence as the basis for preparing a nucleic acid that can be used as a probe to identify either genomic DNA directly, or preferably, to identify a cDNA clone from a library prepared from the target tissue. Once such a cDNA clone has been identified, that sequence can be used to identify the sequence at the 5' end of the transcript of the indicated gene. For differential hybridization, subtractive hybridization and differential display, the nucleic acid sequence identified as enriched in the target tissue is used to identify the sequence at the 5' end of the transcript of the indicated gene. Once such sequences are identified, starting either from protein sequences or nucleic acid sequences, any of these sequences identified as being from the gene transcript can be used to screen a genomic library prepared from the target organism. Methods for identifying and confirming the transcriptional start site are well known in the art.

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In the process of isolating promoters expressed under particular environmental conditions or stresses, or in specific tissues, or at particular developmental stages, a number of genes are identified that are expressed under the desired circumstances, in the desired tissue, or at the desired stage. Further analysis will reveal expression of each particular gene in one or more other tissues of the plant. One can identify a promoter with activity in the desired tissue or condition but that do not have activity in any other common tissue.

To identify the promoter sequence, the 5' portions of the clones described here are analyzed for sequences characteristic of promoter sequences. For instance, promoter sequence elements include the TATA box consensus sequence (TATAAT), which is usually an AT-rich stretch of 5-10 bp located approximately 20 to 40 base pairs upstream of the transcription start site. Identification of the TATA box is well known in the art. For example, one way to predict the location of this element is to identify the transcription start site using standard RNA-mapping techniques such as primer extension, S1 analysis, and/or RNase protection. To confirm the presence of the AT-rich sequence, a structure-function analysis can be performed involving mutagenesis of the putative region and quantification of the mutation's effect on expression of a linked downstream reporter gene. See, e.g., *The Maize Handbook*, Chapter 114, Freeling and Walbot, Eds., Springer, New York, (1994).

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In plants, further upstream from the TATA box, at positions -80 to -100, there is typically a promoter element (i.e., the CAAT box) with a series of adenines surrounding the trinucleotide G (or T) N G. J. Messing et al., in Genetic Engineering in Plants, Kosage, Meredith and Hollaender, Eds., pp. 221-227 1983. In maize, there is no well conserved CAAT box but there are several short, conserved protein-binding motifs upstream of the TATA box. These include motifs for the trans-acting transcription factors involved in light regulation, anaerobic induction, hormonal regulation, or anthocyanin biosynthesis, as appropriate for each gene.

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Once promoter and/or gene sequences are known, a region of suitable size is selected from the genomic DNA that is 5' to the transcriptional start, or the translational start site, and such sequences are then linked to a coding sequence. If the transcriptional start site is used as the point of fusion, any of a number of possible 5' untranslated regions can be used in between the transcriptional start site and the partial coding sequence. If the translational start site at the 3' end of the specific promoter is used, then it is linked directly to the methionine start codon of a coding sequence.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The 3' end sequence to be added can be derived from, for example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

An intron sequence can be added to the 5' untranslated region or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold. Buchman and Berg, *Mol. Cell Biol.* 8: 4395-4405 (1988); Callis *et al.*, *Genes Dev.* 1: 1183-1200 (1987). Such intron enhancement of gene expression is typically greatest when placed near the 5' end of the transcription unit. Use of maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. See generally, *The Maize Handbook*, Chapter 116, Freeling and Walbot, Eds., Springer, New York (1994).

The vector comprising the sequences from a polynucleotide of the present invention will typically comprise a marker gene which confers a selectable phenotype on

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plant cells. Usually, the selectable marker gene will encode antibiotic resistance, with suitable genes including genes coding for resistance to the antibiotic spectinomycin (e.g., the aada gene), the streptomycin phosphotransferase (SPT) gene coding for streptomycin resistance, the neomycin phosphotransferase (NPTII) gene encoding kanamycin or geneticin resistance, the hygromycin phosphotransferase (HPT) gene coding for hygromycin resistance, genes coding for resistance to herbicides which act to inhibit the action of acetolactate synthase (ALS), in particular the sulfonylurea-type herbicides (e.g., the acetolactate synthase (ALS) gene containing mutations leading to such resistance in particular the S4 and/or Hra mutations), genes coding for resistance to herbicides which act to inhibit action of glutamine synthase, such as phosphinothricin or basta (e.g., the bar gene), or other such genes known in the art. The bar gene encodes resistance to the herbicide basta, the nptII gene encodes resistance to the antibiotics kanamycin and geneticin, and the ALS gene encodes resistance to the herbicide chlorsulfuron.

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Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of Agrobacterium tumefaciens described by Rogers et al., Meth. In Enzymol., 153:253-277 (1987). These vectors are plant integrating vectors in that on transformation, the vectors integrate a portion of vector DNA into the genome of the host plant. Exemplary A. tumefaciens vectors useful herein are plasmids pKYLX6 and pKYLX7 of Schardl et al., Gene, 61:1-11 (1987) and Berger et al., Proc. Natl. Acad. Sci. U.S.A., 86:8402-8406 (1989). Another useful vector herein is plasmid pBI101.2 that is available from Clontech Laboratories, Inc. (Palo Alto, CA).

A polynucleotide of the present invention can be expressed in either sense or antisense orientation as desired. It will be appreciated that control of gene expression in either sense or anti-sense orientation can have a direct impact on the observable plant characteristics. Antisense technology can be conveniently used to gene expression in plants. To accomplish this, a nucleic acid segment from the desired gene is cloned and operably linked to a promoter such that the anti-sense strand of RNA will be transcribed. The construct is then transformed into plants and the antisense strand of RNA is produced. In plant cells, it has been shown that antisense RNA inhibits gene expression by preventing the accumulation of mRNA which encodes the enzyme of interest, see,

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e.g., Sheehy et al., Proc. Nat'l. Acad. Sci. (USA) 85: 8805-8809 (1988); and Hiatt et al., U.S. Patent No. 4,801,340.

Another method of suppression is sense suppression. Introduction of nucleic acid configured in the sense orientation has been shown to be an effective means by which to block the transcription of target genes. For an example of the use of this method to modulate expression of endogenous genes see, Napoli *et al.*, *The Plant Cell* 2: 279-289 (1990) and U.S. Patent No. 5,034,323.

Catalytic RNA molecules or ribozymes can also be used to inhibit expression of plant genes. It is possible to design ribozymes that specifically pair with virtually any target RNA and cleave the phosphodiester backbone at a specific location, thereby functionally inactivating the target RNA. In carrying out this cleavage, the ribozyme is not itself altered, and is thus capable of recycling and cleaving other molecules, making it a true enzyme. The inclusion of ribozyme sequences within antisense RNAs confers RNA-cleaving activity upon them, thereby increasing the activity of the constructs. The design and use of target RNA-specific ribozymes is described in Haseloff *et al.*, *Nature* 334: 585-591 (1988).

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A variety of cross-linking agents, alkylating agents and radical generating species as pendant groups on polynucleotides of the present invention can be used to bind, label, detect, and/or cleave nucleic acids. For example, Vlassov, V. V., et al., Nucleic Acids Res (1986) 14:4065-4076, describe covalent bonding of a single-stranded DNA fragment with alkylating derivatives of nucleotides complementary to target sequences. A report of similar work by the same group is that by Knorre, D. G., et al., Biochimie (1985) 67:785-789. Iverson and Dervan also showed sequence-specific cleavage of singlestranded DNA mediated by incorporation of a modified nucleotide which was capable of activating cleavage (J Am Chem Soc (1987) 109:1241-1243). Meyer, R. B., et al., J Am Chem Soc (1989) 111:8517-8519, effect covalent crosslinking to a target nucleotide using an alkylating agent complementary to the single-stranded target nucleotide sequence. A photoactivated crosslinking to single-stranded oligonucleotides mediated by psoralen was disclosed by Lee, B. L., et al., Biochemistry (1988) 27:3197-3203. Use of crosslinking in triple-helix forming probes was also disclosed by Home, et al., J Am Chem Soc (1990) 112:2435-2437. Use of N4, N4-ethanocytosine as an alkylating agent to crosslink to single-stranded oligonucleotides has also been described by Webb and Matteucci, J Am Chem Soc (1986) 108:2764-2765; Nucleic Acids Res (1986) 14:7661-7674; Feteritz

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et al., J. Am. Chem. Soc. 113:4000 (1991). Various compounds to bind, detect, label, and/or cleave nucleic acids are known in the art. See, for example, U.S. Patent Nos. 5,543,507; 5,672,593; 5,484,908; 5,256,648; and, 5,681941.

5 Proteins

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The isolated proteins of the present invention comprise a polypeptide having at least 10 amino acids encoded by any one of the polynucleotides of the present invention as discussed more fully, above, or polypeptides which are conservatively modified variants thereof. The proteins of the present invention or variants thereof can comprise any number of contiguous amino acid residues from a polypeptide of the present invention, wherein that number is selected from the group of integers consisting of from 10 to the number of residues in a full-length polypeptide of the present invention.

Optionally, this subsequence of contiguous amino acids is at least 15, 20, 25, 30, 35, or 40 amino acids in length, often at least 50, 60, 70, 80, or 90 amino acids in length. Further, the number of such subsequences can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5.

As those of skill will appreciate, the present invention includes catalytically active polypeptides of the present invention (i.e., enzymes). Catalytically active polypeptides have a specific activity of at least 20%, 30%, or 40%, and preferably at least 50%, 60%, or 70%, and most preferably at least 80%, 90%, or 95% that of the native (nonsynthetic), endogenous polypeptide. Further, the substrate specificity (k_{cat}/K_m) is optionally substantially similar to the native (non-synthetic), endogenous polypeptide. Typically, the K_m will be at least 30%, 40%, or 50%, that of the native (non-synthetic), endogenous polypeptide; and more preferably at least 60%, 70%, 80%, or 90%. Methods of assaying and quantifying measures of enzymatic activity and substrate specificity (k_{cat}/K_m), are well known to those of skill in the art.

Generally, the proteins of the present invention will, when presented as an immunogen, elicit production of an antibody specifically reactive to a polypeptide of the present invention. Further, the proteins of the present invention will not bind to antisera raised against a polypeptide of the present invention which has been fully immunosorbed with the same polypeptide. Immunoassays for determining binding are well known to those of skill in the art. A preferred immunoassay is a competitive immunoassay as discussed, *infra*. Thus, the proteins of the present invention can be employed as

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immunogens for constructing antibodies immunoreactive to a protein of the present invention for such exemplary utilities as immunoassays or protein purification techniques.

5 Expression of Proteins in Host Cells

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Using the nucleic acids of the present invention, one may express a protein of the present invention in a recombinantly engineered cell such as bacteria, yeast, insect, mammalian, or preferably plant cells. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. One of skill would recognize that modifications can be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located purification sequences. Restriction sites or termination codons can also be introduced.

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A. Expression in Prokaryotes

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al., Nature 198:1056 (1977)), the tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. 8:4057 (1980)) and the lambda derived P L promoter and N-gene ribosome binding site (Shimatake *et al.*, Nature 292:128 (1981)). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using Bacillus sp. and Salmonella (Palva, et al., Gene 22: 229-235 (1983); Mosbach, et al., Nature 302: 543-545 (1983)).

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B. Expression in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. As explained briefly below, a of the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells, as discussed *infra*, are employed as expression systems for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F., et al., Methods in Yeast Genetics, Cold Spring Harbor Laboratory (1982) is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeast for production of eukaryotic proteins are Saccharomyces cerevisiae and Pichia pastoris. Vectors, strains, and protocols for expression in Saccharomyces and Pichia are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as

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promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay of other standard immunoassay techniques.

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The sequences encoding proteins of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Illustrative of cell cultures useful for the production of the peptides are mammalian cells. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV tk promoter or pgk (phosphoglycerate kinase) promoter), an enhancer (Queen et al., Immunol. Rev. 89: 49 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992).

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line (See Schneider, *J. Embryol. Exp. Morphol.* 27: 353-365 (1987).

As with yeast, when higher animal or plant host cells are employed, polyadenlyation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenlyation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague, et al., J. Virol. 45: 773-781 (1983)). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a

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Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D.M. Glover, Ed., IRL Press, Arlington, Virginia pp. 213-238 (1985).

Transfection/Transformation of Cells

The method of transformation/transfection is not critical to the instant invention; various methods of transformation or transfection are currently available. As newer methods are available to transform crops or other host cells they may be directly applied. Accordingly, a wide variety of methods have been developed to insert a DNA sequence into the genome of a host cell to obtain the transcription and/or translation of the sequence to effect phenotypic changes in the organism. Thus, any method which provides for efficient transformation/transfection may be employed.

A. Plant Transformation

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A DNA sequence coding for the desired polynucleotide of the present invention, for example a cDNA or a genomic sequence encoding a full length protein, will be used to construct a recombinant expression cassette which can be introduced into the desired plant.

Isolated nucleic acid acids of the present invention can be introduced into plants according techniques known in the art. Generally, recombinant expression cassettes as described above and suitable for transformation of plant cells are prepared. Techniques for transforming a wide variety of higher plant species are well known and described in the technical, scientific, and patent literature. See, for example, Weising et al., Ann. Rev. Genet. 22: 421-477 (1988). For example, the DNA construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation, PEG poration, particle bombardment, silicon fiber delivery, or microinjection of plant cell protoplasts or embryogenic callus. See e.g., Tomes, et al., Direct DNA Transfer into Intact Plant Cells via Microprojectile Bombardment, pp. 197-213 in Plant Cell, Tissue and Organ Culture, Fundamental Methods, (eds. O.L. Gamborg and G.C. Phillips, Springer-Verlag Berlin Heidelberg New York, 1995). Alternatively, the DNA constructs may be combined with suitable T-DNA flanking regions and introduced into a conventional Agrobacterium tumefaciens host vector. The virulence functions of the Agrobacterium tumefaciens host will direct the insertion of the construct and adjacent

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marker into the plant cell DNA when the cell is infected by the bacteria. See, U.S. Patent No. 5,591,616.

The introduction of DNA constructs using polyethylene glycol precipitation is described in Paszkowski et al., Embo J. 3: 2717-2722 (1984). Electroporation techniques are described in Fromm et al., Proc. Natl. Acad. Sci. 82: 5824 (1985). Ballistic transformation techniques are described in Klein et al., Nature 327: 70-73 (1987).

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Agrobacterium tumefaciens-meditated transformation techniques are well described in the scientific literature. See, for example Horsch et al., Science 233: 496-498 (1984), and Fraley et al., Proc. Natl. Acad. Sci. 80: 4803 (1983). Although Agrobacterium is useful primarily in dicots, certain monocots can be transformed by Agrobacterium. For instance, Agrobacterium transformation of maize is described in U.S. Patent No. 5,550,318.

Other methods of transfection or transformation include (1) Agrobacterium rhizogenes-mediated transformation (see, e.g., Lichtenstein and Fuller In: Genetic Engineering, vol. 6, PWJ Rigby, Ed., London, Academic Press, 1987; and Lichtenstein, C. P., and Draper, J., In: DNA Cloning, Vol. II, D. M. Glover, Ed., Oxford, IRI Press, 1985), Application PCT/US87/02512 (WO 88/02405 published Apr. 7, 1988) describes the use of A. rhizogenes strain A4 and its Ri plasmid along with A. tumefaciens vectors pARC8 or pARC16 (2) liposome-mediated DNA uptake (see, e.g., Freeman et al., Plant Cell Physiol. 25: 1353, 1984), (3) the vortexing method (see, e.g., Kindle, Proc. Natl. Acad. Sci., USA 87: 1228, (1990).

DNA can also be introduced into plants by direct DNA transfer into pollen as described by Zhou *et al.*, Methods in Enzymology, 101:433 (1983); D. Hess, Intern Rev. Cytol., 107:367 (1987); Luo *et al.*, Plane Mol. Biol. Reporter, 6:165 (1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into reproductive organs of a plant as described by Pena *et al.*, Nature, 325.:274 (1987). DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described by Neuhaus *et al.*, Theor. Appl. Genet., 75:30 (1987); and Benbrook *et al.*, in Proceedings Bio Expo 1986, Butterworth, Stoneham, Mass., pp. 27-54 (1986). A variety of plant viruses that can be employed as vectors are known in the art and include cauliflower mosaic virus (CaMV), geminivirus, brome mosaic virus, and tobacco mosaic virus.

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B. Transfection of Prokaryotes, Lower Eukaryotes, and Animal Cells

Animal and lower eukaryotic (e.g., yeast) host cells are competent or rendered competent for transfection by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation, biolistics, and micro-injection of the DNA directly into the cells. The transfected cells are cultured by means well known in the art. Kuchler, R.J., *Biochemical Methods in Cell Culture and Virology*, Dowden, Hutchinson and Ross, Inc. (1977).

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Synthesis of Proteins

The proteins of the present invention can be constructed using non-cellular synthetic methods. Solid phase synthesis of proteins of less than about 50 amino acids in length may be accomplished by attaching the C-terminal amino acid of the sequence to an insoluble support followed by sequential addition of the remaining amino acids in the sequence. Techniques for solid phase synthesis are described by Barany and Merrifield, Solid-Phase Peptide Synthesis, pp. 3-284 in *The Peptides: Analysis, Synthesis, Biology.* Vol. 2: Special Methods in Peptide Synthesis, Part A.; Merrifield, et al., J. Am. Chem. Soc. 85: 2149-2156 (1963), and Stewart et al., Solid Phase Peptide Synthesis, 2nd ed., Pierce Chem. Co., Rockford, Ill. (1984). Proteins of greater length may be synthesized by condensation of the amino and carboxy termini of shorter fragments. Methods of forming peptide bonds by activation of a carboxy terminal end (e.g., by the use of the coupling reagent N,N'-dicycylohexylcarbodiimide)) is known to those of skill.

25 **Purification of Proteins**

The proteins of the present invention may be purified by standard techniques well known to those of skill in the art. Recombinantly produced proteins of the present invention can be directly expressed or expressed as a fusion protein. The recombinant protein is purified by a combination of cell lysis (e.g., sonication, French press) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme releases the desired recombinant protein.

The proteins of this invention, recombinant or synthetic, may be purified to substantial purity by standard techniques well known in the art, including detergent

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solubilization, selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, Protein Purification: Principles and Practice, Springer-Verlag: New York (1982); Deutscher, Guide to Protein Purification, Academic Press (1990). For example, antibodies may be raised to the proteins as described herein. Purification from E. coli can be achieved following procedures described in U.S. Patent No. 4,511,503. The protein may then be isolated from cells expressing the protein and further purified by standard protein chemistry techniques as described herein. Detection of the expressed protein is achieved by methods known in the art and include, for example, radioimmunoassays, Western blotting techniques or immunoprecipitation.

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Transgenic Plant Regeneration

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Transformed plant cells which are derived by any of the above transformation techniques can be cultured to regenerate a whole plant which possesses the transformed genotype. Such regeneration techniques often rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker which has been introduced together with a polynucleotide of the present invention. For transformation and regeneration of maize see, Gordon-Kamm et al., The Plant Cell, 2:603-618 (1990).

Plants cells transformed with a plant expression vector can be regenerated, e.g., from single cells, callus tissue or leaf discs according to standard plant tissue culture techniques. It is well known in the art that various cells, tissues, and organs from almost any plant can be successfully cultured to regenerate an entire plant. Plant regeneration from cultured protoplasts is described in Evans et al., Protoplasts Isolation and Culture, Handbook of Plant Cell Culture, Macmillilan Publishing Company, New York, pp. 124-176 (1983); and Binding, Regeneration of Plants, Plant Protoplasts, CRC Press, Boca Raton, pp. 21-73 (1985).

The regeneration of plants containing the foreign gene introduced by Agrobacterium from leaf explants can be achieved as described by Horsch et al., Science, 227:1229-1231 (1985). In this procedure, transformants are grown in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant species being transformed as described by Fraley et al., Proc. Natl. Acad. Sci. U.S.A., 80:4803 (1983). This procedure typically produces shoots within two to four

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weeks and these transformant shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth.

Transgenic plants of the present invention may be fertile or sterile.

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Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee et al., Ann. Rev. of Plant Phys. 38: 467-486 (1987). The regeneration of plants from either single plant protoplasts or various explants is well known in the art. See, for example, Methods for Plant Molecular Biology, A. Weissbach and H. Weissbach, eds., Academic Press, Inc., San Diego, Calif. (1988). This regeneration and growth process includes the steps of selection of transformant cells and shoots, rooting the transformant shoots and growth of the plantlets in soil. For maize cell culture and regeneration see generally, The Maize Handbook, Freeling and Walbot, Eds., Springer, New York (1994); Corn and Corn Improvement, 3rd edition, Sprague and Dudley Eds., American Society of Agronomy, Madison, Wisconsin (1988).

One of skill will recognize that after the recombinant expression cassette is stably incorporated in transgenic plants and confirmed to be operable, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed.

In vegetatively propagated crops, mature transgenic plants can be propagated by the taking of cuttings or by tissue culture techniques to produce multiple identical plants. Selection of desirable transgenics is made and new varieties are obtained and propagated vegetatively for commercial use. In seed propagated crops, mature transgenic plants can be self-crossed to produce a homozygous inbred plant. The inbred plant produces seed containing the newly introduced heterologous nucleic acid. These seeds can be grown to produce plants that would produce the selected phenotype.

Parts obtained from the regenerated plant, such as flowers, seeds, leaves, branches, fruit, and the like are included in the invention, provided that these parts comprise cells comprising the isolated nucleic acid of the present invention. Progeny and variants, and mutants of the regenerated plants are also included within the scope of the invention, provided that these parts comprise the introduced nucleic acid sequences.

Transgenic plants expressing the selectable marker can be screened for transmission of the nucleic acid of the present invention by, for example, standard immunoblot and DNA detection techniques. Transgenic lines are also typically evaluated

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on levels of expression of the heterologous nucleic acid. Expression at the RNA level can be determined initially to identify and quantitate expression-positive plants. Standard techniques for RNA analysis can be employed and include PCR amplification assays using oligonucleotide primers designed to amplify only the heterologous RNA templates and solution hybridization assays using heterologous nucleic acid-specific probes. The RNA-positive plants can then analyzed for protein expression by Western immunoblot analysis using the specifically reactive antibodies of the present invention. In addition, in situ hybridization and immunocytochemistry according to standard protocols can be done using heterologous nucleic acid specific polynucleotide probes and antibodies, respectively, to localize sites of expression within transgenic tissue. Generally, a number of transgenic lines are usually screened for the incorporated nucleic acid to identify and select plants with the most appropriate expression profiles.

A preferred embodiment is a transgenic plant that is homozygous for the added heterologous nucleic acid; i.e., a transgenic plant that contains two added nucleic acid sequences, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) a heterozygous transgenic plant that contains a single added heterologous nucleic acid, germinating some of the seed produced and analyzing the resulting plants produced for altered expression of a polynucleotide of the present invention relative to a control plant (i.e., native, non-transgenic). Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

Modulating Polypeptide Levels and/or Composition

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The present invention further provides a method for modulating (i.e., increasing or decreasing) the concentration or composition of the polypeptides of the present invention in a plant or part thereof. Modulation can be effected by increasing or decreasing the concentration and/or the composition (i.e., the ratio of the polypeptides of the present invention) in a plant. The method comprises transforming a plant cell with a recombinant expression cassette comprising a polynucleotide of the present invention as described above to obtain a transformed plant cell, growing the transformed plant cell under plant forming conditions, and inducing expression of a polynucleotide of the present invention in the plant for a time sufficient to modulate concentration and/or composition in the plant or plant part.

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In some embodiments, the content and/or composition of polypeptides of the present invention in a plant may be modulated by altering, in vivo or in vitro, the promoter of a non-isolated gene of the present invention to up- or down-regulate gene expression. In some embodiments, the coding regions of native genes of the present invention can be altered via substitution, addition, insertion, or deletion to decrease activity of the encoded enzyme. See, e.g., Kmiec, U.S. Patent 5,565,350; Zarling et al., PCT/US93/03868. And in some embodiments, an isolated nucleic acid (e.g., a vector) comprising a promoter sequence is transfected into a plant cell. Subsequently, a plant cell comprising the promoter operably linked to a polynucleotide of the present invention is selected for by means known to those of skill in the art such as, but not limited to, Southern blot, DNA sequencing, or PCR analysis using primers specific to the promoter and to the gene and detecting amplicons produced therefrom. A plant or plant part altered or modified by the foregoing embodiments is grown under plant forming conditions for a time sufficient to modulate the concentration and/or composition of polypeptides of the present invention in the plant. Plant forming conditions are well known in the art and discussed briefly, above.

In general, concentration or composition is increased or decreased by at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90% relative to a native control plant, plant part, or cell lacking the aforementioned recombinant expression cassette. Modulation in the present invention may occur during and/or subsequent to growth of the plant to the desired stage of development. Modulating nucleic acid expression temporally and/or in particular tissues can be controlled by employing the appropriate promoter operably linked to a polynucleotide of the present invention in, for example, sense or antisense orientation as discussed in greater detail, *above*. Induction of expression of a polynucleotide of the present invention can also be controlled by exogenous administration of an effective amount of inducing compound. Inducible promoters and inducing compounds which activate expression from these promoters are well known in the art. In preferred embodiments, the polypeptides of the present invention are modulated in monocots, particularly maize.

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Molecular Markers

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The present invention provides a method of genotyping a plant comprising a polynucleotide of the present invention. Preferably, the plant is a monocot, such as maize or sorghum. Genotyping provides a means of distinguishing homologs of a chromosome pair and can be used to differentiate segregants in a plant population. Molecular marker methods can be used for phylogenetic studies, characterizing genetic relationships among crop varieties, identifying crosses or somatic hybrids, localizing chromosomal segments affecting monogenic traits, map based cloning, and the study of quantitative inheritance. See, e.g., *Plant Molecular Biology: A Laboratory Manual*, Chapter 7, Clark, Ed., Springer-Verlag, Berlin (1997). For molecular marker methods, see generally, The DNA Revolution by Andrew H. Paterson 1996 (Chapter 2) in: Genome Mapping in Plants (ed. Andrew H. Paterson) by Academic Press/R. G. Landis Company, Austin, Texas, pp.7-21.

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The particular method of genotyping in the present invention may employ any number of molecular marker analytic techniques such as, but not limited to, restriction fragment length polymorphisms (RFLPs). RFLPs are the product of allelic differences between DNA restriction fragments caused by nucleotide sequence variability. As is well known to those of skill in the art, RFLPs are typically detected by extraction of genomic DNA and digestion with a restriction enzyme. Generally, the resulting fragments are separated according to size and hybridized with a probe; single copy probes are preferred. Restriction fragments from homologous chromosomes are revealed. Differences in fragment size among alleles represent an RFLP. Thus, the present invention further provides a means to follow segregation of a gene or nucleic acid of the present invention as well as chromosomal sequences genetically linked to these genes or nucleic acids using such techniques as RFLP analysis. Linked chromosomal sequences are within 50 centiMorgans (cM), often within 40 or 30 cM, preferably within 20 or 10 cM, more preferably within 5, 3, 2, or 1 cM of a gene of the present invention.

In the present invention, the nucleic acid probes employed for molecular marker mapping of plant nuclear genomes selectively hybridize, under selective hybridization conditions, to a gene encoding a polynucleotide of the present invention. In preferred embodiments, the probes are selected from polynucleotides of the present invention. Typically, these probes are cDNA probes or *Pst I* genomic clones. The length of the probes is discussed in greater detail, above, but are typically at least 15 bases in length,

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more preferably at least 20, 25, 30, 35, 40, or 50 bases in length. Generally, however, the probes are less than about 1 kilobase in length. Preferably, the probes are single copy probes that hybridize to a unique locus in a haploid chromosome complement. Some exemplary restriction enzymes employed in RFLP mapping are *EcoRI*, *EcoRv*, and *SstI*. As used herein the term "restriction enzyme" includes reference to a composition that recognizes and, alone or in conjunction with another composition, cleaves at a specific nucleotide sequence.

The method of detecting an RFLP comprises the steps of (a) digesting genomic DNA of a plant with a restriction enzyme; (b) hybridizing a nucleic acid probe, under selective hybridization conditions, to a sequence of a polynucleotide of the present of said genomic DNA; (c) detecting therefrom a RFLP. Other methods of differentiating polymorphic (allelic) variants of polynucleotides of the present invention can be had by utilizing molecular marker techniques well known to those of skill in the art including such techniques as: 1) single stranded conformation analysis (SSCP); 2) denaturing gradient gel electrophoresis (DGGE); 3) RNase protection assays; 4) allele-specific oligonucleotides (ASOs); 5) the use of proteins which recognize nucleotide mismatches, such as the E. coli mutS protein; and 6) allele-specific PCR. Other approaches based on the detection of mismatches between the two complementary DNA strands include clamped denaturing gel electrophoresis (CDGE); heteroduplex analysis (HA); and chemical mismatch cleavage (CMC). Exemplary polymorphic variants are provided in Table I, above. Thus, the present invention further provides a method of genotyping comprising the steps of contacting, under stringent hybridization conditions, a sample suspected of comprising a polynucleotide of the present invention with a nucleic acid probe. Generally, the sample is a plant sample; preferably, a sample suspected of comprising a maize polynucleotide of the present invention (e.g., gene, mRNA). The nucleic acid probe selectively hybridizes, under stringent conditions, to a subsequence of a polynucleotide of the present invention comprising a polymorphic marker. Selective hybridization of the nucleic acid probe to the polymorphic marker nucleic acid sequence yields a hybridization complex. Detection of the hybridization complex indicates the presence of that polymorphic marker in the sample. In preferred embodiments, the nucleic acid probe comprises a polynucleotide of the present invention.

UTR's and Codon Preference

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In general, translational efficiency has been found to be regulated by specific sequence elements in the 5' non-coding or untranslated region (5' UTR) of the RNA. Positive sequence motifs include translational initiation consensus sequences (Kozak, Nucleic Acids Res. 15:8125 (1987)) and the 7-methylguanosine cap structure (Drummond et al., Nucleic Acids Res. 13:7375 (1985)). Negative elements include stable intramolecular 5' UTR stem-loop structures (Muesing et al., Cell 48:691 (1987)) and AUG sequences or short open reading frames preceded by an appropriate AUG in the 5' UTR (Kozak, above, Rao et al., Mol. and Cell. Biol. 8:284 (1988)). Accordingly, the present invention provides 5' and/or 3' UTR regions for modulation of translation of heterologous coding sequences.

Further, the polypeptide-encoding segments of the polynucleotides of the present invention can be modified to alter codon usage. Altered codon usage can be employed to alter translational efficiency and/or to optimize the coding sequence for expression in a desired host or to optimize the codon usage in a heterologous sequence for expression in maize. Codon usage in the coding regions of the polynucleotides of the present invention can be analyzed statistically using commercially available software packages such as "Codon Preference" available from the University of Wisconsin Genetics Computer Group (see Devereaux et al., Nucleic Acids Res. 12: 387-395 (1984)) or MacVector 4.1 (Eastman Kodak Co., New Haven, Conn.). Thus, the present invention provides a codon usage frequency characteristic of the coding region of at least one of the polynucleotides of the present invention. The number of polynucleotides that can be used to determine a codon usage frequency can be any integer from 1 to the number of polynucleotides of the present invention as provided herein. Optionally, the polynucleotides will be full-length sequences. An exemplary number of sequences for statistical analysis can be at least 1, 5, 10, 20, 50, or 100.

Sequence Shuffling

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The present invention provides methods for sequence shuffling using polynucleotides of the present invention, and compositions resulting therefrom. Sequence shuffling is described in PCT publication No. 96/19256. See also, Zhang, J.-H., et al. Proc. Natl. Acad. Sci. USA 94:4504-4509 (1997). Generally, sequence shuffling provides a means for generating libraries of polynucleotides having a desired characteristic which can be selected or screened for. Libraries of recombinant

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polynucleotides are generated from a population of related sequence polynucleotides which comprise sequence regions which have substantial sequence identity and can be homologously recombined in vitro or in vivo. The population of sequence-recombined polynucleotides comprises a subpopulation of polynucleotides which possess desired or advantageous characteristics and which can be selected by a suitable selection or screening method. The characteristics can be any property or attribute capable of being selected for or detected in a screening system, and may include properties of: an encoded protein, a transcriptional element, a sequence controlling transcription, RNA processing. RNA stability, chromatin conformation, translation, or other expression property of a gene or transgene, a replicative element, a protein-binding element, or the like, such as any feature which confers a selectable or detectable property. In some embodiments, the selected characteristic will be a decreased K_m and/or increased K_{cat} over the wild-type protein as provided herein. In other embodiments, a protein or polynculeotide generated from sequence shuffling will have a ligand binding affinity greater than the non-shuffled wild-type polynucleotide. The increase in such properties can be at least 110%, 120%, 130%, 140% or at least 150% of the wild-type value.

Generic and Consensus Sequences

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Polynuclotides and polypeptides of the present invention further include those having: (a) a generic sequence of at least two homologous polynucleotides or polypeptides, respectively, of the present invention; and, (b) a consensus sequence of at least three homologous polynucleotides or polypeptides, respectively, of the present invention. The generic sequence of the present invention comprises each species of polypeptide or polynucleotide embraced by the generic polypeptide or polynucleotide, sequence, respectively. The individual species encompassed by a polynucleotide having an amino acid or nucleic acid consensus sequence can be used to generate antibodies or produce nucleic acid probes or primers to screen for homologs in other species, genera, families, orders, classes, phyla, or kingdoms. For example, a polynucleotide having a consensus sequences from a gene family of *Zea mays* can be used to generate antibody or nucleic acid probes or primers to other *Gramineae* species such as wheat, rice, or sorghum. Alternatively, a polynucleotide having a consensus sequence generated from orthologous genes can be used to identify or isolate orthologs of other taxa. Typically, a polynucleotide having a consensus sequence will be at least 9, 10, 15, 20, 25, 30, or 40

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amino acids in length, or 20, 30, 40, 50, 100, or 150 nucleotides in length. As those of skill in the art are aware, a conservative amino acid substitution can be used for amino acids which differ amongst aligned sequence but are from the same conservative substitution group as discussed above. Optionally, no more than 1 or 2 conservative amino acids are substituted for each 10 amino acid length of consensus sequence.

Similar sequences used for generation of a consensus or generic sequence include any number and combination of allelic variants of the same gene, orthologous, or paralogous sequences as provided herein. Optionally, similar sequences used in generating a consensus or generic sequence are identified using the BLAST algorithm's smallest sum probability (P(N)). Various suppliers of sequence-analysis software are listed in chapter 7 of Current Protocols in Molecular Biology, F.M. Ausubel et al., Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (Supplement 30). A polynucleotide sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.1, more preferably less than about 0.01, or 0.001, and most preferably less than about 0.0001, or 0.00001. Similar polynucleotides can be aligned and a consensus or generic sequence generated using multiple sequence alignment software available from a number of commercial suppliers such as the Genetics Computer Group's (Madison, WI) PILEUP software, Vector NTI's (North Bethesda, MD) ALIGNX, or Genecode's (Ann Arbor, MI) SEQUENCHER. Conveniently, default parameters of such software can be used to generate consensus or generic sequences.

Detection of Nucleic Acids

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The present invention further provides methods for detecting a polynucleotide of the present invention in a nucleic acid sample suspected of comprising a polynucleotide of the present invention, such as a plant cell lysate, particularly a lysate of corn. In some embodiments, a gene of the present invention or portion thereof can be amplified prior to the step of contacting the nucleic acid sample with a polynucleotide of the present invention. The nucleic acid sample is contacted with the polynucleotide to form a hybridization complex. The polynucleotide hybridizes under stringent conditions to a gene encoding a polypeptide of the present invention. Formation of the hybridization complex is used to detect a gene encoding a polypeptide of the present invention in the

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nucleic acid sample. Those of skill will appreciate that an isolated nucleic acid comprising a polynucleotide of the present invention should lack cross-hybridizing sequences in common with non-target genes that would yield a false positive result.

Detection of the hybridization complex can be achieved using any number of well known methods. For example, the nucleic acid sample, or a portion thereof, may be assayed by hybridization formats including but not limited to, solution phase, solid phase, mixed phase, or in situ hybridization assays. Briefly, in solution (or liquid) phase hybridizations, both the target nucleic acid and the probe or primer are free to interact in the reaction mixture. In solid phase hybridization assays, probes or primers are typically linked to a solid support where they are available for hybridization with target nucleic in solution. In mixed phase, nucleic acid intermediates in solution hybridize to target nucleic acids in solution as well as to a nucleic acid linked to a solid support. In in situ hybridization, the target nucleic acid is liberated from its cellular surroundings in such as to be available for hybridization within the cell while preserving the cellular morphology for subsequent interpretation and analysis. The following articles provide an overview of the various hybridization assay formats: Singer et al., Biotechniques 4(3): 230-250 (1986); Haase et al., Methods in Virology, Vol. VII, pp. 189-226 (1984); Wilkinson, The theory and practice of in situ hybridization in: In situ Hybridization, D.G. Wilkinson, Ed., IRL Press, Oxford University Press, Oxford; and Nucleic Acid Hybridization: A Practical Approach, Hames, B.D. and Higgins, S.J., Eds., IRL Press (1987).

Nucleic Acid Labels and Detection Methods

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The means by which nucleic acids of the present invention are labeled is not a

25 critical aspect of the present invention and can be accomplished by any number of methods currently known or later developed. Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, radioisotopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means.

Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads, fluorescent dyes (e.g., fluorescein, texas red, rhodamine, green fluorescent protein, and the like), radiolabels (e.g., ³H, ¹²⁵I, ³⁵S, ¹⁴C, or ³²P), enzymes (e.g., horse radish peroxidase, alkaline phosphatase and others commonly used

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in an ELISA), and colorimetric labels such as colloidal gold or colored glass or plastic (e.g., polystyrene, polypropylene, latex, etc.) beads.

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Nucleic acids of the present invention can be labeled by any one of several methods typically used to detect the presence of hybridized nucleic acids. One common method of detection is the use of autoradiography using probes labeled with ³H, ¹²⁵I, ³⁵S, ¹⁴C, or ³²P, or the like. The choice of radio-active isotope depends on research preferences due to ease of synthesis, stability, and half lives of the selected isotopes. Other labels include ligands which bind to antibodies labeled with fluorophores, chemiluminescent agents, and enzymes. Alternatively, probes can be conjugated directly with labels such as fluorophores, chemiluminescent agents or enzymes. The choice of label depends on sensitivity required, ease of conjugation with the probe, stability requirements, and available instrumentation. Labeling the nucleic acids of the present invention is readily achieved such as by the use of labeled PCR primers.

In some embodiments, the label is simultaneously incorporated during the amplification step in the preparation of the nucleic acids. Thus, for example, polymerase chain reaction (PCR) with labeled primers or labeled nucleotides will provide a labeled amplification product. In another embodiment, transcription amplification using a labeled nucleotide (e.g., fluorescein-labeled UTP and/or CTP) incorporates a label into the transcribed nucleic acids.

Non-radioactive probes are often labeled by indirect means. For example, a ligand molecule is covalently bound to the probe. The ligand then binds to an anti-ligand molecule which is either inherently detectable or covalently bound to a detectable signal system, such as an enzyme, a fluorophore, or a chemiluminescent compound. Enzymes of interest as labels will primarily be hydrolases, such as phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc. Chemiluminescers include luciferin, and 2,3-dihydrophthalazinediones, e.g., luminol. Ligands and anti-ligands may be varied widely. Where a ligand has a natural anti-ligand, namely ligands such as biotin, thyroxine, and cortisol, it can be used in conjunction with its labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

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Probes can also be labeled by direct conjugation with a label. For example, cloned DNA probes have been coupled directly to horseradish peroxidase or alkaline phosphatase, (Renz. M., and Kurz, K., A Colorimetric Method for DNA Hybridization, Nucl. Acids Res. 12: 3435-3444 (1984)) and synthetic oligonucleotides have been coupled directly with alkaline phosphatase (Jablonski, E., et al., Preparation of Oligodeoxynucleotide-Alkaline Phosphatase Conjugates and Their Use as Hybridization Probes, Nuc. Acids. Res. 14: 6115-6128 (1986); and Li P., et al., Enzyme-linked Synthetic Oligonucleotide probes: Non-Radioactive Detection of Enterotoxigenic Escherichia Coli in Faeca Specimens, Nucl. Acids Res. 15: 5275-5287 (1987)).

Means of detecting such labels are well known to those of skill in the art. Thus, for example, radiolabels may be detected using photographic film or scintillation counters, fluorescent markers may be detected using a photodetector to detect emitted light. Enzymatic labels are typically detected by providing the enzyme with a substrate and detecting the reaction product produced by the action of the enzyme on the substrate, and colorimetric labels are detected by simply visualizing the colored label.

Antibodies to Proteins

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Antibodies can be raised to a protein of the present invention, including individual, allelic, strain, or species variants, and fragments thereof, both in their naturally occurring (full-length) forms and in recombinant forms. Additionally, antibodies are raised to these proteins in either their native configurations or in non-native configurations. Anti-idiotypic antibodies can also be generated. Many methods of making antibodies are known to persons of skill. The following discussion is presented as a general overview of the techniques available; however, one of skill will recognize that many variations upon the following methods are known.

A number of immunogens are used to produce antibodies specifically reactive with a protein of the present invention. An isolated recombinant, synthetic, or native polynucleotide of the present invention are the preferred immunogens (antigen) for the production of monoclonal or polyclonal antibodies. Those of skill will readily understand that the proteins of the present invention are typically denatured, and optionally reduced, prior to formation of antibodies for screening expression libraries or other assays in which a putative protein of the present invention is expressed or

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denatured in a non-native secondary, tertiary, or quartenary structure. Non-isolated polypeptides of the present invention can be used either in pure or impure form.

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The protein of the present invention is then injected into an animal capable of producing antibodies. Either monoclonal or polyclonal antibodies can be generated for subsequent use in immunoassays to measure the presence and quantity of the protein of the present invention. Methods of producing polyclonal antibodies are known to those of skill in the art. In brief, an immunogen (antigen), preferably a purified protein, a protein coupled to an appropriate carrier (e.g., GST, keyhole limpet hemanocyanin, etc.), or a protein incorporated into an immunization vector such as a recombinant vaccinia virus (see, U.S. Patent No. 4,722,848) is mixed with an adjuvant and animals are immunized with the mixture. The animal's immune response to the immunogen preparation is monitored by taking test bleeds and determining the titer of reactivity to the protein of interest. When appropriately high titers of antibody to the immunogen are obtained, blood is collected from the animal and antisera are prepared. Further fractionation of the antisera to enrich for antibodies reactive to the protein is performed where desired (See, e.g., Coligan, Current Protocols in Immunology, Wiley/Greene, NY (1991); and Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor Press, NY (1989)).

Antibodies, including binding fragments and single chain recombinant versions thereof, against predetermined fragments of a protein of the present invention are raised by immunizing animals, e.g., with conjugates of the fragments with carrier proteins as described above. Typically, the immunogen of interest is a protein of at least about 5 amino acids, more typically the protein is 10 amino acids in length, preferably, 15 amino acids in length and more preferably the protein is 20 amino acids in length or greater. The peptides are typically coupled to a carrier protein (e.g., as a fusion protein), or are recombinantly expressed in an immunization vector. Antigenic determinants on peptides to which antibodies bind are typically 3 to 10 amino acids in length.

Monoclonal antibodies are prepared from cells secreting the desired antibody. Monoclonals antibodies are screened for binding to a protein from which the immunogen was derived. Specific monoclonal and polyclonal antibodies will usually have an antibody binding site with an affinity constant for its cognate monovalent antigen at least between 10⁶-10⁷, usually at least 10⁸, preferably at least 10⁹, more preferably at least 10¹⁰, and most preferably at least 10¹¹ liters/mole.

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In some instances, it is desirable to prepare monoclonal antibodies from various mammalian hosts, such as mice, rodents, primates, humans, etc. Description of techniques for preparing such monoclonal antibodies are found in, e.g., Basic and Clinical Immunology, 4th ed., Stites et al., Eds., Lange Medical Publications, Los Altos, CA, and references cited therein; Harlow and Lane, Supra; Goding, Monoclonal Antibodies: Principles and Practice, 2nd ed., Academic Press, New York, NY (1986); and Kohler and Milstein, Nature 256: 495-497 (1975). Summarized briefly, this method proceeds by injecting an animal with an immunogen comprising a protein of the present invention. The animal is then sacrificed and cells taken from its spleen, which are fused with myeloma cells. The result is a hybrid cell or "hybridoma" that is capable of reproducing in vitro. The population of hybridomas is then screened to isolate individual clones, each of which secrete a single antibody species to the immunogen. In this manner, the individual antibody species obtained are the products of immortalized and cloned single B cells from the immune animal generated in response to a specific site recognized on the immunogenic substance.

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Other suitable techniques involve selection of libraries of recombinant antibodies in phage or similar vectors (see, e.g., Huse et al., Science 246: 1275-1281 (1989); and Ward, et al., Nature 341: 544-546 (1989); and Vaughan et al., Nature Biotechnology, 14: 309-314 (1996)). Alternatively, high avidity human monoclonal antibodies can be obtained from transgenic mice comprising fragments of the unrearranged human heavy and light chain Ig loci (i.e., minilocus transgenic mice). Fishwild et al., Nature Biotech., 14: 845-851 (1996). Also, recombinant immunoglobulins may be produced. See, Cabilly, U.S. Patent No. 4,816,567; and Queen et al., Proc. Nat'l Acad. Sci. 86: 10029-10033 (1989).

The antibodies of this invention are also used for affinity chromatography in isolating proteins of the present invention. Columns are prepared, e.g., with the antibodies linked to a solid support, e.g., particles, such as agarose, Sephadex, or the like, where a cell lysate is passed through the column, washed, and treated with increasing concentrations of a mild denaturant, whereby purified protein are released.

The antibodies can be used to screen expression libraries for particular expression products such as normal or abnormal protein. Usually the antibodies in such a procedure are labeled with a moiety allowing easy detection of presence of antigen by antibody binding.

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Antibodies raised against a protein of the present invention can also be used to raise anti-idiotypic antibodies. These are useful for detecting or diagnosing various pathological conditions related to the presence of the respective antigens.

Frequently, the proteins and antibodies of the present invention will be labeled by joining, either covalently or non-covalently, a substance which provides for a detectable signal. A wide variety of labels and conjugation techniques are known and are reported extensively in both the scientific and patent literature. Suitable labels include radionucleotides, enzymes, substrates, cofactors, inhibitors, fluorescent moieties, chemiluminescent moieties, magnetic particles, and the like.

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Protein Immunoassays

Means of detecting the proteins of the present invention are not critical aspects of the present invention. In a preferred embodiment, the proteins are detected and/or quantified using any of a number of well recognized immunological binding assays (see, e.g., U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168). For a review of the general immunoassays, see also Methods in Cell Biology, Vol. 37: Antibodies in Cell Biology, Asai, Ed., Academic Press, Inc. New York (1993); Basic and Clinical Immunology 7th Edition, Stites & Terr, Eds. (1991). Moreover, the immunoassays of the present invention can be performed in any of several configurations, e.g., those reviewed in Enzyme Immunoassay, Maggio, Ed., CRC Press, Boca Raton, Florida (1980); Tijan, Practice and Theory of Enzyme Immunoassays, Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers B.V., Amsterdam (1985); Harlow and Lane, above; Immunoassay: A Practical Guide, Chan, Ed., Academic Press, Orlando, FL (1987); Principles and Practice of Immunoassaysm, Price and Newman Eds., Stockton Press, NY (1991); and Non-isotopic Immunoassays, Ngo, Ed., Plenum Press, NY (1988). Immunological binding assays (or immunoassays) typically utilize a "capture agent" to specifically bind to and often immobilize the analyte (in this case, a protein of the present invention). The capture agent is a moiety that specifically binds to the analyte. In a preferred embodiment, the capture agent is an antibody that specifically binds a protein(s) of the present invention. The antibody may be produced by any of a number of means known to those of skill in the art as described herein.

Immunoassays also often utilize a labeling agent to specifically bind to and label the binding complex formed by the capture agent and the analyte. The labeling agent may itself be one of the moieties comprising the antibody/analyte complex. Thus, the labeling agent may be a labeled protein of the present invention or a labeled antibody specifically reactive to a protein of the present invention. Alternatively, the labeling agent may be a third moiety, such as another antibody, that specifically binds to the antibody/protein complex.

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In a preferred embodiment, the labeling agent is a second antibody bearing a label. Alternatively, the second antibody may lack a label, but it may, in turn, be bound by a labeled third antibody specific to antibodies of the species from which the second antibody is derived. The second can be modified with a detectable moiety, such as biotin, to which a third labeled molecule can specifically bind, such as enzyme-labeled streptavidin.

Other proteins capable of specifically binding immunoglobulin constant regions, such as protein A or protein G may also be used as the label agent. These proteins are normal constituents of the cell walls of streptococcal bacteria. They exhibit a strong non-immunogenic reactivity with immunoglobulin constant regions from a variety of species (See, generally Kronval, et al., J. Immunol. 111: 1401-1406 (1973), and Akerstrom, et al., J. Immunol. 135: 2589-2542 (1985)).

Throughout the assays, incubation and/or washing steps may be required after each combination of reagents. Incubation steps can vary from about 5 seconds to several hours, preferably from about 5 minutes to about 24 hours. However, the incubation time will depend upon the assay format, analyte, volume of solution, concentrations, and the like. Usually, the assays will be carried out at ambient temperature, although they can be conducted over a range of temperatures, such as 10°C to 40°C.

While the details of the immunoassays of the present invention may vary with the particular format employed, the method of detecting a protein of the present invention in a biological sample generally comprises the steps of contacting the biological sample with an antibody which specifically reacts, under immunologically reactive conditions, to a protein of the present invention. The antibody is allowed to bind to the protein under immunologically reactive conditions, and the presence of the bound antibody is detected directly or indirectly.

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A. Non-Competitive Assay Formats

Immunoassays for detecting proteins of the present invention include competitive and noncompetitive formats. Noncompetitive immunoassays are assays in which the amount of captured analyte (i.e., a protein of the present invention) is directly measured. In one preferred "sandwich" assay, for example, the capture agent (e.g., an antibody specifically reactive, under immunoreactive conditions, to a protein of the present invention) can be bound directly to a solid substrate where they are immobilized. These immobilized antibodies then capture the protein present in the test sample. The protein thus immobilized is then bound by a labeling agent, such as a second antibody bearing a label. Alternatively, the second antibody may lack a label, but it may, in turn, be bound by a labeled third antibody specific to antibodies of the species from which the second antibody is derived. The second can be modified with a detectable moiety, such as biotin, to which a third labeled molecule can specifically bind, such as enzyme-labeled streptavidin.

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B. Competitive Assay Formats

In competitive assays, the amount of analyte present in the sample is measured indirectly by measuring the amount of an added (exogenous) analyte (e.g., a protein of the present invention) displaced (or competed away) from a capture agent (e.g., an antibody specifically reactive, under immunoreactive conditions, to the protein) by the analyte present in the sample. In one competitive assay, a known amount of analyte is added to the sample and the sample is then contacted with a capture agent that specifically binds a protein of the present invention. The amount of protein bound to the capture agent is inversely proportional to the concentration of analyte present in the sample.

In a particularly preferred embodiment, the antibody is immobilized on a solid substrate. The amount of protein bound to the antibody may be determined either by measuring the amount of protein present in a protein/antibody complex, or alternatively by measuring the amount of remaining uncomplexed protein. The amount of protein may be detected by providing a labeled protein.

A hapten inhibition assay is another preferred competitive assay. In this assay a known analyte, (such as a protein of the present invention) is immobilized on a solid substrate. A known amount of antibody specifically reactive, under immunoreactive

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conditions, to the protein is added to the sample, and the sample is then contacted with the immobilized protein. In this case, the amount of antibody bound to the immobilized protein is inversely proportional to the amount of protein present in the sample. Again, the amount of immobilized antibody may be detected by detecting either the immobilized fraction of antibody or the fraction of the antibody that remains in solution. Detection may be direct where the antibody is labeled or indirect by the subsequent addition of a labeled moiety that specifically binds to the antibody as described above.

C. Generation of pooled antisera for use in immunoassays

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A protein that specifically binds to or that is specifically immunoreactive with an antibody generated against a defined immunogen is determined in an immunoassay. The immunoassay uses a polyclonal antiserum which is raised to a polypeptide of the present invention (i.e., the immunogenic polypeptide). This antiserum is selected to have low crossreactivity against other proteins and any such crossreactivity is removed by immunoabsorbtion prior to use in the immunoassay (e.g., by immunosorbtion of the antisera with a protein of different substrate specificity (e.g., a different enzyme) and/or a protein with the same substrate specificity but of a different form).

In order to produce antisera for use in an immunoassay, a polypeptide of the present invention is isolated as described herein. For example, recombinant protein can be produced in a mammalian or other eukaryotic cell line. An inbred strain of mice is immunized with the protein using a standard adjuvant, such as Freund's adjuvant, and a standard mouse immunization protocol (see Harlow and Lane, above). Alternatively, a synthetic polypeptide derived from the sequences disclosed herein and conjugated to a carrier protein is used as an immunogen. Polyclonal sera are collected and titered against the immunogenic polypeptide in an immunoassay, for example, a solid phase immunoassay with the immunogen immobilized on a solid support. Polyclonal antisera with a titer of 10⁴ or greater are selected and tested for their cross reactivity against polypeptides of different forms or substrate specificity, using a competitive binding immunoassay such as the one described in Harlow and Lane, above, at pages 570-573. Preferably, two or more distinct forms of polypeptides are used in this determination. These distinct types of polypeptides are used as competitors to identify antibodies which are specifically bound by the polypeptide being assayed for. The competitive

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polypeptides can be produced as recombinant proteins and isolated using standard molecular biology and protein chemistry techniques as described herein.

Immunoassays in the competitive binding format are used for crossreactivity determinations. For example, the immunogenic polypeptide is immobilized to a solid support. Proteins added to the assay compete with the binding of the antisera to the immobilized antigen. The ability of the above proteins to compete with the binding of the antisera to the immobilized protein is compared to the immunogenic polypeptide. The percent crossreactivity for the above proteins is calculated, using standard calculations. Those antisera with less than 10% crossreactivity with a distinct form of a polypeptide are selected and pooled. The cross-reacting antibodies are then removed from the pooled antisera by immunoabsorbtion with a distinct form of a polypeptide.

The immunoabsorbed and pooled antisera are then used in a competitive binding immunoassay as described herein to compare a second "target" polypeptide to the immunogenic polypeptide. In order to make this comparison, the two polypeptides are each assayed at a wide range of concentrations and the amount of each polypeptide required to inhibit 50% of the binding of the antisera to the immobilized protein is determined using standard techniques. If the amount of the target polypeptide required is less than twice the amount of the immunogenic polypeptide that is required, then the target polypeptide is said to specifically bind to an antibody generated to the immunogenic protein. As a final determination of specificity, the pooled antisera is fully immunosorbed with the immunogenic polypeptide until no binding to the polypeptide used in the immunosorbtion is detectable. The fully immunosorbed antisera is then tested for reactivity with the test polypeptide. If no reactivity is observed, then the test polypeptide is specifically bound by the antisera elicited by the immunogenic protein.

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D. Other Assay Formats

In a particularly preferred embodiment, Western blot (immunoblot) analysis is used to detect and quantify the presence of protein of the present invention in the sample. The technique generally comprises separating sample proteins by gel electrophoresis on the basis of molecular weight, transferring the separated proteins to a suitable solid support, (such as a nitrocellulose filter, a nylon filter, or derivatized nylon filter), and incubating the sample with the antibodies that specifically bind a protein of the present invention. The antibodies specifically bind to the protein on the solid support. These

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antibodies may be directly labeled or alternatively may be subsequently detected using labeled antibodies (e.g., labeled sheep anti-mouse antibodies) that specifically bind to the antibodies.

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E. Quantification of Proteins.

The proteins of the present invention may be detected and quantified by any of a number of means well known to those of skill in the art. These include analytic biochemical methods such as electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), hyperdiffusion chromatography, and the like, and various immunological methods such as fluid or gel precipitin reactions, immunodiffusion (single or double), immunoelectrophoresis, radioimmunoassays (RIAs), enzyme-linked immunosorbent assays (ELISAs), immunofluorescent assays, and the like.

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F. Reduction of Non-Specific Binding

One of skill will appreciate that it is often desirable to reduce non-specific binding in immunoassays and during analyte purification. Where the assay involves an antigen, antibody, or other capture agent immobilized on a solid substrate, it is desirable to minimize the amount of non-specific binding to the substrate. Means of reducing such non-specific binding are well known to those of skill in the art. Typically, this involves coating the substrate with a proteinaceous composition. In particular, protein compositions such as bovine serum albumin (BSA), nonfat powdered milk, and gelatin are widely used.

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G. Immunoassay Labels

The labeling agent can be, e.g., a monoclonal antibody, a polyclonal antibody, a binding protein or complex, or a polymer such as an affinity matrix, carbohydrate or lipid. Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, radioisotopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Detection may proceed by any known method, such as immunoblotting, western analysis, gel-mobility shift assays, fluorescent in situ hybridization analysis (FISH), tracking of radioactive or

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bioluminescent markers, nuclear magnetic resonance, electron paramagnetic resonance, stopped-flow spectroscopy, column chromatography, capillary electrophoresis, or other methods which track a molecule based upon an alteration in size and/or charge. The particular label or detectable group used in the assay is not a critical aspect of the invention. The detectable group can be any material having a detectable physical or chemical property. Such detectable labels have been well-developed in the field of immunoassays and, in general, any label useful in such methods can be applied to the present invention. Thus, a label is any composition detectable by spectroscopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Useful labels in the present invention include magnetic beads, fluorescent dyes, radiolabels, enzymes, and colorimetric labels or colored glass or plastic beads, as discussed for nucleic acid labels, above.

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The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. As indicated above, a wide variety of labels may be used, with the choice of label depending on the sensitivity required, ease of conjugation of the compound, stability requirements, available instrumentation, and disposal provisions.

Non-radioactive labels are often attached by indirect means. Generally, a ligand molecule (e.g., biotin) is covalently bound to the molecule. The ligand then binds to an anti-ligand (e.g., streptavidin) molecule which is either inherently detectable or covalently bound to a signal system, such as a detectable enzyme, a fluorescent compound, or a chemiluminescent compound. A number of ligands and anti-ligands can be used. Where a ligand has a natural anti-ligand, for example, biotin, thyroxine, and cortisol, it can be used in conjunction with the labeled, naturally occurring anti-ligands. Alternatively, any haptenic or antigenic compound can be used in combination with an antibody.

The molecules can also be conjugated directly to signal generating compounds, e.g., by conjugation with an enzyme or fluorophore. Enzymes of interest as labels will primarily be hydrolases, particularly phosphatases, esterases and glycosidases, or oxidoreductases, particularly peroxidases. Fluorescent compounds include fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, etc.

Chemiluminescent compounds include luciferin, and 2,3-dihydrophthalazinediones, e.g.,

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luminol. For a review of various labeling or signal producing systems which may be used, see, U.S. Patent No. 4,391,904, which is incorporated herein by reference.

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Means of detecting labels are well known to those of skill in the art. Thus, for example, where the label is a radioactive label, means for detection include a scintillation counter or photographic film as in autoradiography. Where the label is a fluorescent label, it may be detected by exciting the fluorochrome with the appropriate wavelength of light and detecting the resulting fluorescence, e.g., by microscopy, visual inspection, via photographic film, by the use of electronic detectors such as charge coupled devices (CCDs) or photomultipliers and the like. Similarly, enzymatic labels may be detected by providing appropriate substrates for the enzyme and detecting the resulting reaction product. Finally, simple colorimetric labels may be detected simply by observing the color associated with the label. Thus, in various dipstick assays, conjugated gold often appears pink, while various conjugated beads appear the color of the bead.

Some assay formats do not require the use of labeled components. For instance, agglutination assays can be used to detect the presence of the target antibodies. In this case, antigen-coated particles are agglutinated by samples comprising the target antibodies. In this format, none of the components need be labeled and the presence of the target antibody is detected by simple visual inspection.

20 Assays for Compounds that Modulate Enzymatic Activity or Expression

The present invention also provides means for identifying compounds that bind to (e.g., substrates), and/or increase or decrease (i.e., modulate) the enzymatic activity of, catalytically active polypeptides of the present invention. The method comprises contacting a polypeptide of the present invention with a compound whose ability to bind to or modulate enzyme activity is to be determined. The polypeptide employed will have at least 20%, preferably at least 30% or 40%, more preferably at least 50% or 60%, and most preferably at least 70% or 80% of the specific activity of the native, full-length polypeptide of the present invention (e.g., enzyme). Generally, the polypeptide will be present in a range sufficient to determine the effect of the compound, typically about 1 nM to 10 µM. Likewise, the compound will be present in a concentration of from about 1 nM to 10 µM. Those of skill will understand that such factors as enzyme concentration, ligand concentrations (i.e., substrates, products, inhibitors, activators), pH, ionic strength, and temperature will be controlled so as to obtain useful kinetic data

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and determine the presence of absence of a compound that binds or modulates polypeptide activity. Methods of measuring enzyme kinetics is well known in the art. See, e.g., Segel, *Biochemical Calculations*, 2nd ed., John Wiley and Sons, New York (1976).

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Although the present invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

10 Example 1

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This example describes the construction cDNA libraries.

Total RNA Isolation

Total RNA was isolated from corn tissues with TRIzol Reagent (Life Technology Inc. Gaithersburg, MD) using a modification of the guanidine isothiocyanate/acid-phenol procedure described by Chomczynski and Sacchi (Chomczynski, P., and Sacchi, N. Anal. Biochem. 162, 156 (1987)). In brief, plant tissue samples were pulverized in liquid nitrogen before the addition of the TRIzol Reagent, and then were further homogenized with a mortar and pestle. Addition of chloroform followed by centrifugation was conducted for separation of an aqueous phase and an organic phase. The total RNA was recovered by precipitation with isopropyl alcohol from the aqueous phase.

Poly(A) + RNA Isolation

The selection of poly(A)+ RNA from total RNA was performed using PolyATact system (Promega Corporation. Madison, WI). In brief, biotinylated oligo(dT) primers were used to hybridize to the 3' poly(A) tails on mRNA. The hybrids were captured using streptavidin coupled to paramagnetic particles and a magnetic separation stand. The mRNA was washed at high stringent condition and eluted by RNase-free deionized water.

cDNA Library Construction

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cDNA synthesis was performed and unidirectional cDNA libraries were constructed using the SuperScript Plasmid System (Life Technology Inc. Gaithersburg, MD). The first stand of cDNA was synthesized by priming an oligo(dT) primer containing a Not I site. The reaction was catalyzed by SuperScript Reverse Transcriptase II at 45°C. The second strand of cDNA was labeled with alpha-³²P-dCTP and a portion of the reaction was analyzed by agarose gel electrophoresis to determine cDNA sizes. cDNA molecules smaller than 500 base pairs and unligated adapters were removed by Sephacryl-S400 chromatography. The selected cDNA molecules were ligated into pSPORT1 vector in between of Not I and Sal I sites.

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Example 2

This example describes cDNA sequencing and library subtraction.

Sequencing Template Preparation

Individual colonies were picked and DNA was prepared either by PCR with M13 forward primers and M13 reverse primers, or by plasmid isolation. All the cDNA clones were sequenced using M13 reverse primers.

O-bot Subtraction Procedure

cDNA libraries subjected to the subtraction procedure were plated out on 22 x 22 cm² agar plate at density of about 3,000 colonies per plate. The plates were incubated in a 37°C incubator for 12-24 hours. Colonies were picked into 384-well plates by a robot colony picker, Q-bot (GENETIX Limited). These plates were incubated overnight at 37°C.

Once sufficient colonies were picked, they were pinned onto 22 x 22 cm² nylon membranes using Q-bot. Each membrane contained 9,216 colonies or 36,864 colonies. These membranes were placed onto agar plate with appropriate antibiotic. The plates were incubated at 37°C for overnight.

After colonies were recovered on the second day, these filters were placed on filter paper prewetted with denaturing solution for four minutes, then were incubated on top of a boiling water bath for additional four minutes. The filters were then placed on filter paper prewetted with neutralizing solution for four minutes. After excess solution was removed by placing the filters on dry filter papers for one minute, the colony side of the filters were place into Proteinase K solution, incubated at 37°C for 40-50 minutes.

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The filters were placed on dry filter papers to dry overnight. DNA was then crosslinked to nylon membrane by UV light treatment.

Colony hybridization was conducted as described by Sambrook, J., Fritsch, E.F. and Maniatis, T., (in Molecular Cloning: A laboratory Manual, 2nd Edition). The following probes were used in colony hybridization:

- First strand cDNA from the same tissue as the library was made from to remove the most redundant clones.
- 2. 48-192 most redundant cDNA clones from the same library based on previous sequencing data.
- 10 3. 192 most redundant cDNA clones in the entire corn partial sequence database.
 - A Sal-A20 oligo nucleotide: TCG ACC CAC GCG TCC GAA AAA AAA AAA AAA AAA AAA AAA, removes clones containing a poly A tail but no cDNA.
 - 5. cDNA clones derived from rRNA.

The image of the autoradiography was scanned into computer and the signal intensity and cold colony addresses of each colony was analyzed. Re-arraying of cold-colonies from 384 well plates to 96 well plates was conducted using Q-bot.

Example 3

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This example describes identification of the gene from a computer homology 20 search. Gene identities were determined by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) J. Mol. Biol. 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches under default parameters for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure 25 Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm. The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the 30 "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. (1993) Nature Genetics 3:266-272) provided by the NCBI. In some cases, the sequencing data from two or more clones containing overlapping segments of DNA were used to construct contiguous DNA sequences.

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The above examples are provided to illustrate the invention but not to limit its scope. Other variants of the invention will be readily apparent to one of ordinary skill in the art and are encompassed by the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference.

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WHAT IS CLAIMED IS:

- 1. An isolated nucleic acid comprising a member selected from the group consisting of:
- 5 (a) a polynucleotide having at least 80% sequence identity, as determined by the BLAST 2.0 algorithm under default parameters, to a polynucleotide encoding a polypeptide selected from the group consisting of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
 - (b) a polynucleotide encoding a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
 - (c) a polynucleotide amplified from a Zea mays nucleic acid library using primers which selectively hybridize, under stringent hybridization conditions, to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
- 15 (d) a polynucleotide which selectively hybridizes, under stringent hybridization conditions and a wash in 2X SSC at 50°C, to a polynucleotide of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
 - (e) a polynucleotide of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57
- 20 (f) a polynucleotide which is complementary to a polynucleotide of (a), (b), (c), (d), or (e); and
 - (g) a polynucleotide comprising at least 25 contiguous nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f).
- 25 2. A recombinant expression cassette, comprising a member of claim 1 operably linked, in sense or anti-sense orientation, to a promoter.
 - 3. A host cell comprising the recombinant expression cassette of claim 2.
- 30 4. A transgenic plant comprising a recombinant expression cassette of claim 2.
 - 5. The transgenic plant of claim 4, wherein the plant is a monocot.

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- 6. The transgenic plant of claim 4, wherein the plant is selected from the group consisting of: maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, and millet.
- 5 7. A transgenic seed from the transgenic plant of claim 4.
 - 8. A method of modulating the level of cellulose synthase in a plant cell capable of plant regeneration, comprising:
 - (a) transforming the plant cell with a recombinant expression cassette comprising a cellulose synthase polynucleotide of claim 1 operably linked to a promoter;
 - (b) culturing the transformed plant cell; and
 - (c) inducing expression of said polynucleotide for a time sufficient to modulate the level of cellulose synthase in said transformed plant cell.
- 15 9. The method of claim 8, wherein a plant is regenerated from the transformed plant cell.
- 10. The method of claim 9, wherein the plant is selected from the group consisting of : maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, and20 millet.
 - 11. The method of claim 8, wherein the promoter is a tissue-preferred promoter.
 - 12. The method of claim 8, wherein the level of cellulose synthase is increased.

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13. The method of claim 8 wherein the cell cycle polynucleotide is amplified from a Zea mays nucleic acid library using primers which selectively hybridize, under stringent hybridization conditions, to loci within a polynucleotide selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57.

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14. The method of claim 8 wherein the cell cycle gene is selected from the group consisting of SEQ ID NOS: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57.

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- 15. An isolated protein comprising a member selected from the group consisting of:
 - (a) a polypeptide of at least 20 contiguous amino acids from a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
 - (b) a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58
 - (c) a polypeptide having at least 80% sequence identity to, and having at least one linear epitope in common with, a polypeptide of SEQ ID NOS: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, and 58, wherein said sequence identity is determined using BLAST 2.0 under default parameters; and,
- 10 (d) a polypeptide encoded by a member of claim 1.

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- 1 -

SEQUENCE LISTING

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<151> August 17, 1998

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50 55 60	
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105

- 2 -

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_ 4 _

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- 6 -													
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His	Met	Met	Ser 170	Pro	Thr	Gly	Asn	Ile 175	Gly	Lys	Arg	Ala	Pro 180	Phe	Pro	
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					aaa Lys								-	-	-	979
_	_		-		ccc Pro 220	_	_				-		-			1027
					ggt Gly											1075
					aac Asn											1123
_					tcc Ser							_	_			1171
_	_	_		_	cta Leu	_			-			_	_			1219
		_		_	tac Tyr 300							-		-		1267
					tcg Ser											1315
					acg Thr			-								1363
					tct Ser									_		1411
					aag Lys											1459
					gtg Val 380											1507
gta Val	tct Ser	gat Asp	gat Asp	gga Gly 395	gct Ala	gcg Ala	atg Met	ctg Leu	aca Thr 400	ttt Phe	gat Asp	gca Ala	cta Leu	gct Ala 405	gag Glu	1555

		gag Glu		_	-			_			-	_	_			1603
	-	cct Pro 425	-	-		-					-			_		1651
-	_	gac Asp							_		_	_		-	_	1699
		gaa Glu														1747
		cag Gln														1795
		cca Pro						_				-		_	· .	1843
		ggt Gly 505														1891
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		gct Ala														1987
		gga Gly														2035
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Thr G	ly Cy	s Val	Phe 635	Asn	Arg	Thr	Ala	Leu 640	туг	Gly	Tyr	Glu	Pro 645	Pro	
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aag a Lys L															2371
aag c Lys H 6															2419
gag g Glu G 695															2467
tct c															2515
gcc to Ala So															2563
gag to Glu So		u Leu													2611
gac as Asp Ly 70						_									2659
aca ga Thr Gi 775							-	_			-				2707
tcg at Ser I		_	_		_			-		_			-		2755
atc as															2803
tcc gt Ser Va		u Ile			_			_		-					2851
gga gg Gly Gl															2899
atc ta Ile Ty 855			-			-									2947

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•			gac ggc gac ttc Asp Gly Asp Phe	
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acc cgc gtc act Thr Arg Val Thr 1065				t 3566
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- 15 -

Phe Val Lys Lys Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Ser Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val His Pro Ser Phe Val Lys Asp Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Val Asn Gly Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Thr Xaa Asp His Pro Gly Met Ile Gln Val Phe Leu Gly His Ser Gly Gly Leu Asp Thr Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg Val Ser Ala Val Leu Thr Asn Gly Gln Tyr Met Leu Asn Leu Asp Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Leu Arg Glu Ala Met Cys Phe Leu Met Asp Pro Asn Leu Gly Arg Ser Val Cys Tyr Val Gln Phe 580 585 Pro Gln Arg Phe Asp Gly Ile Asp Arg Asn Asp Arg Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn Leu Arg Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn Arg Thr Ala Leu Tyr Gly Tyr Glu Pro Pro Ile Lys Gln Lys Lys Gly Gly Phe Leu Ser Ser Leu Cys Gly Gly Arg Lys Lys Ala Ser Lys Ser Lys Gly Ser Asp Lys Lys Ser Gln Lys His Val Asp Ser Ser Val Pro Val Phe Asn Leu Glu Asp Ile Glu Glu Gly Val Glu Gly Ala Gly Phe Asp Asp 690 695 Glu Lys Ser Leu Leu Met Ser Gln Met Ser Leu Glu Lys Arg Phe Gly Gln Ser Ala Ala Phe Val Ala Ser Thr Leu Met Glu Tyr Gly Gly Val Pro Gln Ser Ala Thr Pro Glu Ser Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp Gly Thr Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Ala Arg Gly Trp Arg Ser Ile Tyr Cys Met Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val 805 810 815 Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu Phe Ser Arg His Cys Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys Phe Leu Glu Arg Phe Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr Ser Ile Pro Leu Leu Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile

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120

1.80

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cag ata tct gga gag att cct Gln Ile Ser Gly Glu Ile Pro 160		· · · · · · · · · · · · · · · · ·	: Ile								
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gtt gac tgg aag gaa aga gtt Val Asp Trp Lys Glu Arg Val 205 210											
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	cta Leu										_					1145
	gat Asp	_									_					1193
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	gtt Val	_				-				-	_			_		1385
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	ttt Phe	_				-		-	_							1529
	gtt Val 445	_	-	-	-		-	-				_	-			1577

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	tgg Trp							_								1673
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_	act Thr	-						-		_		_		_		1769
	aga Arg 525															1817
	att Ile									_						1865
	gat Asp	-	-					_	_		-		_	-	-	1913
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	ttt Phe															2009
	cgg Arg 605			_			_			_			_	_		2057
	cag Gln			_						_	_					2105
	ttg Leu															2153
	att Ile													_	_	2201
	tat Tyr															2249
gct Ala	ccc Pro	atc Ile	ttc Phe	aat Asn	atg Met	gaa Glu	gac Asp	atc Ile	gaa Glu	gag Glu	ggt Gly	att Ile	gaa Glu	ggt Gly	tac Tyr	2297

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		cca Pro						_				_	-	-		2441
	_	atc Ile 750	_	_							-					2489
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		att Ile				-										2873
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		ggc Gly 910														2969

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-	cct ccg acc Pro Pro Thr 975	•	Val Ile			3161
	att tct tat Ile Ser Tyr 0	•				3209
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	ctc aag ggt Leu Lys Gly 102	Leu Met Gly		Asn Arg Thr		3305
	gtc tgg tcc Val Trp Ser 1040					3353
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Phe Pr 65	o Val	Cys	Arg	Pro 70	Cys	Tyr	Glu	Tyr	Glu 75	Arg	Lys	Glu	Gly	Asn 80
Gln Cy			85	_	_		_	90	_	_		_	95	
Pro Ar	g Val	His 100	Gly	Asp	Glu	Asp	Glu 105	Glu	Asp	Val	Asp	Asp 110	Leu	Asp
Asn Gl	115		-	_		120		_	-	_	125		_	
Leu Gl	0		_		135					140				
His Hi 145	_			150				_	155				_	160
Ile Pr	_		165		-	_		170		_			175	
Ser Ty		180					185			_		190	_	
Ser Ly	195					200					205			
Arg Va 21	0			_	215	-		-	_	220				
Thr As:				230					235					240
Ser As			245					250	_		_		255	_
Ser Ar	_	260					265					270		
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Arg Ty	_		325		_			330		_	_		335	
Phe Va		340					345					350		_
Asn Th	355			_		360	_				365			
37 Ser Cy	כ				375			_	-	380				
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Lys Hi			405					410					415	
Ile As		420					425					430		
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465				470					475					480
Asp G1			485					490					495	
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cgccggcctc gtcggtgtcg gtggagtgtg aatcggtgtg tgtaggagga gcgcggag
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atg gcg gcc aac aag ggg atg gtg gca ggc tct cac aac cgc aac gag
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Met Ala Ala Asn Lys Gly Met Val Ala Gly Ser His Asn Arg Asn Glu
                                                                 274
tte gte atg ate ege eae gae gge gae geg eet gte eeg get aag eee
Phe Val Met Ile Arg His Asp Gly Asp Ala Pro Val Pro Ala Lys Pro
acg aag agt gcg aat ggg cag gtc tgc cag att tgt ggc gac act gtt
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Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Thr Val
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						gat Asp 55										370
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						aag Lys										466
						gat Asp										514
						caa Gln										562
						gat Asp 135					_	_		-		610
					-	ctt Leu		-			_					658
						gac Asp										706
_		_	_		-	gtt Val		-						_		754
_	_	-	_			tat Tyr				_	_	-		_	-	802
	_					gtt Val 215		_	_			-	_		_	850
						gct Ala										898
		_	_	_		atg Met	-	_		-	_					946
						tca Ser										994
atc	att	ctc	cgt	ctt	atc	atc	ctg	tgc	ttc	ttc	ttc	caa	tat	cgt	atc	1042

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Ile	Ile	Leu 275	Arg	Leu	Ile	Ile	Leu 280	Cys	Phe	Phe	Phe	Gln 285	Tyr	Arg	Ile	
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			tgg Trp													1138
_			atc Ile			-				_			_	_		1186
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	-	_	tcc Ser			_	-	-			_					1330
			tct Ser			_										1378
	_	_	gcc Ala													1426
			gaa Glu 420		-	-		-				_				1474
-		_	aag Lys	-							-	_	-	_	-	1522
-	_	_	aga Arg			_	-				-			_		1570
_	-		gca Ala	_				_					_	-	_	1618
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_			ttg Leu 500			_				-		-				1714

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	cac His 530															1810
	ctg Leu															1858
	aat Asn									_	_		_	_	-	1906
	gct Ala			_			-		-					_		1954
	ggc Gly	_	_	_		-	_		-					_		2002
	gat Asp 610															2050
	gga Gly			-					_	-	_				_	2098
	gtt Val	_				_	_	-					_	_	_	2146
	tgt Cys				_							_				2194
-	cgt Arg		_		-		_			-					-	2242
	gac Asp 690															2290
	atg Met															2338
	att Ile	_				_										2386
aac	cca	gct	tct	cta	ctg	aag	gaa	gct	atc	cat	gtt	atc	agc	tgt	a aa	2434

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Asn	Pro	Ala	Ser 740	Leu	Leu	Lys	Glu	Ala 745	Ile	His	Val	Ile	Ser 750	Cys	Gly	
		gac Asp 755														2482
		aca Thr		-		_					_		-	_		2530
		tca Ser			_	_			_		_		•			2578
_		atc Ile				_	_			_			_		_	2626
		tca Ser				_		-	_		_					2674
		aat Asn 835	_													2722
		gtt Val						_	_			_		_		2770
		gct Ala														2818
		tat Tyr	_		_							-				2866
_		ggt Gly					-		_		_					2914
		aga Arg 915			_			_						-		2962
		gcg Ala										Ala				3010
		ttc Phe		-			_	-		_				-	ttt Phe 960	3058
		cta Leu														3106

act gtt ctt gtc att aac ctg gtc gga atg gtg gca gga att tcg tat Thr Val Leu Val Ile Asn Leu Val Gly Met Val Ala Gly Ile Ser Tyr 980 985 990	3154
gcc att aac agc ggc tac caa tcc tgg ggt ccg ctc ttt gga aag ctg Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys Leu 995 1000 1005	3202
ttc ttc tcg atc tgg gtg atc ctc cat ctc tac ccc ttc ctc aag ggt Phe Phe Ser Ile Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly 1010 1015 1020	3250
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Ile	Pro	Asp	Ala	Ser	Pro	Asp	Arg	His	Ser	Ile	Arg	Ser	Pro	Thr	Ser
	_		_	165	_		_		170		_			175	_
Ser	Tyr	Val	-	Pro	Ser	Val	Pro		Pro	Val	Arg	Ile		Asp	Pro
Ser	Lys	Asn	180	Asn	Ser	ጥህተ	Glv	185	Asn	Ser	Val	Asp.	190	Lvs	Glu
001	2,0	195				-7-	200	200				205		_,,	
Arg	Val	Glu	Ser	Trp	Arg	Val	Lys	Gln	Asp	Lys	Asn	Met	Leu	Gln	Val
	210					215			_		220				_
Thr 225	Asn	Lys	Tyr	Pro	Glu 230	Ala	Arg	Gly	Asp	Met 235	Glu	Gly	Thr	Gly	Ser 240
	Gly	Glu	Asp	Met		Met	Val	Asp	Asp		Arq	Leu	Pro	Leu	
	2			245					250					255	
Arg	Ile	Val		Ile	Ser	Ser	Asn		Leu	Asn	Leu	Tyr	_	Ile	Val
-1.	- 1 -	7	260	• • • •	-1 -	71 -	•	265	Dh a	nh -	Dh.a	~ 1	270	3	- 1-
TTE	Ile	275	Arg	Leu	TIE	ire	280	Cys	Pne	Pne	Pne	285	Tyr	Arg	TTE
Ser	His		Val	Arq	Asn	Ala		Gly	Leu	Trp	Leu		Ser	Val	Ile
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Tip	Tyr	PIO	TTG	325	Arg	GIU	Int	TÀT	330	Asp	Arg	пеα	ALG	335	
Tyr	Asp	Arg	Glu		Glu	Pro	Ser	Gln		Ala	Pro	Ile	Asp		
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Val	Ser		Val	Asp	Pro	Leu	-	Glu	Pro	Pro	Leu		Thr	Ala	Asn
Thr	Val	355 Leu	Ser	Tle	Leu	Ala	360 Val	Asp	Tvr	Pro	Val	365 Asp	Lvs	Val	Ser
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Cys	Tyr	Val	Ser	Asp	Asp	Gly	Ser	Ala	Met		Thr	Phe	Glu	Ser	
385	63	m		03	390	3 3-	3	T	·	395	D	Dh a	C	T	400
Ser	Glu	Thr	Ата	405	Pne	ATa	Arg	ьys	410	Val	Pro	Pne	Cys	LуS 415	nys
His	Asn	Ile	Glu		Arg	Ala	Pro	Glu		Tyr	Phe	Ala	Gln		Ile
			420					425					430		
Asp	Tyr		Lys	Asp	Lys	Ile		Pro	Ser	Phe	Val		Glu	Arg	Arg
Δla	Met	435	Ara	Glu	Tvr	Glu	440 Glu	Phe	Lvs	Tle	Ara	445 Tle	Asn	Ala	Leu
•	450	-3-	9		-1-	455			-1-		460				
Val	Ala	Lys	Ala	Gln	Lys	Val	Pro	Glu	Glu	-	Trp	Thr	Met	Ala	
465	-1			5	470		•	B		475	***	D	~1	Mah	480
GIY	Thr	ALA	Trp	485	GIY	Asn	Asn	Pro	490	Asp	nis	PIO	GIY	495	116
Gln	Val	Phe	Leu		His	Ser	Gly	Gly		Asp	Thr	Asp	Gly		Glu
			500	_			_	505					510		
Leu	Pro		Leu	Val	Tyr	Val		Arg	Glu	Lys	Arg		Gly	Phe	Gln
tri m	***	515	T	21-	~1··	71-	520 Mat	7.55	77 7	f ou	T10	525	Val	Sar	Δla
nis	His 530	Lys	rys	Ala	GIA	535	met	ASII	Ala	neu	540	ALG	Val	Ser	n.u
Val	Leu	Thr	Asn	Gly	Ala		Leu	Leu	Asn	Val		Cys	Asp	His	Tyr
545				_	550	_				555					560
Phe	Asn	Ser	Ser		Ala	Leu	Arg	Glu		Met	CÀa	Phe	Met		Asp
D~^	Ala	T.e.v	Glv	565 Ara	Lve	Thr	Cve	ጥኒንም	570 Val	Gln	Phe	Pro	G) n	575 Ara	Phe
FIO	A1a	Deu	580	~-3	Lys	7 - 1 L	Cys	585					590	3	
Asp	Gly	Ile	Asp	Leu	His	Asp	Arg	Tyr	Ala	Asn	Arg	Asn	Ile	Val	Phe

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59	=				600					605			
Phe Asp Il		Met I	Lys	Gly 615	600 Leu	Asp	Gly	Ile	Gln 620	605 Gly	Pro	Val	Tyr
Val Gly Th	r Gly		Cys 630		Asn	Arg	Gln	Ala 635		Tyr	Gly	Tyr	Asp 640
Pro Val Le	a Thr	Glu <i>1</i> 645	Ala	Asp	Leu	Glu	Pro 650	Asn	Ile	Val	Val	Lys 655	Ser
Cys Cys Gl	Arg 660	Arg I	Lys	Arg	Lys	Asn 665	Lys	Ser	Tyr	Met	Asp 670	Ser	Gln
Ser Arg Il 67	5	-	_		680					685			
Glu Asp Il 690			_	695		_	_		700		-		
Leu Met Se 705		7	710		•	-	_	715	_				720
Phe Ile Al		725					730					735	
Asn Pro Al	740			-		745					750	-	-
Tyr Glu As	5			_	760	_			_	765		_	_
Ser Val Th				775					780				
Trp Gln Se		7	790				_	795	-		-	-	008
Ala Pro Il		805		_	_		810				_	815	
Gly Tyr As:	820					825					830		
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850 Leu Pro Al				855					860		_	_	
865 Ser Asn Ty			370				_	875					880
Ala Thr Gl		885					890					895	
Trp Trp Are	900				_	905		_		-	910		
91 Leu Phe Ala	5				920			-	-	925			
930 Thr Asn Pho				935			-		940		_		
945		9	950	_	_		_	955	_				960
Ala Glu Le	_	965					970					975	
	980					985				-	990		_
Ala Ile Ass	5	_			1000)				1005	;		
Phe Phe Set				1015	;				1020)			
Leu Met Gl	Arg			-	TUY	PTO	rnr			тте	val	rrp	1040
1025	. 23-		1030		C	T 655	T 0	1035		T	T1^	700	
Ile Leu Le		ser 1	тте	Fire	Ser	neu	1050		val	nys	TTE	1055	
Phe Ile Se			31 n	Laze	Δla	Ala			Glv	Gln	Cve		
the tre se	. FIO	1111		-ya	vra	n_a	-1 Cl	سا ب	O-Y	3111	CYS	1	

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geggaagtgg aggggaggaa geg atg gag geg age gee ggg etg gtg gee gge
                                                                      173
                           Met Glu Ala Ser Ala Gly Leu Val Ala Gly
tcc cac aac cgc aac gag ctc gtc gtc atc cgc cgc gac ggc gat ccc
                                                                      221
Ser His Asn Arg Asn Glu Leu Val Val Ile Arg Arg Asp Gly Asp Pro
                                                                      269
ggg ccg aag ccg ccg cgg gag cag aac ggg cag gtg tgc cag att tgc
Gly Pro Lys Pro Pro Arg Glu Gln Asn Gly Gln Val Cys Gln Ile Cys
                                                                      317
ggc gac gac gtc ggc ctt gcc ccc ggc ggg gac ccc ttc gtg gcg tgc
Gly Asp Asp Val Gly Leu Ala Pro Gly Gly Asp Pro Phe Val Ala Cys
                                                                      365
aac gag tgc gcc ttc ccc gtc tgc cgg gac tgc tac gaa tac gag cgc
Asn Glu Cys Ala Phe Pro Val Cys Arg Asp Cys Tyr Glu Tyr Glu Arg
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                          65
cgg gag ggc acg cag aac tgc ccc cag tgc aag act cga tac aag cgc
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Arg Glu Gly Thr Gln Asn Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg
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	-	gag Glu 125		_					_	-			_			557
_		aat Asn					-		_					-		605
		acc Thr				-		_	-			_		_		653
	_	gtg Val				_					_					701
		tat Tyr		_		_								_	_	749
		aag Lys 205	_		_	_					_				_	797
_		atg Met	_				_				_			_		845
		gat Asp														893
_	-	gaa Glu	_	_			_									941
		att Ile														989
		ttc Phe 285														1037
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tct Ser	tgg Trp	att Ile	ctt Leu	gat Asp	caa Gln	ttc Phe	cca Pro	aag Lys	tgg Trp	ttc Phe	cct Pro	att Ile	gag Glu	aga Arg	gag Glu	1133

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	gaa Glu															1277
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_	gca Ala	-		_		-	-			_			-			1373
-	aaa Lys		_			-						_		_	-	1421
	gag Glu															1469
_	gca Ala			_				-		_						1517
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	gaa Glu	_				_		-	_							1613
	gtt Val															1661
	ggc															1709
tct Ser	aga Arg	gag Glu 525	aaa Lys	cga Arg	cca Pro	ggc Gly	tat Tyr 530	aac Asn	cat His	cat His	aag Lys	aaa Lys 535	gct Ala	ggt Gly	gct Ala	1757
atg Met	aat Asn 540	gca Ala	ttg Leu	gtc Val	cga Arg	gtc Val 545	tct Ser	gct Ala	gta Val	cta Leu	aca Thr 550	aat Asn	gct Ala	cca Pro	tat Tyr	1805

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				atg Met													1901
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	-	-		att Ile	_					_				_	_		2045
A	-			gca Ala						-				_	_		2093
				act Thr	_		_			_		_		_	_	-	2141
				aat Asn 670		_		_	_							_	2189
	-		_	tta Leu				_		-						-	2237
		-		ggt Gly	_		-	-	_			-					2285
L	_	_		att Ile	-						-	_					2333
			_	ttt Phe	-												2381
				agt Ser 750													2429
				tat Tyr													2477
				tca Ser													2525

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	780					785				790					
tgt (Cys) 795															2573
aaa (2621
cgg (_			-		2669
ctt (Leu :										_	-	_			2717
tac a													_	_	2765
tac 1 Tyr (875															2813
cca (_	_			_	_	-	_		_					2861
tgc a															2909
att o	_	_						_		 _					2957
tcc t Ser s															3005
ggt g Gly V 955															3053
gag t Glu I			-									-			3101
cct a Pro T			_	Leu		_			Ile	 	_	-	Gly	-	3149
tca a Ser A	Asn		Ile					Glu		 		Leu			3197

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PCT/US99/18760

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Gln Cys Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Gln Lys Gly Ser

Pro Arg Val His Gly Asp Asp Glu Glu Glu Asp Val Asp Asp Leu Asp

105

514

85

100

- 41 -

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tgc tgt ggt Cys Cys Gly						_	2194
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gct gag cta tat go Ala Glu Leu Tyr Va 90	7		_	
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- 45 -

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	atc Ile		_	Thr	-		_	-	Ala	_			-	Gly		3394
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Pro	Arg	Val	His 100	Gly	Asp	Asp	Glu	Glu 105	Glu	Asp	Val	Asp	Asp 110	Leu	Asp	
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260 265 270

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Thr	Val 370		Ser	Ile	Leu	Ala 375	Val	Asp	Tyr	Pro	Val 380	Asp	Lys	Val	Ser
Cys 385	Tyr	Val	Ser	Asp	Asp 390		Ser	Ala	Met	Leu 395		Phe	Glu	Ser	Leu 400
	Glu	Thr	Ala	Glu 405		Ala	Arg	Lys	Trp 410		Pro	Phe	Суз	Lys 415	
His	Asn	Ile	Glu 420		Arg	Ala	Pro	Glu 425		Tyr	Phe	Ala	Gln 430		Ile
Asp	Tyr	Leu 435		Asp	Lys	Ile	Gln 440		Ser	Phe	Val	Lys 445		Arg	Arg
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His	His 530	Lys	Lys	Aļa	Gly	Ala 535	Met	Asn	Ala	Leu	Ile 540	Arg	Val	Ser	Ala
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Trp Gln Ser Ile Tyr Cys Met Pro Pro Arg Pro Cys Phe Lys Gly Ser
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                                          815
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          885 890
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	gcc Ala															283
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	gag Glu															475
	cgc Arg															523
	gag Glu 105															571
	gcc Ala															619
	gcg Ala															667

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		_		_	gta Val					-				-	_	1771
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		tg gct tac tgt acc t eu Ala Tyr Cys Thr L 885	
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			PRT Zea		s											
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Met		400> Ala		Ala	Glv	Len	Val	Δla	G1v	Ser	Hic	Δen	Ara	λen	Glu	
1				5					10					15		
			20					25					30		Gly	
Ala	Ala	Arg 35	Arg	Ala	Glu	Ala	Pro	Cys	Gln	Ile	Cys	Gly 45	Asp	Glu	Val	
Gly	Val		Phe	Asp	Gly	Glu 55		Phe	Val	Ala	Cys 60		Glu	Cys	Ala	
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Ser	Ser	Ser 275	Arg	Ile	Asn	Pro	Tyr 280	Arg	Met	Ile	Ile	Val 285	Ile	Arg	Leu	
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Asp	Gly	Ala	Ala	Met 405	Leu	Thr	Phe	Glu	Ala 410	Leu	Ser	Glu	Thr	Ser 415	Glu
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	770		Суз			775					780				
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Phe Ser Tyr Ile Asn Ser Ile Val Tyr Pro Trp Thr Ser Ile Pro Leu
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Ile Thr Pro Glu Leu Thr Asn Val Ala Ser Ile Trp Phe Met Ala Leu
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915 920 925
Val Ala Ile Asp Asp Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly
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Gly Val Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val
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Phe Ala Gly Ile Asp Thr Ser Phe Thr Val Thr Ser Lys Ala Gly Asp
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Asp Glu Glu Phe Ser Glu Leu Tyr Thr Phe Lys Trp Thr Thr Leu Leu
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Ile Pro Pro Thr Thr Leu Leu Leu Leu Asn Phe Ile Gly Val Val Ala
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Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile Val His Leu Tyr Pro
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Phe Leu Lys Gly Leu Val Gly Arg Gln Asn Arg Thr Pro Thr Ile Val
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Ile Val Trp Ser Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val
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	cca Pro															1265
	tgg Trp															1313
	tac Tyr															1361
	cag Gln															1409
	gag Glu 365															1457
	gat Asp				_	_	-		_		-					1505
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	gaa Glu															1649
	cct Pro 445															1697
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	gag Glu	_				_		_								1793
	acc Thr		_				-		_	_					_	1841
	ggc										_	_	_		-	1889
	cgt Arg 525	-	_	-				_			_		_		-	1937
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-	tat Tyr 605	_								_			-	_		2177
	gat Asp															2225

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	gtg Val 685															2417
	gga Gly															2465
	aag Lys	_		•	~		_			_	-			_	~	2513
	tat Tyr			_		_		_		_				_		2561
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	gga Gly		-	-			-				-					2705
	aag Lys			-		_			_							2753
	ctg Leu															2801
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	ctg Leu 845															2897
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Ser 860	Ile	Pro	Leu	Leu	Ile 865	Tyr	Cys	Ile	Leu	Pro 870	Ala	Ile	Cys	Leu	Leu 875	
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	tgg Trp	_					-									3089
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-	gcc Ala	-	-		_	-	_		_	_	_		-			3233
	acg Thr															3281
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	tgg Trp 100	Gly					Lys					Phe				3377
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	ccg Pro				Val					Leu					Phe	3473
	ttg Leu			Val					Phe					Thr		3521
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365

395

355 360

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	690		Asp			695	_			-	700	_			
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60

120

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						gga Gly										324
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-		_	_			gat Asp		_		_	_	-	-	-		516
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Val	Thr 225	Asn	Lys	Tyr	Pro	Glu 230	Ala	Arg	Gly	Gly	Asp 235	Met	Glu	Gly	Thr	
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_		tgc Cys		-			_	_					_	_		1140
		tgg Trp					-					_			_	1188
_		tat Tyr	_							_	-	_			_	1236
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-		act Thr 370	_	_						_			-	-		1332
		tgc Cys		-		_	-			-	_	-			-	1380
		tca Ser	Glu	Thr		_	Phe		Arg	Lys	Trp	-		Phe		1428
-	_	cac His			-		_	-		_				-		1476
		gat Asp		_	_	-									_	1524
		gca Ala 450														1572

gcc Ala	ctt Leu 465	gtt Val	gcc Ala	aaa Lys	gca Ala	cag Gln 470	aaa Lys	gtg Val	cct Pro	gaa Glu	gag Glu 475	Gly 999	tgg Trp	acc Thr	atg Met	1620
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aat Asn	gag Glu	tta Leu	cca Pro 515	cgt Arg	ctt Leu	gtc Val	tat Tyr	gtc Val 520	tct Ser	cgt Arg	gaa Glu	aag Lys	aga Arg 525	cca Pro	Gly	1764
Phe	Gln	His 530	cac His	Lys	Lys	Ala	Gly 535	Ala	Met	Asn	Ala	Leu 540	Ile	Arg	Val	1812
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			gtt Val													2148
			tgt Cys													2196
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			atg Met													2340
			att Ile													2388
			cca Pro													2436
			gag Glu 755				-									2484
			gta Val													2532
			caa Gln				-	_			_		_		_	2580
			cca Pro					_	_			_			_	2628
			gjå 333			-		-		-	-		-			2676
			tac Tyr 835					_		_			_			2724
			att Ile	•							-			-		2772
-			ccc Pro	-		_						Phe				2820
			aat Asn													2868
			act Thr													2916
-	_		tgg Trp 915	_			_			_						2964

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gcc Ala i																3012
att g																3060
gac (Asp) 960								_			_	_				3108
eeg a																3156
tct (Asn					Ser					Phe		3204
aag d Lys 1	-		Phe	-				Ile					Pro			3252
aag q Lys (Leu	_			_	Asn	_				Ile	-			3300
tgg t Trp S						Ser					Leu					3348
gat o					Pro					Ala					Cys	3396
ggc g			t go	tgat	cgag	g aca	igtga	actc	ttat	ttga	iag a	ıggct	caat	c		3446
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Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Ser Val 35 40 Gly Val Ser Ala Thr Gly Asp Val Phe Val Ala Cys Asn Glu Cys Ala Phe Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Glu Gly Asn 75 70 Gln Cys Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Gln Lys Gly Ser 90 85 Pro Arg Val His Gly Asp Glu Asp Glu Glu Asp Val Asp Asp Leu Asp 105 Asn Glu Phe Asn Tyr Lys Gln Gly Ser Gly Lys Gly Pro Glu Trp Gln 115 120 125 Leu Gln Gly Asp Asp Ala Asp Leu Ser Ser Ala Arg His Glu Pro 135 140 His His Arg Ile Pro Arg Leu Thr Ser Gly Gln Gln Ile Ser Gly Glu 150 155 Ile Pro Asp Ala Ser Pro Asp Arg His Ser Ile Arg Ser Pro Thr Ser 165 170 175 Ser Tyr Val Asp Pro Ser Val Pro Val Pro Val Arg Ile Val Asp Pro 180 185 190 Ser Lys Asp Leu Asn Ser Tyr Gly Leu Asn Ser Val Asp Trp Lys Glu 200 205 Arg Val Glu Ser Trp Arg Val Lys Gln Asp Lys Asn Met Met Gln Val 215 220 Thr Asn Lys Tyr Pro Glu Ala Arg Gly Gly Asp Met Glu Gly Thr Gly 230 235 240 Ser Asn Gly Glu Xaa Met Gln Met Val Asp Asp Ala Arg Leu Pro Leu 245 250 255 Ser Arg Ile Val Pro Ile Ser Ser Asn Gln Leu Asn Leu Tyr Arg Val 260 265 270 Val Ile Ile Leu Arg Leu Ile Ile Leu Cys Phe Phe Phe Gln Tyr Arg 275 280 285 Val Ser His Pro Val Arg Asp Ala Tyr Gly Leu Trp Leu Val Ser Val 295 300 Ile Cys Glu Val Trp Phe Ala Leu Ser Trp Leu Leu Asp Gln Phe Pro 310 315 Lys Trp Tyr Pro Ile Asn Arg Glu Thr Tyr Leu Asp Arg Leu Ala Leu 325 330 335 Arg Tyr Asp Arg Glu Gly Glu Pro Ser Gln Leu Ala Pro Ile Asp Val 340 345 350 Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Ile Thr Ala · 355 360 Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Asp Lys Val 370 375 380 Ser Cys Tyr Val Ser Asp Asp Gly Ser Ala Met Leu Thr Phe Glu Ser 395 400 390 Leu Ser Glu Thr Ala Glu Phe Ala Arg Lys Trp Val Pro Phe Cys Lys 405 410 415 Lys His Asn Ile Glu Pro Arg Ala Pro Glu Phe Tyr Phe Ala Gln Lys 420 425 430 Ile Asp Tyr Leu Lys Asp Lys Ile Gln Pro Ser Phe Val Lys Glu Arg 435 440 445 Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala 460 450 455 Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr Met Ala 470 475 480 Asp Gly Thr Ala Trp Pro Gly Asn Asn Pro Arg Asp His Pro Gly Met 490

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Ile	Gln	Val	Phe 500	Leu	Gly	His	Ser	Gly 505	Gly	Leu	Asp	Thr	Asp 510	Gly	Asn
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Phe	Phe 610	Asp	Ile	Asn	Met	Lys 615	Gly	Leu	Asp	Gly	Ile 620	Gln	Gly	Pro	Val
Tyr 625	Val	Gly	Thr	Gly	Cys 630	Cys	Phe	Asn	Arg	Gln 635	Ala	Leu	Tyr	Gly	Tyr 640
Asp	Pro	Val	Leu	Thr 645	Glu	Ala	Asp	Leu	Glu 650	Pro	Asn	Ile	Val	Ile 655	Lys
Ser	Cys	Суз	Gly 660	Arg	Arg	Lys	Lys	Lys 665	Asn	Lys	Ser	Tyr	Met 670	Asp	Ser
		675	Ile		_	_	680					685			
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705			Ser		710	_			_	715		-			720
			Ala	725					730					735	
			Ala 740					745					750		
		755	Asp				760					765			
_	770		Thr		_	775					780				
785			Ser		790					795					800
			Ile	805					810					815	
		_	Ser 820					825					830		
-		835	Asn	_	_		840					845			
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	930		Ala			935					940				
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Tyr Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys
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                                        1005
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  1010 1015 1020
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Ser Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Lys Ile Asp
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Ser His Asn Arq Asn Glu Leu Val Val Ile Arg Arg Asp Gly Asp Pro
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Gly Pro Lys Pro Pro Arg Glu Gln Asn Gly Gln Val Cys Gln Ile Cys
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	gac Asp															509
	gcc Ala			-						-			-			557
	cct Pro 140															605
	ct <i>c</i> Leu							-								653
	ctg Leu															701
	cct Pro			-		_										749
	tcc Ser															797
	cgg Arg 220															845
	aat Asn															893
	gat Asp															941
agc	cag	att	aat	сса	tat	agg	atg	att	atc	att	att	cgg	ctt	gtg	gtt	989

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Ser	Gln	Ile	Asn 270	Pro	Tyr	Arg	Met	Ile 275	Ile	Ile	Ile	Arg	Leu 280	Val	Val	
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	gct Ala 300															108
	tgg Trp															113
	tac Tyr															118
	caa Gln								_	-	_	_	_			122
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	gat Asp 380			-		-	_		_		_		-	_		132
	gca Ala	_				_	_			-			_		-	137
_	aaa Lys					-						_		-		142
	gag Glu					_	_		_		_		-	_		1469
	gca Ala			Val	Arg		Arg	Arg								151
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	gaa Glu	_				_		-				-				1613
	gtt Val															1661

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	aga Arg			_							_		_		_	1757
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_	tta Leu											_	_	_		1853
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	tat Tyr								_			-	_		-	1949
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	ttt Phe															2189
_	aaa Lys	_					_		-						_	2237
	gct Ala 700			_		-	-	_	_			_			-	2285
	gcc Ala															2333
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agt Ser	tgt Cys	ggt Gly 765	tat Tyr	gaa Glu	gac Asp	aag Lys	aca Thr 770	gac Asp	tgg Trp	gga Gly	aaa Lys	gag Glu 775	att Ile	ggc	tgg Trp	2477
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		_	tgg Trp										_	_		2573
aaa Lys	ggt Gly	tct Ser	gca Ala	cct Pro 815	ctg Leu	aat Asn	ctt Leu	tca Ser	gat Asp 820	cgt Arg	ctt Leu	cac His	cag Gln	gtg Val 825	ctt Leu	2621
			ctt Leu 830													2669
			Gly ggg													2717
			tcc Ser													2765
			ttg Leu													2813
		_	aat Asn		-		_	_			_					2861
			gct Ala 910						_	_		-		-		2909
			tgg Trp													2957
			ctc Leu													3005
	-	-	aca Thr	-						_			_	_		3053

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cct acc Pro Thr		Leu L	-			Ile			_	-	Gly	_	3149
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aaa ggt Lys Gly 1035		Gly A				Thr		Thr		_		-	3293
tgg tcc . Trp Ser	_					_	Leu			_		Ile	3341
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70 Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Leu Lys Gly Cys Gln Arĝ 85 90 95 Val Thr Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Asp Asn Glu 100 105 Phe Asn Trp Asp Gly His Asp Ser Gln Ser Val Ala Glu Ser Met Leu 120 125 Tyr Gly His Met Ser Tyr Gly Arg Gly Gly Asp Pro Asn Gly Ala Pro 135 140 Gln Ala Phe Gln Leu Asn Pro Asn Val Pro Leu Leu Thr Asn Gly Gln 150 Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu Val Pro Ser Phe 165 170 Met Gly Gly Gly Lys Arg Ile His Pro Leu Pro Tyr Ala Asp Pro 180 185 190 Ser Leu Pro Val Gln Pro Arg Ser Met Asp Pro Ser Lys Asp Leu Ala 200 Ala Tyr Gly Tyr Gly Ser Val Ala Trp Lys Glu Arg Met Glu Asn Trp 215 220 Lys Gln Arg Gln Glu Arg Met His Gln Thr Gly Asn Asp Gly Gly Gly 230 235 Asp Asp Gly Asp Asp Ala Asp Leu Pro Leu Met Asp Glu Ala Arg Gln 245 250 Gln Leu Ser Arg Lys Ile Pro Leu Pro Ser Ser Gln Ile Asn Pro Tyr 260 265 270 Arg Met Ile Ile Ile Arg Leu Val Val Leu Gly Phe Phe Phe His 275 280 285 Tyr Arg Val Met His Pro Val Asn Asp Ala Phe Ala Leu Trp Leu Ile 295 Ser Val Ile Cys Glu Ile Trp Phe Ala Met Ser Trp Ile Leu Asp Gln 310 315 320 Phe Pro Lys Trp Phe Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu 330 325 Ser Leu Arg Phe Asp Lys Glu Gly Gln Pro Ser Gln Leu Ala Pro Ile 340 345 Asp Phe Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Val 360 Thr Thr Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Asp 370 375 380 Lys Val Ser Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe 390 395 Glu Ala Leu Ser Glu Thr Ser Glu Phe Ala Lys Lys Trp Val Pro Phe 410 Cys Lys Arg Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Gln 425 420 430 Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val Ala Ala Asn Phe Val Arg 440 Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile 455 460 Asn Ala Leu Val Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr 470 475 Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Val Arg Asp His Pro 485 490 Gly Met Ile Gln Val Phe Leu Gly Gln Ser Gly Gly Leu Asp Cys Glu 500 505 Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro 520 Gly Tyr Asn His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg

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705		Ala			710					715		-			720
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-	770	Ile	_	_	-	775		-	_		780				
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		Ser		805					810					815	
		Ile	820					825					830		
		835 Leu					840					845			
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865					870					875	-2-				880
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		Glu 915					920					925			
	930	Gln		-		935	_	_			940				
Val 945	Phe	Gln	Gly	Leu	Leu 950	Lys	Val	Ile	Ala	955	Val	Asp	Thr	ser	960
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Met Ala Ala Asn Lys Gly Met Val Ala Gly Ser His Asn Arg Asn Glu
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                                                                274
Phe Val Met Ile Arg His Asp Gly Asp Ala Pro Val Pro Ala Lys Pro
                              25
                                                                322
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Thr Lys Ser Ala Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Thr Val
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gge gtt tea gee act ggt gat gte ttt gtt gee tge aat gag tgt gee
Gly Val Ser Ala Thr Gly Asp Val Phe Val Ala Cys Asn Glu Cys Ala
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		tgc Cys														466
	_	gtt Val			_	-	-		-	-		-	-	_	-	514
	_	ttc Phe 115			_								-			562
		gga Gly	-	-	-	_	_				-	_				610
		cgg Arg														658
		gat Asp														706
_		gtt Val	_		_	_		-						_		754
_	_	gac Asp 195	-												_	802
		gag Glu														850
		aaa Lys					-				-					898
		gaa Glu														946
		gtg Val														994
		ctc Leu 275											Tyr			1042
agt	cat	cca	gtg	cgt	aat	gct	tat	gga	ttg	tgg	cta	gta	tct	gtt	atc	1090

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					Arg						agg Arg					118	6
											ccc Pro					123	4
											ctg Leu					128	2
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tgc Cys 385	tat Tyr	gtt Val	tct Ser	gat Asp	gat Asp 390	ggc Gly	tca Ser	gct Ala	atg Met	ctg Leu 395	act Thr	ttt Phe	gag Glu	tct Ser	ctc Leu 400	1371	8
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											ttt Phe					1474	1
											gtt Val					1522	2
-	_	_	_			_	~				aga Arg 460			_		1570)
			Ala	Gln		Val			Glu		tgg Trp			Ala		1618	3
		-								_	cat His			_		1666	5
			-	-		_				-	act Thr	_				1714	Ŀ
		_		-		-		_	_	_	aga Arg				-	1762	<u>}</u>

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		aag Lys					_		_	_		-	-		-	1810
		aca Thr														1858
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		aca Thr														2098
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_	_	att Ile 675	_	_	-		_			_					_	2242
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		gca Ala														2386
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Tyr	Glu	Asp 755	Lys	Thr	Glu	Trp	Gly 760	Lys	Glu	Ile	Gly	Trp 765	Ile	Tyr	Gly	
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	caa Gln				_	_			_		_		_			2578
_	cca Pro						-			-			-		-	2626
	eja aaa			-		_		_	-		_					2674
	tac Tyr				-			_	-		_					2722
	att Ile 850	-						_	_			-		-		2770
	cct Pro	-		-												2818
_	aat Asn		-		_									_	_	2866
_	act Thr			_			-		_		-				-	2914
	tgg Trp															2962
	ttc Phe 930				_		_	_			_	-		_		3010
	aac Asn			_			-	-		-		-				3058
	gag Glu															3106
	gtt Val															3154

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Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys Leu 995 1000 1005	
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ctc atg ggc agg cag aac cgc acg cca aca atc gtc atc gtt tgg tcc Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp Ser 1025 1030 1035 1040	3298
atc ctc ctt gcg tct atc ttc tcc ttg ctg tgg gtg aag atc gat cct Ile Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Lys Ile Asp Pro 1045 1050 1055	3346
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Cys 305	Glu	Val	Trp	Phe	Ala 310	Leu	Ser	Trp	Leu	Leu 315	qaA	Gln	Phe	Pro	Lys 320
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Ser	Glu	Thr	Ala	Glu 405	Phe	Ala	Arg	Lys	Trp 410	Val	Pro	Phe	Cys	Lys 415	Lys
	Asn		420		_			425		•			430	-	
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	Met 450	_	_		-	455			_		460				
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	Thr			485					490					495	
	Val		500	_			_	505		_		_	510		
	Pro	515			-		520	_		-	_	525	_		
	His 530					535					540				
545	Leu			_	550	_				555	_	_			560
	Asn			565					570					575	
	Ala		580					585					590		
_	Gly	595	_			_	600	_			_	605		_	
	Asp 610				_	615		_	_		620	_			
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625	**- 1	T	Mh	a1	630		•	01	D	635	71.	77- 1	770 7	*	640
Pro	vaı	Leu	Thr	645	Ata	Asp	Leu	GIU	650	ASI	TTE	var	val	655	ser
Cvs	Cva	Glv	Arg		Lvs	Ara	I.ve	Δsn		Ser	TVY	Met	Asp		Gln
Cyb	- 70	 2	660		-,-	9		665	-7-		- 7 -		670		
Ser	Arg	Ile	Met	Lys	Arq	Thr	Glu		Ser	Ala	Pro	Ile		Asn	Met
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Glu	Asp	Ile	Glu	Glu	Gly	Ile	Glu	Gly	Tyr	Glu	Asp	Glu	Arg	Ser	Val
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Leu	Met	Ser	Gln	Arg	Lys	Leu	Glu	Lys	Arg	Phe	Gly	Gln	Ser	Pro	Ile
705					710					715					720
Phe	Ile	Ala	Ser		Phe	Met	Thr	Gln	-	Gly	Ile	Pro	Pro		Thr
_	_		_	725		_	~-		730					735	~1 ·-
Asn	Pro	Ala	Ser	Leu	Leu	гÀг	Glu		lle	HIS	vaı	TTE		Cys	GIY
Tr. cor	C1	200	740 Lys	Thr	C1.,	æν	C111	745	C111	T10	Clv	Tvn	750	The	Gly
ıyı	GIU	755	nys	1111	GIU	115	760	nys	GIU	116	Gry	765	116	1 y 1	Gry
Ser	Val		Glu	Asp	Ile	Leu		Glv	Phe	Lvs	Met		Ala	Ara	Glv
	770			- to-p		775		,		-,-	780			3	,
Trp		Ser	Ile	Tyr	Cys		Pro	Pro	Arg	Pro		Phe	Lys	Gly	Ser
785				•	790				_	795	•		-	-	800
Ala	Pro	Ile	Asn	Leu	Ser	Asp	Arg	Leu	Asn	Gln	Val	Leu	Arg	Trp	Ala
				805					810					815	
Leu	Gly	Ser	Val	Glu	Ile	Leu	Leu		Arg	His	Суз	Pro		Trp	Tyr
			820					825					830		
Gly	Tyr		Gly	Arg	Leu	Lys		Leu	Glu	Arg	Leu		Tyr	ITe	Asn
ml	-1 -	835		D	71 -	mla sa	840	*7-1	D	*	71 ~	845	M	Crea	17-1
Thr		vaı	Tyr	Pro	IIe	855	ser	vai	Pro	Leu	860 TTG	Ата	TYE	Cys	vai
T.011	850 Pro	A 1 =	Ile	Cve	T.e.ii		Thr	Δan	LVS	Phe		Tle	Pro	Glu	Ile
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Trp	\mathtt{Trp}		Asn	Glu	Gln	Phe		Val	Ile	Gly	Gly		Ser	Ala	His
_		915	•			~-	920	_				925	~ 1	- 1-	3
Leu		Ala	Val	Phe	Gin	-	Leu	Leu	гуз	val		Ala	GIY	TIE	Asp
PT las sa	930	Dho	Thr	3703	The sec	935	7	חות	602) co	940	N G YO	Glv	y en	Dhe
945	ASII	Pne	1111	vaı	950	ser	цуз	ALA	Set	955	Gra	ASP	Gry	qaa	960
	Glu	T.e.11	Tyr	Val		Lvs	Trn	Thr	Ser		Leu	Ile	Pro	Pro	
7124	0.2	200	-7-	965		_,_			970					975	
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			980					985				_	990		
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		995					1000					1005			
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102		_		_	1030		_	_	_	1039		_			1040
Ile	Leu	Leu	Ala			Pne	ser	Leu			val	гÀг	TTG		
D !	-7 -	0	D	1045		T	71-	A 7 -	1050		G1	G3~	Care	1055	
Fue	тте	ser	Pro 1060		GIII	туа	WTG	1065		neu	GTÅ	G 711	1070		· u.i
Acr	Cys		1000	•				700	•					-	
Part	Cys														

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	-	_				-	_						ctg Leu	_		571
													tcc Ser			619
		_	-	-			-			_			ccc Pro			667
													atg Met 165			715
_		-	_							_			atg Met	_		763
					_				_				gca Ala	_		811
													gat Asp			859
_					-		_				-	_	gag Glu			907
_	_		-		-	-			-				ggt Gly 245			955
_		-	_		-	-							gaa Glu			1003
_		-		_		_					_	_	att Ile			1051
													ttc Phe			1099
		_		-		_			_	_		-	ttg Leu			1147
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cag	ttc	cca	aag	tgg	ctt	cca	atc	gag	aga	gag	act	tac	ctg	gac	cgt	1243

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Gln	Phe	Pro 330	Lys	Trp	Leu	Pro	Ile 335	Glu	Arg	Glu	Thr	Tyr 340	Leu	Asp	Arg		
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		ttc Phe															1339
		gcg Ala															1387
		gtc Val						_	-		_	_	_		_		1435
		gca Ala 410	_		-			-		_	_			_			1483
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		gcc Ala	_	_	_		-		_	-			_			:	1675
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		atg Met			Val		Leu									;	1771
-		aat Asn		_		_	_	_		-	_	-	_	_	-	:	1819
		tat Tyr				_	_	-		_	_		_	_		3	1867
		tct Ser														3	1915

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	gat Asp												_	_	_	19	63
	atg Met 585							_			_		-	_		20	11
	cag Gln															20	59
	gtt Val															21	07
	ccc Pro								_		_		-	_	_	21	55
	ggt Gly			_									-		_	220	03
	tgc Cys 665			_		_			_	-	_	-			-	22	51
	aaa Lys		-					_	_		_	_		-		229	99
	ttc Phe	_		-	-					-		_	_			234	17
	gat Asp	-		-			-				-	-				235	₹5
	caa Gln	_				_				-			_		_	244	13
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	tct Ser															253	39
-	aag Lys		-											_		258	37
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											-		_	ctt Leu		2731
	_	-				-	_		_					gga Gly		2779
						_	_							tcc Ser 870		2827
-									-	-		_		ttg Leu		2875
_		-	-		_		_							acc Thr		2923
-	_	_				_	-				_			gtg Val		2971
		_	_	-			_			_		-	-	tgg Trp		3019
			_			-				-	_			ctg Leu 950		3067
			_		_	_	_	~ ~		_			_	acg Thr	-	3115
			_	_	-	_		_	-				_	gag Glu		3163
	_	Phe	_				Leu	_				Thr		ctc Leu		3211
	Leu					Val					Ser			atc Ile		3259
					Trp					Gly				ttc Phe 1030	Ala	3307

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agg cag aac agg acg ccg acg atc gtc atc gtc tgg tcc atc ctg ctg Arg Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp Ser Ile Leu Leu 1050 1055 1060	3403
gcc tcg atc ttc tcg ctc ctg tgg gtc cgc gtc gac ccg ttc ctc gcc Ala Ser Ile Phe Ser Leu Leu Trp Val Arg Val Asp Pro Phe Leu Ala 1065 1070 1075	3451
aag age aac ggc ccg ctc ctg gag gag tgt ggc ctg gac tgc a Lys Ser Asn Gly Pro Leu Leu Glu Glu Cys Gly Leu Asp Cys 1080 1085 1090	3494
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Pro Arg Val Ala Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Glu 100 105 110 Gly Glu Phe Gly Leu Gln Asp Gly Ala Ala His Glu Asp Asp Pro Gln 115 120 125 Tyr Val Ala Glu Ser Met Leu Arg Ala Gln Met Ser Tyr Gly Arg Gly 130 135 140 Gly Asp Ala His Pro Gly Phe Ser Pro Val Pro Asn Val Pro Leu Leu 145 150 155 160 Thr Asn Gly Gln Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu	
Pro Arg Val Ala Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Glu 100 105 110 Gly Glu Phe Gly Leu Gln Asp Gly Ala Ala His Glu Asp Asp Pro Gln 115 120 125 Tyr Val Ala Glu Ser Met Leu Arg Ala Gln Met Ser Tyr Gly Arg Gly 130 135 140 Gly Asp Ala His Pro Gly Phe Ser Pro Val Pro Asn Val Pro Leu Leu 145 150 155 160 Thr Asn Gly Gln Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu 165 170 175 Val Pro Ser Tyr Met Ser Gly Gly Gly Gly Gly Gly Lys Arg Ile His	
Pro Arg Val Ala Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Glu 100 105 110 Gly Glu Phe Gly Leu Gln Asp Gly Ala Ala His Glu Asp Asp Pro Gln 125 Tyr Val Ala Glu Ser Met Leu Arg Ala Gln Met Ser Tyr Gly Arg Gly 130 135 Gly Asp Ala His Pro Gly Phe Ser Pro Val Pro Asn Val Pro Leu Leu 140 Thr Asn Gly Gln Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu 160 Thr Asn Gly Gln Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu 175	

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Val	Val 290	Leu	Gly	Phe	Phe	Phe 295	His	Tyr	Arg	Val	Met 300	His	Pro	Ala	Lys
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				325					330				Pro	335	
			340					345					Lys 350		_
		355					360					365	Thr		_
	370	_				375					380		Leu		
Leu 385	Ser	Val	Asp	Tyr	9ro 390	Val	Glu	Lys	Val	Ser 395	Cys	Tyr	Val	Ser	Asp 400
Asp	Gly	Ala	Ala	Met 405	Leu	Thr	Phe	Glu	Ala 410	Leu	Ser	Glu	Thr	Ser 415	Glu
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Lys	Val 450	Ala	Ala	Ser	Phe	Val 455	Arg	Glu	Arg	Arg	Ala 460	Met	Lys	Arg	Glu
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Lys	Val	Pro	Glu	Glu 485	Gly	Trp	Thr	Met	Gln 490	Asp	Gly	Ser	Pro	Trp 495	Pro
Gly	Asn	Asn	Val 500	Arg	Asp	His	Pro	Gly 505	Met	Ile	Gln	Val	Phe 510	Leu	Gly
Gln	Ser	Gly 515	Gly	Arg	Asp	Val	Glu 520	Gly	Asn	Glu	Leu	Pro 525	Arg	Leu	Val
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Lys	Val	Cys 595	Tyr	Val	Gln	Phe	Pro 600	Gln	Arg	Phe	Asp	Gly 605	Ile	Asp	ГÀЗ
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Lys 625	Gly	Leu	Asp	Gly	Ile 630	Gln	Gly	Pro	Ile	Tyr 635	Val	Gly	Thr	Gly	Cys 640
	Phe	Arg	Arg	Gln 645		Leu	Tyr	Gly	Tyr 650		Ala	Pro	Lys	Thr 655	
Lys	Pro	Pro	Ser 660		Thr	Cys	Asn	Cys 665		Pro	Lys	Trp	Cys 670		Ser
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Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Asp Trp Gly Lys Glu Ile
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Gly Trp Ile Tyr Gly Ser Ile Thr Glu Asp Ile Leu Thr Gly Phe Lys
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Met His Cys His Gly Trp Arg Ser Ile Tyr Cys Ile Pro Lys Arg Pro
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Ala Phe Lys Gly Ser Ala Pro Leu Asn Leu Ser Asp Arg Leu His Gln
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Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe Phe Ser Lys His
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Cys Pro Leu Trp Tyr Gly Tyr Gly Gly Gly Leu Lys Phe Leu Glu Arg
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Phe Ser Tyr Ile Asn Ser Ile Val Tyr Pro Trp Thr Ser Ile Pro Leu
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Leu Ala Tyr Cys Thr Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe
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Phe Ile Cys Ile Ser Val Thr Gly Ile Leu Glu Met Arg Trp Ser Gly
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gtoottttot otogtoooto otoccoccgt atagttaago cocgocccgo tactactact
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                                                                      240
tetegggaet ggtgeegget etgeeeagge eecaggetee aggeeagete eetegaegtt
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Arg Arg Gly Gly Gln Val Cys Gln Ile Cys Gly Asp Gly Val Gly
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Thr Thr Ala Glu Gly Asp Val Phe Ala Ala Cys Asp Val Cys Gly Phe
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Pro Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Asp Gly Thr Gln
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                         50
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Ala Cys Pro Gln Cys Lys Thr Lys Tyr Lys Arg His Lys Gly Ser Pro
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                     65
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                                                                      593
Ala Ile Arg Gly Glu Glu Gly Asp Asp Thr Asp Ala Asp Ser Asp Phe
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                                     85
                                                                      641
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Asn Tyr Leu Ala Ser Gly Asn Glu Asp Gln Lys Gln Lys Ile Ala Asp
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						cct Pro										1121
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						tac Tyr 290										1217
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	-	tgg Trp 430				-			-		-	_	-			1649
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		mgg Xaa	_				_		_	-	,					1841
		ctt Leu 510														1889
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		gct Ala														1985
		aat Asn														2033
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			_		tca Ser		_				_	_	-	-		2	2321
	-	_		_	gac Asp	_	-	_	-		_			_	_	2	2369
			-	_	aac Asn			-					-	_		2	2417
					gag Glu 705					-			_	_	_	2	2465
	_	-			cag Gln		-			_	_			-	-	2	2513
					cct Pro											2	2561
_	_			_	ata Ile	_	-				_			_		2	2609
					tgg Trp											2	2657
					cac His 785											2	2705
					ttc Phe											2	2753

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		gct ctt ggg tcc gtg gag atc ctc Ala Leu Gly Ser Val Glu Ile Leu 820 825	2801
		tac ggc tac gga ggg cgg ctc aag Tyr Gly Tyr Gly Gly Arg Leu Lys 840	2849
		aac acc acc atc tac ccg ctc acg Asn Thr Thr Ile Tyr Pro Leu Thr 855	2897
•	-	atc ctg ccc gcc atc tgt ctg ctc Ile Leu Pro Ala Ile Cys Leu Leu 870 875	2945
	: Ile Pro Glu	atc agc aac ttc gcc agc atc tgg Ile Ser Asn Phe Ala Ser Ile Trp 885 890	2993
	_	ttc gcc acg ggc atc ctg gag atg Phe Ala Thr Gly Ile Leu Glu Met 900 905	3041
		gag tgg tgg agg aac gag cag ttc Glu Trp Trp Arg Asn Glu Gln Phe 920	3089
		cac ctc ttc gcc gtg ttc cag ggc His Leu Phe Ala Val Phe Gln Gly 935	3137
		gac acc aac ttc acc gtc acc tcc Asp Thr Asn Phe Thr Val Thr Ser 950 955	3185
	Asp Gly Asp	ttc gcg gag ctg tac atg ttc aag Phe Ala Glu Leu Tyr Met Phe Lys 965 970	3233
		acc acc atc ctg atc atc aac ctg Thr Thr Ile Leu Ile Ile Asn Leu 980 985	3281
		tac gcc atc aac agc gga tac cag Tyr Ala Ile Asn Ser Gly Tyr Gln 5 1000	3329
		ctc ttc ttc gcc ttc tgg gtc atc Leu Phe Phe Ala Phe Trp Val Ile 1015	3377
gtc cac ctg tac ccg Val His Leu Tyr Pro 1020	ttc ctc aag Phe Leu Lys 1025	ggc ctc atg ggc agg cag aac cgc Gly Leu Met Gly Arg Gln Asn Arg 1030 1035	3425
acc ccg acc atc gto Thr Pro Thr Ile Val		gccatcctgc tggcgtccat cttctccttg	3479

1040

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SO	
Lys Thr Lys Tyr Lys Arg His Lys Gly Ser Pro Ala Ile Arg Gly Glu 65	
Glu Gly Asp Asp Thr Asp Ala Asp Ser Asp Phe Asn Tyr Leu Ala Ser 85 90 95 Gly Asn Glu Asp Gln Lys Gln Lys Ile Ala Asp Arg Met Arg Ser Trp 100 105 110 Arg Met Asn Val Gly Gly Ser Gly Asp Val Gly Arg Pro Lys Tyr Asp 115 120 125 Ser Gly Glu Ile Gly Leu Thr Lys Tyr Asp Ser Gly Glu Ile Pro Arg 130 135 140 Gly Tyr Ile Pro Ser Val Thr Asn Ser Gln Ile Ser Gly Glu Ile Pro 160 Gly Ala Ser Pro Asp His His Met Met Ser Pro Thr Gly Asn Ile Gly 165 150 Lys Arg Ala Pro Phe Pro Tyr Val Asn His Ser Pro Asn Pro Ser Arg 180 185 190 Glu Phe Ser Gly Ser Ile Gly Asn Val Ala Trp Lys Glu Arg Val Asp 195 Gly Trp Lys Met Lys Gln Asp Lys Gly Thr Ile Pro Met Thr Asn Gly 210 Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230	
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Arg Met Asn Val Gly Gly Ser Gly Asp Val Gly Arg Pro Lys Tyr Asp 115	
Ser Gly Glu Ile Gly Leu Thr Lys Tyr Asp Ser Gly Glu Ile Pro Arg 130 135 140 Gly Tyr Ile Pro Ser Val Thr Asn Ser Gln Ile Ser Gly Glu Ile Pro 145 150 155 160 Gly Ala Ser Pro Asp His His Met Met Ser Pro Thr Gly Asn Ile Gly 165 170 175 175 Lys Arg Ala Pro Phe Pro Tyr Val Asn His Ser Pro Asn Pro Ser Arg 180 185 190 561 190 Glu Phe Ser Gly Ser Ile Gly Asn Val Ala Trp Lys Glu Arg Val Asp 195 200 205 205 Gly Trp Lys Met Lys Gln Asp Lys Gly Thr Ile Pro Met Thr Asn Gly 215 220 220 Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230 235 240	
Gly Tyr Ile Pro Ser Val Thr Asn Ser Gln Ile Ser Gly Glu Ile Pro 145	
Gly Ala Ser Pro Asp His His Met Met Ser Pro Thr Gly Asn Ile Gly 165 170 175 Lys Arg Ala Pro Phe Pro Tyr Val Asn His Ser Pro Asn Pro Ser Arg 180 185 190 Glu Phe Ser Gly Ser Ile Gly Asn Val Ala Trp Lys Glu Arg Val Asp 195 200 205 Gly Trp Lys Met Lys Gln Asp Lys Gly Thr Ile Pro Met Thr Asn Gly 210 215 Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230 235 240	
Lys Arg Ala Pro Phe Pro Tyr Val Asn His Ser Pro Asn Pro Ser Arg 180	
Glu Phe Ser Gly Ser Ile Gly Asn Val Ala Trp Lys Glu Arg Val Asp 195 200 205 Gly Trp Lys Met Lys Gln Asp Lys Gly Thr Ile Pro Met Thr Asn Gly 210 215 220 Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230 240	
Gly Trp Lys Met Lys Gln Asp Lys Gly Thr Ile Pro Met Thr Asn Gly 210 215 220 Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230 235 240	
Thr Ser Ile Ala Pro Ser Glu Gly Arg Gly Val Gly Asp Ile Asp Ala 225 230 235 240	
Ser the ASD Tyr ASH Met Gid ASD Ala Led Led ASH ASD Gid inc Ald	
245 250 255	
Gln Pro Leu Ser Arg Lys Val Pro Leu Pro Ser Ser Arg Ile Asn Pro 260 265 270	
Tyr Arg Met Val Ile Val Leu Arg Leu Ile Val Leu Ser Ile Phe Leu 275 280 285	
His Tyr Arg Ile Thr Asn Pro Val Arg Asn Ala Tyr Pro Leu Trp Leu 290 295 300	
Leu Ser Val Ile Cys Glu Ile Trp Phe Ala Leu Ser Trp Ile Leu Asp	
305 310 315 320	
Gln Phe Pro Lys Trp Phe Pro Ile Asn Arg Glu Thr Tyr Leu Asp Arg 325 330 335	
Leu Ala Leu Arg Tyr Asp Arg Glu Gly Glu Pro Ser Gln Leu Ala Ala 340 345 350	

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Val	Asp	11e 355	Phe	Val	Ser	Thr	Val 360	Asp	Pro	Met	Lys	Glu 365	Pro	Pro	Leu
Val	Thr 370	Ala	Asn	Thr	Val	Leu 375	Ser	Ile	Leu	Ala	Val 380	Asp	Tyr	Pro	Val
Asp 385	Lys	Val	Ser	Cys	Tyr 390	Val	Ser	Asp	Asp	Gly 395	Ala	Ala	Met	Leu	Thr 400
	Asp	Ala	Leu	Ala 405		Thr	Ser	Glu	Phe 410		Arg	Lys	Trp	Val 415	
Phe	Val	Lys	Lys 420		Asn	Ile	Glu	Pro 425		Ala	Pro	Glu	Trp		Phe
Ser	Gln	Lys		Asp	Tyr	Leu	Lys		Lys	Val	His			Phe	Val
Lys	-	435 Arg	Arg	Ala	Met		_	Glu	Tyr	Glu		445 Phe	Lys	Val	Arg
	450	~3	•		• • •	455	-1	~ 3	•	**- 3	460	~1	a 1	~ 1	
465		Gly			470	_			_	475				_	480
Ile	Met	Gln	Asp	Gly 485	Thr	Pro	Trp	Pro	Gly 490	Asn	Asn	Thr	Xaa	Asp 495	His
Pro	Gly	Met	Ile 500	Gln	Val	Phe	Leu	Gly 505	His	Ser	Gly	Gly	Leu 510	Asp	Thr
Glu	Gly	Asn 515	Glu	Leu	Pro	Arg	Leu 520	Val	Tyr	Val	Ser	Arg 525	Glu	Lys	Arg
Pro	Gly 530	Phe	Gln	His	His	Lys 535	Lys	Ala	Gly	Ala	Met 540	Asn	Ala	Leu	Val
Arg 545		Ser	Ala	Val	Leu 550		Asn	Gly	Gln	Tyr 555		Leu	Asn	Leu	Asp 560
	Asp	His	Tyr	Ile 565		Asn	Ser	Lys	Ala 570		Arg	Glu	Ala	Met 575	
Phe	Leu	Met	Asp 580		Asn	Leu	Gly	Arg 585		Val	Суз	Tyr	Val 590		Phe
Pro	Gln	Arg 595	-	Asp	Gly	Ile	Asp 600		Asn	Asp	Arg	Tyr 605		Asn	Arg
Asn		Val	Phe	Phe	Asp	Ile 615		Leu	Arg	Gly			Gly	Ile	Gln
G147	610 Pro	Val	Tirr	Wa 1	Glv		G117	Cva	17a 1	Dhe	620	Ara	Thr	בומ	T.211
625			_		630		_	_		635					640
TYL	GIĀ	Tyr	GIU	645	PLO	116	цуз	GIII	650	цуs	ĢIY	GIY	PILE	655	Ser
Ser	Leu	Cys	Gly 660		Arg	Lys	Lys	Ala 665	Ser	Lys	Ser	Lys	Lys 670	Gly	Ser
Asp	Lys	Lys 675		Ser	Glņ	Lys	His 680		Asp	Ser	Ser	Val 685		Val	Phe
Asn		Glu	Asp	Ile	Glu	Glu 695		Val	Glu	Gly	Ala 700		Phe	qeA	Asp
Glu 705	_	Ser				Ser			Ser	Leu 715	Glu	Lys	Arg	Phe	
		71.			710				7 011			The root	C144	C111	720
		Ala		725					730					735	
		Ser	740					745					750		
		Cys 755					760					765			
Trp	Ile 770	Tyr	Gly	Ser	Val	Thr 775	Glu	Asp	Ile	Leu	Thr 780	Gly	Phe	Lys	Met
His 785	Ala	Arg	Gly	Trp	Arg 790	Ser	Ile	Tyr	Cys	Met 795	Pro	Lys	Arg	Pro	Ala 800
	Lys	Gly	Ser	Ala 805		Ile	Asn	Leu	Ser 810		Arg	Leu	Asn	Gln 815	Val

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Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu Phe Ser Arg His Cys
       820
                 825 830
Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys Phe Leu Glu Arg Phe
   835
            840
                             845
Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr Ser Ile Pro Leu Leu
 850
                   855
Ile Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Gly Lys Phe Ile
      870
                               875
Ile Pro Glu Ile Ser Asn Phe Ala Ser Ile Trp Phe Ile Ser Leu Phe
            885
                            890
Ile Ser Ile Phe Ala Thr Gly Ile Leu Glu Met Arg Trp Ser Gly Val
         900
                         905
Gly Ile Asp Glu Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly
    915 920
                             925
Ile Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu
         935
                                  940
Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu
             950
                              955
Asp Gly Asp Phe Ala Glu Leu Tyr Met Phe Lys Trp Thr Thr Leu Leu
          965 970 975
Ile Pro Pro Thr Thr Ile Leu Ile Ile Asn Leu Val Gly Val Val Ala
                        985
Gly Ile Ser Tyr Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu
          1000 1005
Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile Val His Leu Tyr Pro
 1010 1015
                            1020
Phe Leu Lys Gly Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val
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                               1035
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ctagcagttg atgccacacg tctgg
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    <220>
    <221> CDS
    <222> (184)...(3406)
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- 103 -

- 103 -																	
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								-					-			geete	120
																ggagcg	180
				gcc													228
		Met	Ala	Ala	Asn	Lys	Gly	Met	Val	Ala	Gly	Ser	His	Asn	Arg	Asn	
		1				5					10					15	
				atg													276
	Glu	Phe	Val	Met	Ile	Arg	His	Asp	Gly	Asp	Val	Pro	Gly	Ser	Ala	Lys	
					20					25					30		
			_	agt				_	-	_	_		_		_		324
	Pro	Thr	Lys	Ser	Ala	Asn	Gly	Gin		Cys	Gin	Ile	Cys	_	qeA	Ser	
				35					40					45			
	~÷~	~a+	~++	tas	~~~	200	~~+	~a+	~+ a		~++	~~~	+~~	22+	~~~	tat	372
	-		_	tca Ser	_			_	_		-	_	_			_	3/2
	vaı	GIY	50	Ser	Ala	1111	GIY	55	vaı	File	val	Ата	60	Maii	Gra	Cys	
			30					33					00				
	acc	ttc	cct	gtc	tac	cac	cca	tac	tat	gag	tat	gag	cac	aag	gag	aaa	420
				Val													
		65			-2-	3	70	-1-	-1-		- 4 -	75	5			,	
	aac	caa	tgc	tgc	ccc	cag	tgc	aag	act	aga	tac	aag	aga	cag	aaa	ggt	468
	Asn	Gln	Cys	Суз	Pro	Gln	Cys	Lys	Thr	Arg	Tyr	Lys	Arg	Gln	Lys	Gly	
	80					85					90					95	
	agc	cct	cga	gtt	cat	ggt	gat	gag	gat	gag	gaa	gat	gtt	gat	gac	cta	516
	Ser	Pro	Arg	Val	His	Gly	Asp	Glu	Asp	Glu	Glu	Asp	Val	Asp	Asp	Leu	
					100					105					110		
																_	
	_		-	ttc			-										564
	Asp	Asn	GIU	Phe	Asn	Tyr	гàа	GIN	_	ser	GTA	ràs	GIA		GIU	Trp	
				115					120					125			
		ata	~~~	gga	an t	ast	aat	~at	cta	tat	tra	tet	act	cac	cat	asa	612
				Gly													0.2.2
	GLII	neu	130	Gry	Азр	voh	мла	135	neu	Der	Ser	Ser	140	7-9	112.5	014	
			130					133									
	cca	cat	cat	cgg	att	cca	cac	cta	aca	age	aat	caa	caq	ata	tct	qqa.	660
				Arg			-	_									
		145					150				•	155				=	
	gag	att	cct	gat	gct	tcc	cct	gac	cgt	cat	tct	atc	cgc	agt	cca	aca	708
				Asp													
	160					165					170					175	
	tcg	agc	tat	gtt	gat	cca	agc	gtc	cca	gtt	cct	gtg	agg	att	gtg	gac	756
	Ser	Ser	Tyr	Val	Asp	Pro	Ser	Val	Pro	Val	Pro	Val	Arg	Ile	Val	Asp	
					180					185					190		
				gac													804
	Pro	Ser	Lys	qzA	Leu	Asn	Ser	Tyr		Leu	Asn	Ser	Val		Trp	Lys	
				195					200					205			
																	050
	gaa	aga	gtt	gag	agc	tgg	agg	gtt	aaa	cag	gac	aaa	aat	atg	arg	caa cl=	852
	Glu	Arg	Val	Glu	Ser	Trp	Arg	Val	Lys	Gin	Asp	гàа	Asn	met	Met	GTII	

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210	21	15	220	
	a tat cca gag go s Tyr Pro Glu Al 230	la Arg Gly Gly	·	
= =	a gaa nat atg ca y Glu Xaa Met Gl 245			
	c gtg cca att to e Val Pro Ile Se 260	-		
	t ctc cgt ctt at e Leu Arg Leu Il 5		•	
	t cca gtg cgt ga s Pro Val Arg As 29	sp Ala Tyr Gly		
	g gtc tgg ttt gc u Val Trp Phe Al 310	la Leu Ser Trp		
	t cca atc aac co r Pro Ile Asn Ar 325			•
	t aga gag gga ga p Arg Glu Gly Gl 340	· · · · · · · · · · · · · · · · · · ·		-
	t aca gtg gat co r Thr Val Asp Pr 5		-	
	t ttg tcc att ct l Leu Ser Ile Le 37	eu Ser Val Asp '		
	t gtt tct gat ga r Val Ser Asp As 390	sp Gly Ser Ala i		
	a acc gca gaa tt u Thr Ala Glu Ph 405		•	_
	t att gaa cca ag n Ile Glu Pro Ar 420		-	
_	c ctg aag gac aa r Leu Lys Asp Ly 5		-	- <u>-</u>

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								-								
_	_	gca Ala 450	_	_				_	_							1572
-		gtt Val	-			_				_			-		_	1620
_	_	gga Gly		-												1668
_		cag Gln	_		_			_				_		_		1716
		tta Leu		-		-		_		_	-	_	-		· =	1764
		cat His 530														1812
	_	gtg Val					-					-				1860
		ttc Phe		_	-		-		_	-	_	_	-			1908
atg Met	gat Asp	ccg Pro	gct Ala	cta Leu 580	gga Gly	agg Arg	aaa Lys	act Thr	tgt Cys 585	tat Tyr	gta Val	caa Gln	ttt Phe	cca Pro 590	cag Gln	1956
		gat Asp														2004
		ttt Phe 610														2052
		gtg Val							_	_					~ 1	2100
		cct Pro														2148
aag Lys	agc Ser	tgc Cys	tgt Cys	ggt Gly 660	aga Arg	agg Arg	aag Lys	aaa Lys	aag Lys 665	aac Asn	aag Lys	agt Ser	tat Tyr	atg Met 670	gat Asp	2196
agt Ser	caa Gln	agc Ser	cgt Arg	att Ile	atg Met	aag Lys	aga Arg	aca Thr	gaa Glu	tct Ser	tca Ser	gct Ala	ccc Pro	atc Ile	ttc Phe	2244

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6	675	680)	685	
			gaa ggt tac ga Glu Gly Tyr Gl 70	u Asp Glu Arg	2292
			g gag aaa cgc tt 1 Glu Lys Arg Ph 715		2340
			g aca caa ggt gg Thr Gln Gly Gl 730		2388
			ggaa gct atc car Glu Ala Ile His 745		2436
Cys Gly Tyr C			gga aaa gag at Gly Lys Glu Ile		2484
			act ggg ttt aaa Thr Gly Phe Lys 780	Met His Ala	2532
	Gln Ser Ile		cca cca cga cci Pro Pro Arg Pro 795	-	2580
-:	_	_	cgt ctt aat cag Arg Leu Asn Glr 810		2628
		-	ctt agt aga cat Leu Ser Arg His 825	-	2676
Trp Tyr Gly T			ctt ttg gag agg		2724
	•		tcc att ccg ctt Ser Ile Pro Leu 860	ı Ile Ala Tyr	2772
	Pro Ala Ile	-	acc aat aaa ttt Thr Asn Lys Phe 875		2820
			ttc att ctt ctt Phe Ile Leu Leu 890		2868
			aga tgg agt ggt Arg Trp Ser Gly 905	• • • • • • • • • • • • • • • • • • • •	2916

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-	gat Asp		_				-			_						2964
gcc Ala	cat His	ctc Leu 930	ttc Phe	gca Ala	gtg Val	ttc Phe	cag Gln 935	ggt Gly	ctg Leu	ctg Leu	aaa Lys	gtg Val 940	ttg Leu	gct Ala	gl ^à aaa	3012
att Ile	gat Asp 945	acc Thr	aac Asn	ttc Phe	aca Thr	gtt Val 950	acc Thr	tca Ser	aag Lys	gca Ala	tct Ser 955	gat Asp	gag Glu	gat Asp	ggc Gly	30,60
	ttt Phe															3108
_	acc Thr							_	_		_		_			3156
	tat Tyr			Asn					Ser					Phe		3204
	ctg Leu		Phe	_				Ile					Pro			3252
	ggt Gly 102	Leu	_	_			Asn	_				Ile	_		_	3300
	tcc Ser				-	Ser				_	Leu			-		3348
_	cct Pro				Pro		_		-	Ala	-	-			Cys	3396
	gtc Val		t go	tgat	cgag	g aca	igtga	ictc	ttat	ttga	ag a	ıggct	caat	cc		3446
gtga tgct aaga	iggat gegg tgtg gtttg	gg a yac t yaa t	tttg aaga tttg	cato atca jaagt	et as ac go et tt	igtta jagco :gtta	tgcc tttc tgcc	tet tac	gtto ctto agtt	att cat tat	aget gtag tgtt	tett regeo	cc g ag d	tgcc cago aaat	tgtag ggtgc agegt tatca aaaaaa	3506 3566 3626 3686 3746 3753

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<213> Zea mays

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- 108 -

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			20					25					30		
	Lys	35			_		40	_			_	45	_		
Gly	Val 50	Ser	Ala	Thr	Gly	Asp 55	Val	Phe	Val	Ala	Cys 60	Asn	Glu	Суз	Ala
Phe 65	Pro	Val	Суз	Arg	Pro 70	Cys	Tyr	Glu	Tyr	Glu 75	Arg	Lys	Glu	Gly	Asn 80
Gln	Cys	Суз	Pro	Gln 85	Cys	Lys	Thr	Arg	Tyr 90	Lys	Arg	Gln	Lys	Gly 95	Ser
Pro	Arg	Val	His 100	Gly	Asp	Glu	Asp	Glu 105	Glu	Asp	Val	Asp	Asp 110	Leu	Asp
Asn	Glu	Phe 115	Asn	Tyr	Lys	Gln	Gly 120	Ser	Gly	Lys	Gly	Pro 125	Glu	Trp	Gln
Leu	Gln 130	Gly	Asp	Asp	Ala	Asp 135	Leu	Ser	Ser	Ser	Ala 140	Arg	His	Glu	Pro
His 145	His	Arg	Ile	Pro	Arg 150	Leu	Thr	Ser	Gly	Gln 155	Gln	Ile	Ser	Gly	Glu 160
Ile	Pro	Asp	Ala	Ser 165	Pro	Asp	Arg	His	Ser 170	Ile	Arg	Ser	Pro	Thr 175	Ser
Ser	Tyr	Val	Asp 180	Pro	Ser	Val	Pro	Val 185	Pro	Val	Arg	Ile	Val 190	Asp	Pro
Ser	Lys	Asp 195	Leu	Asn	Ser	Tyr	Gly 200	Leu	Asn	Ser	Val	Asp 205	Trp	Lys	Glu
Arg	Val 210	Glu	Ser	Trp	Arg	Val 215	Lys	Gln	Asp	Lys	Asn 220	Met	Met	Gln	Val
Thr 225	Asn	Lys	Tyr	Pro	Glu 230	Ala	Arg	Gly	Gly	Asp 235	Met	Glu	Gly	Thr	Gly 240
Ser	Asn	Gly	Glu	Xaa 245	Met	Gln	Met	Val	Asp 250	qaA	Ala	Arg	Leu	Pro 255	Leu
	Arg		260					265					270		
	Ile	275					280					285			
	Ser 290					295					300				
305	Cys				310					315					320
-	Trp	-		325		_			330		_			335	
Arg	Tyr	qeA	Arg 340	Glu	Gly	Glu	Pro	Ser 345	Gln	Leu	Ala	Pro	Ile 350	Asp	Val
	Val	355			_		360	-				365			
	Thr 370					375					380				
Ser 385	Cys	Tyr	Val	Ser	Asp 390	Asp	Gly	Ser	Ala	Met 395	Leu	Thr	Phe	Glu	Ser 400
Leu	Ser	Glu	Thr	Ala 405	Glu	Phe	Ala	Arg	Lys 410	Trp	Val	Pro	Phe	Cys 415	Lys
	His		420					425					430		
Ile	Asp	Tyr 435	Leu	Lys	Asp	Lys	Ile 440	Gln	Pro	Ser	Phe	Val 445	Lys	Glu	Arg
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Leu	Val	Ala	Lys	Ala	Gln	Lys	Va1	Pro	Glu	Glu	Gly	Trp	Thr	Met	Ala

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	Ser			885					890					895	
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_	_	_		- .	~1	~ 2	51 .	m-	**- *	- 7	~ 1	~1 · ·	mb	0	70 T -
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(51) International Patent Classification 7: C12N 15/54, 9/10, 5/10, 15/82 (11) International Publication Number: WO 00/09	706
C12N 15/54, 9/10, 5/10, 15/82 A3 (43) International Publication Date: 24 February 2000 (24.0)	2.00)
(21) International Application Number: PCT/US99/18760 (22) International Filing Date: 16 August 1999 (16.08.99) (23) Priority Data: 60/096,822 17 August 1998 (17.08.98) (24) International Filing Date: 16 August 1999 (16.08.99) (25) International Filing Date: 16 August 1999 (16.08.99) (26) Priority Data: 60/096,822 17 August 1998 (17.08.98) (27) Applicant (for all designated States except US): PIONEER HI-BRED INTERNATIONAL, INC. [US/US]; 800 Capital	GB, KP, MK, G, SI, ZA, SZ, MD, DK,
Square, 400 Locust Street, Des Moines, IA 50309 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): DHUGGA, Kanwarpal, S. [US/US]; 8320 Barnham Drive, Johnston, IA 50131 (US). HELENTJARIS, Timothy, G. [US/US]; 2960 N.W. 73rd Lane, Ankeny, IA 50021 (US). BOWEN, Benjamin, A. [GB/US]; 7027 Buckingham Boulevard, Berkeley, CA 94705 (US). WANG, Xun [CN/US]; 12524 Caminito Vista Soledad, San Diego, CA 92130 (US). (74) Agents: BLAIR, Debra, L. et al.; 7100 N.W. 62nd Avenue, Darwin Building, Johnston, IA 50131–1000 (US).	
(54) Title: MAIZE CELLULOSE SYNTHASES AND USES THEREOF (57) Abstract The invention provides isolated cellulose synthase nucleic acids and their encoded proteins. The present invention provides met and compositions relating to altering cellulose synthase concentration and/or composition of plants. The invention further provides met and composition cassettes, host cells, and transgenic plants.	hods

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DK	Denmark	LK	Sri Lanka	SE	Sweden		
EE	Estonia	LR	Liberia	SG	Singapore		

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A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12N15/54 C12N9/10 C12N5/10 C12N15/82 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) C12N IPC 7 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consuted during the international search (name of data base and, where practical, search terms used) STRAND, EPO-Internal, WPI Data, PAJ C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No WO 98 00549 A (WILLIAMSON RICHARD EDWARD 1-15 Х :PENG LIANGCAI (AU); ARIOLI ANTONIO (AU)) 8 January 1998 (1998-01-08) abstract page 4, line 10 - line 14 page 7, line 19 - line 29 page 8, line 16 - line 21 page 11, line 6 - line 12 page 17, line 4 - line 19 page 24, line 15 - line 18 page 28, line 15 - line 21 WO 98 18949 A (CALGENE INC ; PEAR JULIE R 1-15 Α (US); STALKER DAVID M (US); DELMER DEBOR) 7 May 1998 (1998-05-07) cited in the application abstract page 7, line 14 -page 9, line 25 -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. X X Special categories of cited documents : "T" later occurrent published after the international filing date or priority date and not in conflict with the application out cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the international "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taxen alone filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-"O" document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled in the art. document published pnor to the international filing date but later than the priority date claimed *&* document member of the same patent family Date or mailing of the international search report Date of the actual completion of the international search 14. 7. 00 22 June 2000 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5318 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl. Ceder, 0 Fax: (+31-70) 340-3016

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Interraponal Application No
PCT/US 99/18760

	ation) DOCUMENTS C NSIDERED T BE RELEVANT	
ategory *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 1 June 1998 (1998-06-01), XP002125688 HEIDELBERG DE Ac 048947	15
A	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 3 February 1998 (1998-02-03), XP002125689 HEIDELBERG DE AC AF027174	1
x	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 1 June 1998 (1998-06-01), XP002140697 HEIDELBERG DE Ac 048948	15
x	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 20 January 1998 (1998-01-20), XP002140698 HEIDELBERG DE Ac AF030052	1
x	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 1 June 1998 (1998-06-01), XP002140699 HEIDELBERG DE AC 048946	15
A	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" EMBL SEQUENCE DATABASE, 3 February 1998 (1998-02-03), XP002140700 HEIDELBERG DE Ac AF027173	15
X	the whole document -& ARIOLI ET AL.: "Molecular analysis of cellulose biosynthesis in Arabidopsis" SCIENCE, vol. 279, no. 5351, 30 January 1998 (1998-01-30), pages	15
	717-720, XP002124283 abstract; figure 3	1

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Inten. , anal Application No PCT/US 99/18760

		PCT/US 99/18760
	ntion) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WU ET AL.: "Cellulose synthase" EMBL SEQUENCE DATABASE, 1 August 1998 (1998-08-01), XP002140701 HEIDELBERG DE Ac 065338 the whole document	15
A	PEAR J R ET AL: "HIGHER PLANTS CONTAIN HOMOLOGS OF THE BACTERIAL CELA GENES ENCODINGTHE CATALYTIC SUBUNIT OF CELLULOSE SYNTHASE" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, US, NATIONAL ACADEMY OF SCIENCE. WASHINGTON, vol. 93, page 12637-12642 XP002061424 ISSN: 0027-8424	
A	AMOR Y ET AL: "EVIDENCE FOR A CYCLIC DIGUANYLIC ACID-DEPENDENT CELLULOSE SYNTHASE IN PLANTS" PLANT CELL,US,AMERICAN SOCIETY OF PLANT PHYSIOLOGISTS, ROCKVILLE, MD, vol. 3, page 989-995 XP002061420 ISSN: 1040-4651	
١	US 5 723 764 A (SINGLETARY GEORGE WILLIAM ET AL) 3 March 1998 (1998-03-03) abstract; claims	2-12
	WO 00 04166 A (THORPE ET AL.) 27 January 2000 (2000-01-27) abstract; claims	1-15

International application No. PCT/US 99/18760

B x I Observations whir is intains claims were found unsearchabli (Continuation of item 1 if firstish let)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see additional sheet
As a result of the prior review under R. 40.2(e) PCT, no additional fees are to be refunded.
1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
1-15 all partly (inventions $1,2,3,4,5,7$ and 15 searched)
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: Invention 1: 1-15 all partial

An isolated nucleic acid selected from the groups as defined in claim 1 and uses of said nucleic acid, and a protein selected from the groups as defined in claim 15, where the nucleic acid sequence is SEQ ID NO 1 and the protein sequence is SEQ ID NO 2.

2. Claims: Inventions 2-15: Claims 1-15 all partial

Idem as subject 1 but limited to each of the nucleic acid sequences as in SEQ ID NOS 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, and 57 and corresponding protein sequences SEQ ID NOS 6, 10, 14, 18, 22, 26, 30, 34, 42, 46, 50, 54, and 58, where invention 2 is limited to SEQ ID NOS 5 and 6, invention 3 is limited to SEQ ID NOS 9 and 10,, invention 15 is limited to SEQ ID NOS 57 and 58.

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